

Designation: C1205 – 20

Standard Test Method for The Radiochemical Determination of Americium-241 in Soil by Alpha Spectrometry¹

This standard is issued under the fixed designation C1205; the number immediately following the designation indicates the year of original adoption or, in the case of revision, the year of last revision. A number in parentheses indicates the year of last reapproval. A superscript epsilon (ε) indicates an editorial change since the last revision or reapproval.

1. Scope

1.1 This test method covers the determination of ²⁴¹Am in soil by means of chemical separations and alpha spectrometry. It is designed to analyze up to 10 g of soil or other sample matrices that contain up to 30 mg of combined rare earths. This test method allows the determination of ²⁴¹Am concentrations from ambient levels to applicable standards.

1.2 The values stated in SI units are to be regarded as standard. The values given in parentheses after SI units are provided for information only and are not considered standard.

1.3 This standard does not purport to address all of the safety concerns, if any, associated with its use. It is the responsibility of the user of this standard to establish appropriate safety, health, and environmental practices and determine the applicability of regulatory limitations prior to use.

1.4 This international standard was developed in accordance with internationally recognized principles on standardization established in the Decision on Principles for the Development of International Standards, Guides and Recommendations issued by the World Trade Organization Technical Barriers to Trade (TBT) Committee.

https://standards.iteh.ai/catalog/standards/sist/412f0ato

2. Referenced Documents

2.1 ASTM Standards:²

C859 Terminology Relating to Nuclear Materials

C998 Practice for Sampling Surface Soil for Radionuclides C999 Practice for Soil Sample Preparation for the Determination of Radionuclides

- C1163 Practice for Mounting Actinides for Alpha Spectrometry Using Neodymium Fluoride
- D1193 Specification for Reagent Water
- D3084 Practice for Alpha-Particle Spectrometry of Water

D3648 Practices for the Measurement of Radioactivity

3. Terminology

3.1 Definitions:

3.1.1 Except as otherwise defined herein, definitions of terms are as given in Terminology C859.

4. Summary of Test Method

4.1 Americium-241 is determined in prepared soil samples of up to 10 g. The soil is completely dissolved by use of pyrosulfate fusion. After an initial separation on barium sulfate and extraction with an organophosphorous compound, the americium is separated from the other trivalent actinides and the rare earths by oxidation of the americium and precipitation of the interferences. The americium is prepared for alpha spectrometry by coprecipitation with neodymium fluoride and the ²⁴¹Am determined by alpha spectrometry using ²⁴³Am as a yield monitor.

4.2 Typical radiochemical recoveries of this test method as determined by the yield monitor, are between 75 and 90 %. Decontamination factors from other radionuclides that may interfere with the determination of americium in this energy range are 10^4 – 10^5 .

4.3 The reagent blank contains all reagents plus the 243 Am tracer. Five samples and a reagent blank can be completed and ready for alpha spectrometry in approximately 6 h. The full-width at half-maximum (FWHM) detector resolution ranges between 43 and 65 keV.

5. Significance and Use

5.1 This test method provides the speed and high decontamination factors attainable with liquid-liquid extraction of the actinides and eliminates filtration techniques that are more time consuming.

5.2 This test method provides a precise determination of americium in concentrations normally found in environmental samples.

6. Interferences

6.1 Plutonium, if inadequately separated, may interfere with the alpha spectrometric determination of ²⁴¹Am. Thorium-228,

¹ This test method is under the jurisdiction of ASTM Committee C26 on Nuclear Fuel Cycle and is the direct responsibility of Subcommittee C26.05 on Methods of Test.

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² For referenced ASTM standards, visit the ASTM website, www.astm.org, or contact ASTM Customer Service at service@astm.org. For *Annual Book of ASTM Standards* volume information, refer to the standard's Document Summary page on the ASTM website.

identifiable by its progeny, is a serious interference to the final determination of americium by alpha spectrometry if decontamination factors are not sufficiently high. An inadequate separation of ²¹⁰Po may result in an inaccurate determination of the ²⁴³Am yield monitor but this is unlikely when using the neodymium fluoride precipitation method. If high concentrations of these radionuclides are known to be present, a preliminary separation may be required.

7. Apparatus

7.1 Alpha pulse height analysis system as in Practice D3084.

7.1.1 A system consisting of a silicon surface barrier detector capable of 50 keV or better resolution on standards electrodeposited on a flat, mirror finished disk is required. Samples prepared for alpha spectrometry using neodymium fluoride mounting by Practice C1163 should be capable of 60 to 70 keV resolution. The resolution is defined as the width of an alpha energy peak when the counts on either side of the peak are equal to one-half of the counts at the maximum of the peak (FWHM).

7.1.2 The counting efficiency of the system (that is, count/ disintegration) should be greater than 20 % and the instrument background in the region of each energy peak used for analysis should be less than five counts in 60 000 s (1000 min).

7.2 Membrane Filter (such as cellulose nitrate or cellulose acetate), 47 mm diameter, 0.45 µm pore size.

- 7.3 Platinum dish, 250 mL.
- 7.4 Blast burner.
- 7.5 Ring stand.
- 7.6 Glass beaker, 800 mL.
- 7.7 Hot plate with fiberglass mat cover. ards/sist/412/0afe-9 acid with water and dilute to 1 L.
- 7.8 Erlenmeyer flask, 125 mL.
- 7.9 Separatory funnel, 60 mL.
- 7.10 Polycarbonate counting bottle, 70 mL.
- 7.11 Centrifuge.
- 7.12 Hot water bath.
- 7.13 Tetrafluoroethylene (TFE) fluorocarbon beaker.
- 7.14 Conical polycarbonate centrifuge tube, 50 mL.
- 7.15 Round bottom polycarbonate centrifuge tube, 50 mL.
- 7.16 Ice water bath.

8. Reagents and Materials

8.1 *Purity of Reagents*—Reagent grade chemicals shall be used in all tests. Unless otherwise indicated, it is intended that all reagents shall conform to the specifications of the Committee on Analytical Reagents of the American Chemical Society, where such specifications are available. Other grades may be used, provided it is first ascertained that the reagent is of sufficiently high purity to permit its use without lessening the accuracy of the determination.

8.2 *Purity of Water*—Unless otherwise indicated, reference to water shall be understood to mean reagent water as defined in Specification D1193, Type III or better.

8.3 *Americium Tracer*—Purify the ²⁴³Am tracer.³ The ²⁴³Am tracer may be available from NIST or other recognized standards laboratories.

8.4 Potassium Fluoride, anhydrous.

8.5 Potassium Sulfate, anhydrous.

8.6 Sodium Sulfate, anhydrous.

8.7 Ammonium Persulfate (ammonium peroxydisulfate).

8.8 Hydrochloric Acid (sp gr 1.19)—Concentrated hydrochloric acid (HCl).

8.9 *Hydrofluoric Acid (sp gr 1.20)*—Concentrated hydrofluoric acid (HF).

8.10 Sulfuric Acid (sp gr 1.84)—Concentrated sulfuric acid (H_2SO_4) .

8.11 *Sulfuric Acid Solution 0.5* %—Mix 5 mL of concentrated sulfuric acid with water and dilute to 1 L.

8.12 *Nitric Acid (sp gr 1.42)*—Concentrated nitric acid (HNO₃).

8.13 *Potassium Metabisulfite Solution 25 %*—Dissolve 25 g of potassium metabisulfite in water and dilute to 100 mL.

8.14 Sodium Bromate Solution 10 %—Dissolve 10 g of sodium bromate in water and dilute to 100 mL.

8.15 *HDEHP Solution 15* %—Dissolve 150 mL of bis(2-ethylhexyl)phosphoric acid in 850 mL of n-heptane.

8.16 *Barium Chloride Solution 0.5* %—Dissolve 0.5 g of barium chloride in water and dilute to 100 mL.

<u>205-8.17</u> 5M Nitric Acid—Mix 312 mL of concentrated nitric acid with water and dilute to 1 L. Jackson at 2005-200

8.18 *Silver Nitrate Solution 0.5* %—Dissolve 0.5 g of silver nitrate in water and dilute to 100 mL.

8.19 *Lanthanum Carrier (5 mg La/mL)*— Dissolve 1.17 g of lanthanum nitrate in 75 mL of 5M nitric acid and dilute to 100 mL with water.

8.20 *Phosphoric Acid (sp gr 1.83)*—Concentrated phosphoric acid (H_3PO_4) .

8.21 *0.2M Ammonium Persulfate*—Dissolve 2.3 g of ammonium persulfate in water and dilute to 50 mL. Prepare daily.

8.22 *6M Ammonium Fluoride*—Dissolve 22.2 g of ammonium fluoride in water and dilute to 100 mL.

8.23 0.10M Ammonium Persulfate-3N Ammonium Fluoride—Mix 20 mL of 0.2M ammonium persulfate with 20 mL of 6M ammonium fluoride. Prepare daily.

8.24 Hydrogen Peroxide Solution 30 %.

8.25 *Perchloric Acid (sp gr 1.67)*—Concentrated perchloric acid (HClO₄).

³ For a description of this process, see Sill, C. W., *Analytical Chemistry*, 46, 1974, pp. 1426–1431.

8.26 *Neodymium Carrier (10 mg Nd/mL)*—Heat 25 mL of 12M hydrochloric acid and 1.17 g of neodymium oxide on a hot plate until the neodymium oxide is in solution. Cool the solution and dilute to 100 mL with water.

8.27 *Neodymium Carrier (0.5 mg Nd/mL)*—Dilute 5 mL of the 10 mg Nd/mL neodymium carrier solution to 100 mL with water.

9. Sampling

9.1 Collect the sample in accordance with Practice C998.

9.2 Prepare the sample for analysis in accordance with Practice C999.

10. Hazards

10.1 In addition to other precautions, adequate laboratory facilities, such as perchloric acid fume hoods and controlled ventilation, along with safe techniques must be used in this procedure. Extreme care should be exercised in using hydro-fluoric acid and other hot concentrated acids, particularly hot perchloric acid. Use of safety equipment, especially safety glasses and rubber gloves, is recommended.

10.2 Hydrofluoric acid is a highly corrosive and toxic acid that can severely burn skin, eyes, and mucous membranes. Hydrofluoric acid differs from other acids because the fluoride ion readily penetrates the skin, causing destruction of deep tissue layers. Unlike other acids that are rapidly neutralized, hydrofluoric acid reactions with tissue may continue for days if left untreated. Familiarization and compliance with the Safety Data Sheet is essential.

10.3 Perchloric acid is one of the strongest acids known. At room temperature, aqueous solutions up to 72 % do not have significant oxidizing power, and the corrosive properties are very similar to other mineral acids. However, the oxidizing power increases with concentration and temperature. Concentrated perchloric acid (72 %) heated above 150 °C is a strong oxidizer, and reacts violently with organic material, which has resulted in devastating explosions in specific situations. Familiarization and compliance with the Safety Data Sheet is essential.

11. Calibration and Standardization

11.1 If an ²⁴³Am solution traceable to a national standards body is not available for use as a tracer, standardize a freshly prepared sample of ²⁴³Am using the guidance in Practice D3648. These practices may also be used to establish the counting efficiency of the alpha spectrometer which then can be used to calculate the chemical recovery.

12. Procedure

12.1 Weigh 10 g of ~35 mesh soil, to 0.001 g, into a 250 mL platinum dish and add 30 g of anhydrous potassium fluoride, 10 mL of water, 10 mL of concentrated hydrofluoric acid, and 2 mL of concentrated nitric acid. Slurry the contents of the dish together and add an ²⁴³Am tracer at the level of approximately 0.1 Bq. Place the dish on a fiberglass matcovered hot plate and evaporate the contents to dryness. Place the dish on a ring stand and heat with a high temperature blast

burner until the contents have dissolved completely. Swirl the contents gently to dissolve any sample on the sides of the dish. When the sample has dissolved completely, carefully remove the platinum dish from the ring stand and swirl the melt as it cools to deposit the melt evenly along the sides and the bottom of the dish.

12.2 After the contents of the dish have cooled to room temperature, carefully add 40 mL of concentrated sulfuric acid to transpose the fluoride cake. After the initial vigorous reaction has subsided, place the dish on a fiberglass matcovered hot plate and heat until the fluoride cake has been completely transposed. Add 20 g of anhydrous sodium sulfate, place the dish on the ring stand and heat gently until the viscous solution begins to boil. Increase the temperature from the blast burner until a smooth boiling mixture is obtained and continue heating until a clear melt results. Cool the dish to solidify the melt and then place the dish into a cold water bath to facilitate cake removal.

12.3 Bring 500 mL of water and 150 mL of concentrated hydrochloric acid to a boil in a 800 mL beaker and carefully add the contents of the platinum dish to the beaker. Continue boiling until a clear solution results. Add 50 g of anhydrous potassium sulfate and 2 mL of a 25 % potassium metabisulfite solution to the beaker and continue boiling for two minutes.

12.4 To the boiling contents of the beaker, add four 10 mL portions of a 0.5 % barium chloride solution with a 5-min boiling interval between each addition. Stir the contents of the beaker after each addition of barium chloride. Filter the hot solution through a 47 mm membrane filter using either a glass or polycarbonate filtering apparatus. Rinse the beaker and the precipitate with a 0.5 % sulfuric acid solution. Place the filter containing the barium sulfate precipitate into a 125 mL Erlenmeyer flask containing 30 mL of concentrated perchloric acid and heat the contents to fumes of perchloric acid. The use of perchloric acid during the above procedure is used to dissolve the barium sulfate precipitate and the filter and presents little or no safety hazard. The addition of nitric acid during this step is not necessary. Cool the contents of the flask to room temperature.

12.5 Add 1 mL of a 10 % sodium bromate solution to the flask containing the perchloric acid and transfer the contents of the flask to a 60 mL separatory funnel containing 10 mL of 15 % HDEHP in n-heptane and shake for five minutes. After complete phase separation, draw off the lower aqueous layer and discard. Wash the organic extract twice with 5 mL portions of concentrated perchloric acid for two minutes. Discard the wash solutions.

12.6 Strip the trivalent actinides and lanthanides twice for four minutes each with 10 mL portions of 5M nitric acid containing one mL of 10 % sodium bromate solution. Transfer the solution containing the trivalent actinides and lanthanides to another separatory funnel containing 10 mL of 15 % HDEHP in n-heptane and extract for two minutes to remove any plutonium, thorium, or tetravalent cerium that may have stripped during the back extraction of the trivalent actinides. After complete phase separation, draw off the lower aqueous