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Designation: D6139 - 18 D6139 - 22

### Standard Test Method for Determining the Aerobic Aquatic Biodegradation of Lubricants or Their Components Using the Gledhill Shake Flask<sup>1</sup>

This standard is issued under the fixed designation D6139; the number immediately following the designation indicates the year of original adoption or, in the case of revision, the year of last revision. A number in parentheses indicates the year of last reapproval. A superscript epsilon ( $\varepsilon$ ) indicates an editorial change since the last revision or reapproval.

#### 1. Scope\*

1.1 This test method covers the determination of the degree of aerobic aquatic biodegradation of fully formulated lubricants or their components on exposure to an inoculum under controlled laboratory conditions. This test method is an ultimate biodegradation test that measures carbon dioxide  $(CO_2)$  evolution.

1.2 This test method is intended to specifically address the difficulties associated with testing water insoluble materials and complex mixtures such as are found in many lubricants.

1.3 This test method is designed to be applicable to all non-volatile lubricants or lubricant components that are not toxic and not inhibitory at the test concentration to the organisms present in the inoculum.

1.4 The values stated in SI units are to be regarded as standard. No other units of measurement are included in this standard.

1.5 This standard does not purport to address all of the safety concerns, if any, associated with its use. It is the responsibility of the user of this standard to establish appropriate safety, health, and environmental practices and determine the applicability of regulatory limitations prior to use. Specific hazards are discussed in Section 10.

1.6 This international standard was developed in accordance with internationally recognized principles on standardization established in the Decision on Principles for the Development of International Standards, Guides and Recommendations issued by the World Trade Organization Technical Barriers to Trade (TBT) Committee.

#### 2. Referenced Documents

2.1 ASTM Standards:<sup>2</sup>

D1193 Specification for Reagent Water

D1293 Test Methods for pH of Water

D4447 Guide for Disposal of Laboratory Chemicals and Samples

D5291 Test Methods for Instrumental Determination of Carbon, Hydrogen, and Nitrogen in Petroleum Products and Lubricants D5864 Test Method for Determining Aerobic Aquatic Biodegradation of Lubricants or Their Components

E943 Terminology Relating to Biological Effects and Environmental Fate

\*A Summary of Changes section appears at the end of this standard

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<sup>&</sup>lt;sup>1</sup> This test method is under the jurisdiction of ASTM Committee D02 on Petroleum Products, Liquid Fuels, and Lubricants and is the direct responsibility of Subcommittee D02.12 on Environmental Standards for Lubricants.

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<sup>&</sup>lt;sup>2</sup> For referenced ASTM standards, visit the ASTM website, www.astm.org, or contact ASTM Customer Service at service@astm.org. For Annual Book of ASTM Standards volume information, refer to the standard's Document Summary page on the ASTM website.

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2.2 ISO Standard:<sup>3</sup>
4259:1992(E) Petroleum Products—Determination and application of precision data in relation to methods of test
2.3 APHA Standards:<sup>4</sup>
2540B Total Solids Dried at 103–105°C
9215 Heterotrophic Plate Count

#### 3. Terminology

3.1 Definitions:

# iTeh Standards (https://standards.iteh.ai) Document Preview

ASTM D6139-22

https://standards.iteh.ai/catalog/standards/sist/7cacfca8-4729-4c8a-be02-f473a5cae6e2/astm-d6139-22

<sup>&</sup>lt;sup>3</sup> Available from American National Standards Institute (ANSI), 25 W. 43rd St., 4th Floor, New York, NY 10036, http://www.ansi.org.

<sup>&</sup>lt;sup>4</sup> Methods from *Standard Methods for the Examination of Water and Wastewater*, latest edition. Available from the American Public Health Association (APHA), 800 I Street, NW, Washington, DC 20001.

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3.1.1 Definitions of terms applicable to this test method that are not described herein appear in the ASTM Online Dictionary of Engineering Science and Technology<sup>5</sup> or Terminology E943.

<u>3.1.2 activated sludge</u>, n—the precipitated solid matter, consisting mainly of bacteria and other aquatic microorganisms, that is produced at a domestic wastewater treatment plant and is used primarily in secondary sewage treatment to microbially oxidize dissolved organic matter in the effluent.

3.1.3 *aerobic*, *adj*—(1) taking place in the presence of oxygen; (2) living or active in the presence of oxygen.

3.1.4 *biodegradation*, *n*—the process of chemical breakdown or transformation of a material caused by organisms or their enzymes.

3.1.4.1 Discussion—

Biodegradation is only one mechanism by which materials are transformed in the environment.

<u>3.1.5 *biomass*</u>, *n*—biological material including any material other than fossil fuels which is or was a living organism or component or product of a living organism.

3.1.5.1 Discussion—

In biology and environmental science, biomass is typically expressed as density of biological material per unit sample volume, area, or mass (g biomass / g (or / mL or / cm<sup>2</sup>) sample); when used for products derived from organisms biomass is typically expressed in terms of mass (kg, MT, etc.) or volume (L, m<sup>3</sup>, bbl, etc.).

3.1.5.2 Discussion—

Products of living organisms include those materials produced directly by living organisms as metabolites (for example, ethanol, various carbohydrates and fatty acids), materials manufactured by processing living organisms (for example: pellets manufactured by shredding and pelletizing plant material) and materials produced by processing living organisms, their components or metabolites (for example, transesterified oil; also called biodiesel).

3.1.6 blank, n-in biodegradability testing, a test system containing all system components with the exception of the test material.

3.1.7 inoculum, n-spores, bacteria, single celled organisms, or other live materials, that are introduced into a test medium.

<u>3.1.8 lag phase, n—the period of diminished physiological activity and cell division following the addition of microorganisms to a new culture medium.</u>

3.1.9 log phase, n-the period of growth of microorganisms during which cells divide at a positive constant rate.

3.1.10 *mixed liquor, n*—in sewage treatment, the contents of an aeration tank including the activated sludge mixed with primary effluent or the raw wastewater and return sludge.

3.1.11 *pre-adaptation*, *n*—the pre-incubation of an inoculum in the presence of the test material under conditions similar to the test conditions.

3.1.11.1 Discussion—

The aim of pre-adaptation is to improve the precision of the test method by decreasing variability in the rate of biodegradation produced by the inoculum. Pre-adaptation may mimic the natural processes which cause changes in the microbial population of the inoculum leading to a more rapid rate of biodegradation of the test material but not to a change in the final degree of biodegradation.

3.1.12 pre-condition, n-the pre-incubation of an inoculum under the conditions of the test in the absence of the test material.

3.1.13 supernatant, n-the liquid above settled solids.

3.1.14 suspended solids (of activated sludge or other inoculum samples), n—solids present in activated sludge or inoculum samples that are not removed by settling under specified conditions.

3.1.15 theoretical CO<sub>2</sub>, n-the amount of CO<sub>2</sub> which could in theory be produced from the complete oxidation of all of the carbon

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#### in a material.

3.1.16 *ultimate biodegradation, n*—degradation achieved when a material is totally utilized by microorganisms resulting in the production of  $CO_2$  (and possibly methane in the case of anaerobic biodegradation), water, inorganic compounds, and new microbial cellular constituents (biomass or secretions, or both).

3.2 Definitions of terms applicable to this test method that are not described herein appear in the ASTM Online Dictionary of Engineering Science and Technology<sup>5</sup> or Terminology E943.

3.3 activated sludge, n—the precipitated solid matter, consisting mainly of bacteria and other aquatic microorganisms, that is produced at a domestic wastewater treatment plant; activated sludge is used primarily in secondary sewage treatment to microbially oxidize dissolved organic matter in the effluent.

3.4 aerobic, adj-(1) taking place in the presence of oxygen; (2) living or active in the presence of oxygen.

3.5 *biodegradation, n*—the process of chemical breakdown or transformation of a material caused by organisms or their enzymes. 3.5.1 *Discussion*—

Biodegradation is only one mechanism by which materials are transformed in the environment.

3.6 *biomass, n*—biological material including any material other than fossil fuels which is or was a living organism or component or product of a living organism.

3.6.1 Discussion—

In biology and environmental science, biomass is typically expressed as density of biological material per unit sample volume, area, or mass (g biomass / g (or / mL or / cm<sup>2</sup>) sample); when used for products derived from organisms biomass is typically expressed in terms of mass (kg, MT, etc.) or volume (L, m<sup>3</sup>, bbl, etc.).

3.6.2 Discussion-

Products of living organisms include those materials produced directly by living organisms as metabolites (for example, ethanol, various carbohydrates and fatty acids), materials manufactured by processing living organisms (for example: pellets manufactured by shredding and pelletizing plant material) and materials produced by processing living organisms, their components or metabolites (for example, transesterified oil; also called biodiesel).

3.7 blank, n-in biodegradability testing, a test system containing all system components with the exception of the test material.

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3.8 inoculum, n-spores, bacteria, single celled organisms, or other live materials, that are introduced into a test medium.

3.9 lag phase, n—the period of diminished physiological activity and cell division following the addition of microorganisms to a new culture medium.

3.10 log phase, n-the period of growth of microorganisms during which cells divide at a positive constant rate.

3.11 *mixed liquor*, *n*—in sewage treatment, the contents of an aeration tank including the activated sludge mixed with primary effluent or the raw wastewater and return sludge.

3.12 *pre-adaptation*, *n*—the pre-incubation of an inoculum in the presence of the test material under conditions similar to the test conditions.

3.12.1 Discussion—

The aim of pre-adaptation is to improve the precision of the test method by decreasing variability in the rate of biodegradation produced by the inoculum. Pre-adaptation may mimic the natural processes which cause changes in the microbial population of the inoculum leading to a more rapid rate of biodegradation of the test material but not to a change in the final degree of biodegradation.

3.13 pre-condition, n—the pre-incubation of an inoculum under the conditions of the test in the absence of the test material.

<sup>&</sup>lt;sup>5</sup> ASTM Online Dictionary of Engineering Science and Technology (Stock#DEFONLINE) is available on the ASTM website, www.astm.org, or contact ASTM Customer Service at service@astm.org.

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3.14 supernatant, n-the liquid above settled solids.

3.15 suspended solids (of activated sludge or other inoculum samples), n—solids present in activated sludge or inoculum samples that are not removed by settling under specified conditions.

3.16 *theoretical CO*<sub>2</sub>, n—the amount of CO<sub>2</sub> which could in theory be produced from the complete oxidation of all of the carbon in a material.

3.17 ultimate biodegradation, n—degradation achieved when a material is totally utilized by microorganisms resulting in the production of CO<sub>2</sub> (and possibly methane in the case of anaerobic biodegradation), water, inorganic compounds, and new microbial cellular constituents (biomass and secretions).

#### 4. Summary of Test Method

4.1 Biodegradation of a lubricant or the component(s) of a lubricant is estimated by collecting and measuring the  $CO_2$  produced when the lubricant or component is exposed to microorganisms under controlled aerobic aquatic conditions. This value is then compared to the theoretical amount of  $CO_2$  which could be generated if all of the carbon in the test material were converted to  $CO_2$ . Carbon dioxide is a product of aerobic microbial metabolism of carbon-containing materials and so is a direct measure of the test material's ultimate biodegradation. The evolved  $CO_2$  is trapped in a Ba(OH)<sub>2</sub> or other alkaline solution and the amount of  $CO_2$  absorbed is determined by titrating the remaining hydroxide in solution.

4.2 The carbon content of the test material is determined by Test Methods D5291 or another appropriate method and the theoretical  $CO_2$  is calculated from that measurement. It is necessary to directly measure the carbon content of the test material instead of calculating this number, because of the complexity of the mixture of compounds present in lubricants.

4.3 Biodegradability is expressed as a percentage of theoretical  $CO_2$  production.

#### 5. Significance and Use

5.1 Results from this  $CO_2$  evolution test method suggest, within the confines of a controlled laboratory setting, the degree of ultimate aerobic aquatic biodegradability of a lubricant or components of a lubricant. Test materials which achieve a high degree of biodegradation in this test method may be assumed to easily biodegrade in many aerobic aquatic environments. (See also Test Method D5864.)

5.2 Because of the stringency of this test method, a low yield of  $CO_2$  does not necessarily mean that the test material is not biodegradable under environmental conditions, but indicates that further testing needs to be carried out in order to establish biodegradability.

5.3 Information on the toxicity of the test material to the inoculum may be useful in the interpretation of low biodegradation results.

5.4 Activated sewage-sludge from a sewage treatment plant that principally treats domestic waste may be used as an aerobic inoculum. An inoculum derived from soil or natural surface waters, or any combination of the three sources, may also be used in this test method.

NOTE 1—Allowance for various and multiple inoculum sources provides access to a greater diversity of biochemical competency and potentially represents more accurately the capacity for biodegradation.

5.5 A reference or control material known to biodegrade under the conditions of this test method is necessary in order to verify the activity of the inoculum. The test method must be regarded as invalid and should be repeated using a fresh inoculum if the reference does not demonstrate biodegradation to the extent of >60 % of the theoretical  $CO_2$  within 28 days.

5.6 The water solubility or dispersibility of the lubricant or components may influence the results obtained and hence the procedure may be limited to comparing lubricants or components with similar solubilities.



FIG. 1 NaOH Scrubber – Flask Trap Assembly for Providing CO<sub>2</sub>-Free Air

5.7 The ratio of carbon incorporated into cellular material to carbon metabolized to  $CO_2$  will vary depending on the organic substrate, on the particular microorganisms carrying out the conversion, and on the environmental conditions under which the conversion takes place. In principle, this variability complicates the interpretation of the results from this test method.

5.8 The behavior of complex mixtures may not always be consistent with the individual properties of the components. The biodegradability of the components may be suggestive of whether a mixture containing these components (that is, a fully formulated lubricant) is biodegradable but such information should be used judiciously.

#### 6. Apparatus

## 6.1 Carbon Dioxide Scrubbing Apparatus (see Fig. 1): Standards

6.1.1 The following are required to produce a stream of  $CO_2$ -free air for aeration and for sparging aqueous solutions and mixtures (for example, test medium, sewage inoculum):

6.1.1.1 *Erlenmeyer flask*, one 1 L with side arm containing 500 mL of 10 *M* sodium hydroxide (NaOH), and fitted with a rubber stopper and an inlet tube that extends below the level of the NaOH solution or an equivalent apparatus or system.

6.1.1.2 *Erlenmeyer flask*, one 1 L with side arm containing 500 mL of distilled water and fitted with a stopper and inlet tube, or an equivalent apparatus or system. alog/standards/sist/7cacfca8-4729-4c8a-be02-473a5cae6e2/astm-d6139-22

6.1.1.3 It is optional to add an empty 1 L Erlenmeyer flask in series with the flasks to prevent liquid carryover.

6.1.1.4 It is optional to add a 1 L Erlenmeyer flask containing 500 mL of 0.1 *M* barium hydroxide  $[Ba(OH)_2]$  solution to monitor for possible breakthrough CO<sub>2</sub>.

6.1.2 Connect the flasks in series as shown in Fig. 1, using vinyl or other suitable non-gas-permeable tubing, to a pressurized air system and purge air through the scrubbing solution.

6.1.3 The CO<sub>2</sub> scrubbing apparatus upstream of the Erlenmeyer flask containing the  $Ba(OH)_2$  may be substituted with an alternative system which effectively and consistently produces CO<sub>2</sub>-free air (that is, containing <1 ppm CO<sub>2</sub>).

6.2 *Incubation/Biodegradation Apparatus—Gledhill-type Shake Flask Units*<sup>6</sup> (see Fig. 2)—Each test material, reference, or blank control requires the following:

6.2.1 *Erlenmeyer Flasks,* 2L—2L Erlenmeyer flasks are used to hold the 1 L of total final aqueous volume but larger volume Erlenmeyer flasks (as large as 3 L to 4 L) may be used if 2 L to 3 L final aqueous volumes are required. The amounts described here are for 1 L final aqueous volumes carried out in 2 L Erlenmeyer flasks; scale procedure accordingly if larger final aqueous volumes and larger Erlenmeyer flasks are necessary.

<sup>&</sup>lt;sup>6</sup> Gledhill, W. E., "Screening Test for Assessment of Ultimate Biodegradability: Linear Alkyl Benzene Sulfonate," *Applied Microbiology* Vol 30, 1975, pp. 992–929. Also see description of Gledhill shake flask unit in EPA Chemical Fate Testing Guidelines for Aerobic Aquatic Biodegradation, EPA Publication 560/6-82-003, No. CG-2000 (August 1982); Federal Register, September 27, 1985, p. 39277, Section 796.3100; 40 CFR 796.3100, 1994.



6.2.2 *Stoppers*—Each stopper is fitted with a conical alkaline trap, an outlet and an inlet vent tube (see Fig. 2). Ensure that the stopper fits tightly in the Erlenmeyer flask to prevent any leaks.

6.2.3 *Conical Alkaline Trap Tube or Unit*—Glass, 40 mL conical tube (borosilicate glass, No. 8120 centrifuge tube or equivalent) welded to a glass support rod, or an equivalent apparatus, will be used to hold the  $Ba(OH)_2$  solution for trapping the evolved  $CO_2$  from aerobic biodegradation. The opening in the alkaline trap tube is large enough to permit  $CO_2$  diffusion into the barium hydroxide solution. The support rod of the conical trap shall fit tightly in the stopper.

6.2.4 *Inlet and Outlet Vent Tubes*—The inlet vent tube attached to the stopper extends down into the flask so that it will be immersed below the surface of the aqueous medium and will be used for sparging. The outlet vent tube will be situated significantly above the level of the aqueous medium and will be used for venting. The two vent tubes shall fit tightly in the stopper.

6.2.5 Flexible tubing which is non-permeable to  $CO_2$  will be used to connect the tops of inlet and outlet vent tubes to form a closed system.

6.2.6 Agitators—Incubator-shaker table unit or equivalent, or stirrers may be used to agitate the aqueous mixture in the Erlenmeyer flasks.

6.3 Analytical Balance, to weigh out test material or reference material to be added to the test flask (capable of weighing to appropriate precision and accuracy, for example,  $\pm 0.0001$  g).

6.4 Titration Apparatus for Measuring the Production of  $CO_2$ :

6.4.1 Appropriate graduated burette filled with standard HCl solution.

6.4.2 Alternatively, an automatic titration apparatus in which the burette dispenser is filled with standard HCl solution. Automatic titrations are carried out to a potentiometric end point of pH 8.3 (that is, phenolphthalein end point equivalent)

6.5 Glass Wool, for filtering the inoculum.

#### 7. Reagents and Materials

7.1 *Purity of Reagents*—Reagent grade chemicals shall be used in all tests. Unless otherwise indicated, it is intended that all reagents conform to the specifications of the Committee on Analytical Reagents of the American Chemical Society where such specifications are available.<sup>7</sup> Other grades may be used, provided it is first ascertained that the reagent is of sufficiently high purity to permit its use without lessening the accuracy of the determination.

7.2 *Purity of Water*—Unless otherwise indicated, references to water shall be understood to mean reagent water as defined by Type II of Specification D1193.

<sup>&</sup>lt;sup>7</sup> Reagent Chemicals, American Chemical Society Specifications, ACS Reagent Chemicals, Specifications and Procedures for Reagents and Standard-Grade Reference <u>Materials</u>, American Chemical Society, Washington, D.C.DC. For suggestions on the testing of reagents not listed by the American Chemical Society, see Analar Standards for Laboratory Chemicals, BDH Ltd., Poole, <del>Dorset, U.K.,</del> <u>Dorset, U.K.,</u> and the United States Pharmacopeia and National Formulary, U.S. Pharmacopeial Convention, Inc. (USPC), Rockville, MD.