



Designation: D8024 – 23

Standard Test Method for Determination of (Tri-n-butyl)-n-tetradecylphosphonium chloride (TTPC) in Water by Multiple Reaction Monitoring Liquid Chromatography/Mass Spectrometry (LC/MS/MS)¹

This standard is issued under the fixed designation D8024; the number immediately following the designation indicates the year of original adoption or, in the case of revision, the year of last revision. A number in parentheses indicates the year of last reapproval. A superscript epsilon (ϵ) indicates an editorial change since the last revision or reapproval.

1. Scope

1.1 This test method covers the determination of (Tri-n-butyl)-n-tetradecylphosphonium chloride (TTPC) in water by dilution with acetone, filtration and analysis by liquid chromatography/tandem mass spectrometry. This test method is not amenable for the analysis of isomeric mixtures of Tributyl-tetradecylphosphonium chloride. TTPC is a biocide that strongly adsorbs to soils.² The water samples are prepared in a solution of 75 % acetone and 25 % water because TTPC has an affinity for surfaces and particles. The reporting range for this method is from 100 ng/L to 4000 ng/L. This analyte is qualitatively and quantitatively determined by this method. This test method adheres to multiple reaction monitoring (MRM) mass spectrometry.

1.2 A full collaborative study to meet the requirements of Practice D2777 has not been completed. This test method contains single-operator precision and bias based on single-operator data. Publication of standards that have not been fully validated is done to make the current technology accessible to users of standards, and to solicit additional input from the user community.

1.3 The Method Detection Limit³ (MDL) and Reporting Range⁴ for the target analyte are listed in Table 1.

1.3.1 The reporting limit in this test method is the minimum value below which data are documented as non-detects. Analyte detections between the method detection limit and the reporting limit are estimated concentrations and are not reported following this test method. The reporting limit is calculated from the concentration of the Level 1 calibration standard as shown in Table 4 for TTPC after taking into account a 2.5 mL water sample volume and a final diluted sample volume of 10 mL (75 % acetone/25 % water). The final solution volume is 10 mL because a 7.5 mL volume of acetone is added to each 2.5 mL water sample which is shaken and filtered.

1.4 *Units*—The values stated in SI units are to be regarded as standard. No other units of measurement are included in this standard.

1.5 *This standard does not purport to address all of the safety concerns, if any, associated with its use. It is the responsibility of the user of this standard to establish appropriate safety, health, and environmental practices and determine the applicability of regulatory limitations prior to use.*

1.6 *This international standard was developed in accordance with internationally recognized principles on standardization established in the Decision on Principles for the Development of International Standards, Guides and Recommendations issued by the World Trade Organization Technical Barriers to Trade (TBT) Committee.*

¹ This test method is under the jurisdiction of ASTM Committee D19 on Water and is the direct responsibility of Subcommittee D19.06 on Methods for Analysis for Organic Substances in Water.

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² More information on TTPC can be found at <http://www.buruenergy.com/wp-content/uploads/BE-Environmental-Properties-of-Proposed-Biocide-BE-91.pdf> (2014) and http://iaspub.epa.gov/sor_internet/registry/substreg/searchandretrieve/advancedsearch/externalSearch.do?p_type=CASNO&p=81741-28-8 (2014).

³ The MDL is determined following the Code of Federal Regulations, 40 CFR Part 136, Appendix B, as a guide. A detailed process determining the MDL is explained in the reference and is beyond the scope of this test method to be explained here.

⁴ Reporting range concentration is calculated from Table 4 concentrations assuming a 50 μ L injection of the Level 1 calibration standard for TTPC, and the highest level calibration standard with a 10 mL final diluted sample volume starting with a 2.5 mL water sample. Volume variations will change the reporting limit and ranges.

2. Referenced Documents

2.1 ASTM Standards:⁵

D1129 Terminology Relating to Water

D1193 Specification for Reagent Water

D2777 Practice for Determination of Precision and Bias of Applicable Test Methods of Committee D19 on Water

D3856 Guide for Management Systems in Laboratories Engaged in Analysis of Water

⁵ For referenced ASTM standards, visit the ASTM website, www.astm.org, or contact ASTM Customer Service at service@astm.org. For *Annual Book of ASTM Standards* volume information, refer to the standard's Document Summary page on the ASTM website.

TABLE 1 Method Detection Limit and Reporting Range

Analyte	MDL (ng/L)	Reporting Range (ng/L)
TTPC	13	100–4000

D4841 Practice for Estimation of Holding Time for Water Samples Containing Organic and Inorganic Constituents

D5847 Practice for Writing Quality Control Specifications for Standard Test Methods for Water Analysis

E2554 Practice for Estimating and Monitoring the Uncertainty of Test Results of a Test Method Using Control Chart Techniques

2.2 Other Standards:

40 CFR Part 136, Appendix B Definition and Procedure for the Determination of the Method Detection Limit⁶

EPA Publication SW-846 Test Methods for Evaluating Solid Waste, Physical/Chemical Methods⁷

3. Terminology

3.1 Definitions:

3.1.1 For definitions of terms used in this standard, refer to Terminology **D1129**.

3.2 Definitions of Terms Specific to This Standard:

3.2.1 *reporting limit, RL, n*—the minimum concentration below which data are documented as non-detects.

3.2.2 *reporting limit check sample, RLCS, n*—a sample used to ensure that if the analyte was present at the reporting limit, it would be confidently identified.

3.2.3 *batch QC, n*—all the quality control samples and standards included in an analytical procedure.

3.2.4 *independent reference material, IRM, n*—a material of known purity and concentration obtained either from the National Institute of Standards and Technology (NIST) or other reputable supplier.

3.2.4.1 *Discussion*—The IRM must be obtained from a different lot of material than is used for calibration.

3.3 Acronyms:

3.3.1 *CCC, n*—Continuing Calibration Check

3.3.2 *CRW, n*—Chicago River Water

3.3.3 *IC, n*—Initial Calibration

3.3.4 *LC, n*—Liquid Chromatography

3.3.5 *LCS/LCSD, n*—Laboratory Control Sample/
Laboratory Control Sample Duplicate

3.3.6 *MDL, n*—Method Detection Limit

3.3.7 *MeOH, n*—Methanol

3.3.8 *mM, n*—millimolar, 1×10^{-3} moles/L

3.3.9 *MRM, n*—Multiple Reaction Monitoring

3.3.10 *MS/MSD, n*—Matrix Spike/Matrix Spike Duplicate

⁶ Available from U.S. Government Printing Office, Superintendent of Documents, 732 N. Capitol St., NW, Washington, DC 20401-0001, <http://www.access.gpo.gov>.

⁷ Available from United States Environmental Protection Agency (EPA), William Jefferson Clinton Bldg., 1200 Pennsylvania Ave., NW, Washington, DC 20460, <http://www.epa.gov>.

3.3.11 *NA, adj*—Not Available

3.3.12 *ND, n*—non-detect

3.3.13 *P&A, n*—Precision and Accuracy

3.3.14 *ppt, n*—parts-per-trillion

3.3.15 *QA, adj*—Quality Assurance

3.3.16 *QC, adj*—Quality Control

3.3.17 *RL, n*—Reporting Limit

3.3.18 *RLCS, n*—Reporting Limit Check Sample

3.3.19 *RSD, n*—Relative Standard Deviation

3.3.20 *RT, n*—Retention Time

3.3.21 *SDS, n*—Safety Data Sheets

3.3.22 *SRM, n*—Single Reaction Monitoring

3.3.23 *SS, n*—Surrogate Standard

3.3.24 *TC, n*—Target Compound

3.3.25 *TTPBr, n*—(Tri-*n*-butyl)-*n*-tetradecylphosphonium bromide

3.3.26 *TTPC, n*—(Tri-*n*-butyl)-*n*-tetradecylphosphonium chloride

3.3.27 *VOA, n*—Volatile Organic Analysis

4. Summary of Test Method

4.1 The operating conditions presented in this test method have been successfully used in the determination of TTPC in water; however, this test method is intended to be performance based and alternative operating conditions can be used to perform this method provided data quality objectives are attained.

4.2 For TTPC analysis, samples are shipped to the lab on ice and analyzed within 14 days of collection. A sample (2.5 mL) is transferred to a VOA vial, a TTPC spike solution is added to Laboratory Control and Matrix Spike samples before the addition of acetone. An isotopically labeled TTPC surrogate would be added at this point. An isotopically labeled TTPBr-D29 surrogate is now available and should be incorporated into this method by the user if requested by the customer.⁸ 7.5 mL of acetone is then added and the solution is mixed by hand or vortex for 1 min. The samples are then filtered through a Nylon membrane syringe driven filter unit⁹ and then analyzed by LC/MS/MS. All concentrations reported only to the reporting limit.

⁸ A surrogate, TTPBr-D29, was custom synthesized by Cambridge Isotope Laboratories Inc., Andover, MA 01810 and was found to be acceptable. Any commercial source of TTPBr-D29 surrogate may be used, SRM transition 428.6 > 372.5 was used. If you are aware of an alternative source that meets the performance of the standard, please provide this information to ASTM International Headquarters. Your comments will receive careful consideration at a meeting of the responsible technical committee,¹ which you may attend.

⁹ A Whatman Puradisc (a trademark of Whatman International Limited of Maidstone, Kent) 25 NYL Disposable Filter unit (Diameter 25 mm, 0.2 μm Nylon membrane syringe driven filter unit has been found suitable for use for this method, any filter unit may be used that meets the performance of this method may be used. If you are aware of an alternative filter that meets the performance of the standard, please provide this information to ASTM International Headquarters. Your comments will receive careful consideration at a meeting of the responsible technical committee,¹ which you may attend.

TABLE 2 Gradient Conditions for Liquid Chromatography

Time (min)	Flow (mL/min)	Percent 95 % Water: 5 % Acetonitrile	Percent Acetonitrile	Percent 400 mM Ammonium Acetate (95 % Water: 5 % Acetonitrile)
0	0.3	95	0	5
1	0.3	95	0	5
4	0.4	0	95	5
11	0.4	0	95	5
12	0.4	95	0	5
15	0.4	95	0	5

TABLE 3 Retention Time, SRM Ions, and Analyte-Specific Mass Spectrometer Parameters

Chemical	Primary/Confirmatory	SRM Transition	Cone (V)	Collision (eV)	Retention Time (minutes)	Primary/Confirmatory SRM Area Ratio
TTPC	Primary (Quantitation)	399.5→229.3	40	45	8.1	NA
	First Confirmatory	399.5→75.9	40	46		0.92
	Second Confirmatory	399.5→343.5	40	10		3.02

TABLE 4 Concentrations of Calibration Standards (ng/L)

Concentrations (ng/L)	LV1	LV2	LV3	LV4	LV5	LV6	LV7	LV8
TTPC	25	50	100	200	400	600	800	1000

4.3 TTPC is identified by comparing the single reaction monitoring (SRM) transition and its confirmatory SRM transitions if correlated to the known standard SRM transition (Table 3) and quantitated utilizing an external calibration. The final report issued for each sample lists the concentration of TTPC, if detected, or RL, if not detected, in ng/L and surrogate recovery, if available.

5. Significance and Use

5.1 TTPC may be used in various industrial and commercial products for use as a biocide. Products containing TTPC have been approved for controlling algal, bacterial, and fungal slimes in industrial water systems.² TTPC should not be persistent in water but may be deposited in sediments at concentrations of concern. Hence, there is a need for quick, easy and robust method to determine TTPC concentration at trace levels in water matrices for understanding the sources and concentration levels in affected areas.

5.2 This method has been used to determine TTPC in reagent water and a river water (Table 8).

NOTE 1—This test method has been used to characterize TTPC in real world water samples with success and similar recoveries as shown in Table 8.

6. Interferences

6.1 All glassware is washed in hot water with detergent and rinsed in hot water followed by distilled water. The glassware is then dried and heated in an oven at 250 °C for 15 min to 30 min. All glassware is subsequently rinsed or sonicated, or both, with acetone, n-propanol, or acetonitrile, or combinations thereof.

6.2 TTPC should not be a common contaminant found in a laboratory, unless involved in the analysis or matrices that contain TTPC. TTPC has been found to continue to adhere to glassware and syringes after routine glassware washing. Rinsing glassware with acetone, n-propanol, or acetonitrile, or combinations thereof, or even sonication, may be required to remove TTPC. All of the materials and supplies are routinely demonstrated to be free from interferences of TTPC by analyzing laboratory blanks under the same conditions as the samples. If found, measures should be taken to remove the contamination or data should be qualified, background subtraction of blank contamination is not allowed.

6.3 All reagents and solvents should be pesticide residue purity or higher to minimize interference problems.

TABLE 5 Preparation of Calibration Standards

Solution	LV1	LV2	LV3	LV4	LV5	LV6	LV7	LV8
A ^A	25 µL	50 µL	100 µL	200 µL	400 µL	600 µL	800 µL	1000 µL
B ^B	975 µL	950 µL	900 µL	800 µL	600 µL	400 µL	200 µL	0 µL

^A Solution A: Level 8 stock solution prepared according to Section 12 and at Table 4 concentrations.

^B Solution B: 75 % Acetone, 25 % Water.

TABLE 6 QC Acceptance Criteria

NOTE 1—Table 6 data is preliminary until a multi-lab validation study is completed.

Analyte	Spike Conc. ng/L	Initial Demonstration of Performance			Laboratory Control Sample	
		Recovery (%)		Precision	Recovery (%)	
		Lower Limit	Upper Limit	Maximum % RSD	Lower Control Limit (LCL) %	Upper Control Limit (UCL) %
TTPC	2000	70	130	30	70	130

TABLE 7 MS/MSD QC Acceptance Criteria

NOTE 1—Table 7 data is preliminary until a multi-lab validation study is completed.

Analyte	Spike Conc. ng/L	MS/MSD		Precision
		Recovery (%)		RPD (%)
		Lower Limit	Upper Limit	
TTPC	2000	70	130	30

TABLE 8 Single-Laboratory Recovery Data in Water

Sample	Reagent Water P&A Data (2000 ng/L spike)	CRW P&A Data (2000 ng/L spike)
	TTPC	TTPC
MB 1	<RL	<RL
MB 2	<RL	<RL
P&A 1	1861	1898
P&A 2	1891	1965
P&A 3	1882	1933
P&A 4	1894	1871
P&A 5	1975	1929
P&A 6	1975	1974
Average Recovery (ng/L)	1893	1928
% Average Recovery	94.6	96.4
Standard Deviation	42.8	39.1
RSD (%)	2.26	2.03

6.4 Matrix interferences may be caused by contaminants in the sample. The extent of matrix interferences can vary considerably depending on variations in the sample matrices.

6.5 Automatic pipettes with polypropylene tips were used to prepare the standards, spiking, and calibration solutions. The use of glass syringes for preparing standards, spiking, and calibration solutions generated erratic results; glass syringes should be avoided for these critical tasks. A thoroughly cleaned 10 mL or 20 mL hypodermic glass syringe with a nylon filter was found to perform well to filter the 10 mL prepared sample. 20 mL Luer-Lock polypropylene syringes have also been shown to meet the performance of this test method; these are disposable and do not require cleaning before use. It seems when preparing small volumes of solutions, concentrations are affected by adhesion of TTPC to the syringe barrel or plunger.

NOTE 2—The use of PTFE, PVDF and polypropylene filter units resulted in poor performance and low recoveries.

7. Apparatus

7.1 LC/MS/MS System:

7.1.1 *Liquid Chromatography System*—A complete LC system is required in order to analyze samples, this should include a sample injection system, a solvent pumping system capable of mixing solvents, a sample compartment capable of maintaining required temperature and a temperature controlled

column compartment. A LC system that is capable of performing at the flows, pressures, controlled temperatures, sample volumes, and requirements of this test method must be used.

7.1.2 *Analytical Column*¹⁰—A reverse phase C18 particle column was used to develop this test method. Any column that achieves adequate resolution may be used. The retention times and order of elution may change depending on the column used and need to be monitored.

7.2 *Tandem Mass Spectrometer System*—A MS/MS system capable of multiple reaction monitoring (MRM) analysis or any system that is capable of meeting the requirements in this test method must be used.

7.3 *Adjustable Volume Pipettes*—10 µL, 20 µL, 100 µL, 1000 µL, 5 mL, and 10 mL.

NOTE 3—Any pipette may be used providing the data generated meets the performance of this test method.

7.3.1 *Pipette Tips*—Polypropylene pipette tips free of release agents or low retention coating of various sizes.

¹⁰ A Waters Acquity UPLC BEH C18, 2.1 mm × 100 mm and 1.7 µm particle size column was used. If you are aware of an alternative column that meets the performance of the standard, please provide this information to ASTM International Headquarters. Your comments will receive careful consideration at a meeting of the responsible technical committee,¹ which you may attend.

7.4 *Class A Volumetric Glassware.*

7.5 *Filtration Device.*

7.5.1 *Hypodermic Syringe*—A Luer-Lock tip glass or polypropylene syringe capable of holding a syringe driven filter unit.

7.5.2 A 10 mL or 20 mL Lock Tip Syringe size is recommended since a 10 mL prepared sample size is used in this test method. If a smaller volume syringe is used, do not wash out the syringe or change filters while filtering the same sample if multiple refills of the syringe is required in order to filter the 10 mL prepared sample.

7.5.3 *Filter Unit*¹¹—Nylon filter units were used to filter the samples.

7.6 *Vials*—2-mL autosampler vials (LC vials) with pre-slit PTFE/silicone septa or equivalent.

7.7 *VOA Vials*—40 mL.

7.8 *Sonicator.*

7.9 *Oven*, capable to achieve 250 °C.

8. Reagents and Materials

8.1 *Purity of Reagents*—High Performance Liquid Chromatography (HPLC) pesticide residue analysis and spectrophotometry grade chemicals must be used in all tests. Unless indicated otherwise, it is intended that all reagents must conform to the Committee on Analytical Reagents of the American Chemical Society.¹² Other reagent grades may be used provided they are first determined to be of sufficiently high purity to permit their use without affecting the accuracy of the measurements.

8.2 *Purity of Water*—Unless otherwise indicated, references to water must be understood to mean reagent water conforming to Type 1 of Specification D1193. It must be demonstrated that this water does not contain contaminants at concentrations sufficient to interfere with the analysis.

8.3 All prepared solutions are routinely replaced every year if not previously discarded for quality control failure.

8.4 *Gases*—Ultrapure nitrogen and argon.

8.5 Acetone (CAS # 67-64-1).

8.6 Acetonitrile (CAS # 75-05-8).

8.7 Methanol (CAS # 67-56-1).

8.8 Ammonium Acetate (CAS # 631-61-8).

8.9 2-Propanol (isopropyl alcohol, CAS # 67-63-0).

8.10 Ottawa Sand (CAS # 14808-60-7).

8.11 (Tri-n-butyl)-n-tetradecylphosphonium chloride (CAS # 81741-28-8).

8.12 (Tri-n-butyl)-n-tetradecylphosphonium bromide –D29 (TTPBr-D29, CAS # NA, optional surrogate).

9. Hazards

9.1 Normal laboratory safety applies to this method. Analysts should wear safety glasses, gloves, and lab coats when working in the lab. Analysts should review the Safety Data Sheets (SDS) for all reagents used in this method.

10. Sampling and Preservation

10.1 Grab samples are collected in glass containers with inert-lined caps. As part of the overall quality assurance program for this test method, field blanks exposed to the same field conditions as samples are collected and analyzed according to this test method to assess the potential for field contamination, refer to Guide D3856 as a guide for sampling. This test method is based on a 2.5 mL sample size per analysis. If different sample sizes are used, spiking solution amounts may need to be modified. EPA Publication SW-8466 may be used as a sampling guide. Samples must be shipped with a trip blank between above freezing and 6 °C.

10.2 Once received the sample temperature is taken and should be less than 6 °C. If the receiving temperature is greater than 6 °C, the sample temperature is noted in the case narrative accompanying the data. Samples should be stored refrigerated between above freezing and 6 °C from the time of collection until analysis.

10.3 The sample should be analyzed within 14 days of collection. No holding time study has been done on water matrices tested in this test method. Holding time may vary depending on the matrix and individual laboratories should determine the holding time in their matrix, refer to Practice D4841.

11. Preparation of LC/MS/MS

11.1 *LC Chromatograph Operating Conditions:*

11.1.1 Injections of all standards and samples are made at a 50 µL volume. Other injection volumes may be used to optimize conditions. Standards and sample extracts must be in a 75:25 acetone:water solution. In the case of extreme concentration differences amongst samples, it is wise to analyze a blank after a concentrated sample and before a dilute sample to minimize carry-over of analytes from injection to injection. However, there should not be carry-over between samples. The LC utilized to develop this test method has a flow through LC needle design. The gradient conditions for liquid chromatography are shown in Table 2.

11.2 *LC Sample Manager Conditions:*

11.2.1 *Needle Wash Solvent*—60 % acetonitrile/40 % 2-propanol. Eight-second wash time before and after injection. Instrument manufacturer's specifications should be followed in order to eliminate sample carry-over.

11.2.2 *Temperatures*—Column, 35 °C; sample compartment, 15 °C.

¹¹ A Whatman Puradisc 25 NYL Disposable Filter unit (diameter 25 mm, 0.2 µm nylon membrane syringe driven filter unit) was used. If you are aware of an alternative filter that meets the performance of the standard, please provide this information to ASTM International Headquarters. Your comments will receive careful consideration at a meeting of the responsible technical committee,¹ which you may attend.

¹² *ACS Reagent Chemicals, Specifications and Procedures for Reagents and Standard-Grade Reference Materials*, American Chemical Society, Washington, DC. For suggestions on the testing of reagents not listed by the American Chemical Society, see *Analar Standards for Laboratory Chemicals*, BDH Ltd., Poole, Dorset, U.K., and the *United States Pharmacopeia and National Formulary*, U.S. Pharmacopeial Convention, Inc. (USPC), Rockville, MD.

11.2.3 *Seal Wash*—Solvent: 50 % methanol/50 % water; time: 5 min.

11.3 *Mass Spectrometer Parameters:*

11.3.1 To acquire the maximum number of data points per SRM channel while maintaining adequate sensitivity, the tune parameters may be optimized according to the instrument used. Each peak requires at least 10 scans per peak for adequate quantitation. Variable parameters regarding SRM transitions, and cone and collision energies are shown in [Table 3](#). The values for the following parameters are shown here for information only. These conditions should be checked and optimized when required. Mass spectrometer parameters used in the development of this method are listed below:

The instrument is set in the Electrospray positive source setting:

Capillary Voltage: 1 kV
 Cone: Variable depending on analyte
 Source Temperature: 150 °C
 Desolvation Gas Temperature: 500 °C
 Desolvation Gas Flow: 900 L/h
 Cone Gas Flow: 150 L/h
 Collision Gas Flow: 0.15 mL/min
 Low Mass Resolution 1:3
 High Mass Resolution 1:14
 Ion Energy 1:1
 Entrance Energy: 1
 Collision Energy: Variable depending on analyte
 Exit Energy: 1
 Low Mass Resolution 2: 2.8
 High Mass Resolution 2:14
 Ion Energy 2:1
 Gain: 1.0
 Multiplier: 512.82
 Inter-Scan Delay: 0.003 s

12. Calibration and Standardization

12.1 The mass spectrometer must be calibrated as in accordance with manufacturer’s specifications before analysis. Analytical values satisfying test method criteria have been achieved using the following procedures. Prepare all solutions in the lab using Class A volumetric glassware.

12.2 *Calibration and Standardization*—To generate a calibration curve, analyze eight calibration standards of the TTPC compound prior to sample analysis as shown in [Table 4](#). Calibration stock standard solution is prepared from the target spike solution directly to ensure consistency. Stock standard Solution A containing the TTPC is prepared at Level 8 concentration and aliquots of that solution are diluted to prepare Levels 1 through 7. The following steps will produce 1 mL calibration standards with the concentration values shown in [Table 4](#). The analyst is responsible for recording initial component weights carefully when working with pure materials and correctly carrying the weights through the dilution calculations.

12.3 At a minimum, five calibration levels are required when using a linear calibration curve and six calibration levels are required when using a quadratic calibration curve. An initial eight-point curve may be used to allow for the dropping of the lower level calibration points if the individual laboratory’s instrument can’t achieve low detection limits. This should allow for at least a five- or six-point calibration curve to be obtained.

12.3.1 Calibration stock standard Solution A (Level 8, [Table 4](#)) is prepared from the target spike solution directly to ensure

consistency. 500 µL of TTPC Target Spike Solution (100 µg/L, [12.7](#)) is added to a 50 mL volumetric flask and diluted to 50 mL with 75:25 acetone:water. The preparation of the Level 8 standard can be accomplished using appropriate volumes and concentrations of stock solutions as in accordance with a particular laboratory’s standard procedure.

12.3.2 Aliquots of Solution A are then diluted with 75:25 acetone:water to prepare the 1 mL desired calibration levels in 2 mL amber glass LC vials ([Table 5](#)). The calibration vials must be used within 24 h to ensure optimum results. Calibration standards are not filtered.

12.3.3 Inject each standard and obtain its chromatogram. An external calibration technique is used to monitor the primary and confirmatory SRM transitions of TTPC. Calibration software is utilized to conduct the quantitation of the target analyte using the primary SRM transition. The ratios of the primary/confirmatory SRM transitions area counts are given in [Table 3](#) and will vary depending on the individual tuning conditions. The primary/confirmatory SRM transitions area ratio must be within 35 % of the individual labs’ accepted primary/confirmatory SRM transitions area ratio. The primary SRM transition of TTPC is used for quantitation and the confirmatory SRM transitions for confirmation. This gives added confirmation by isolating the parent ion, forming three product ions by means of fragmentation, and relating it to the retention time in the calibration standard.

12.3.4 Depending on sensitivity and matrix interference issues dependent on sample type, a confirmatory SRM transition may be substituted as the primary SRM transition for quantitation during analysis. This must be explained in a narrative accompanying the data. New primary/confirmatory ion ratios will then be determined if switching the SRM transitions used to quantitate and confirm. The new primary/confirmatory SRM transitions area ratio is required to be within 35 % of the individual labs’ new primary/confirmatory SRM transitions area ratio.

12.3.5 The calibration software manual should be consulted to use the software correctly. The quantitation method is set as an external calibration using the peak areas in ppt units. Concentrations may be calculated using the data system software to generate linear regression or quadratic calibration curves. Forcing the calibration curve through the origin ($X=0$, $Y=0$) is not recommended. Curves should be evaluated using relative error or relative standard error.¹³

12.3.6 Linear calibration may be used if the point of origin is excluded and a fit weighting of $1/X$ is used in order to give more emphasis to the lower concentrations. Each calibration point used to generate the curve must have a calculated percent deviation less than 25 % from the generated curve. If the low or high point(s), or both, are excluded, minimally a five-point curve is acceptable but the reporting range must be modified to reflect this change.

12.3.7 Quadratic calibration may be used if the point of origin is excluded, and a fit weighting of $1/X$ is used in order

¹³ Management and Technical Requirements for Laboratories Performing Environmental Analysis; Module 4: Quality Systems for Chemical Testing; The NELAC Institute, 2017.

to give more emphasis to the lower concentrations. If the low or high point(s), or both, are excluded, minimally a six-point curve is acceptable but the reporting range must be modified to reflect this change. Each calibration point used to generate the curve must have a calculated percent deviation less than 25 % from the generated curve.

12.3.8 The retention time window of the SRM transitions must be within 5 % of the retention time of the analyte in a midpoint calibration standard. If this is not the case, re-analyze the calibration curve to determine if there was a shift in retention time during the analysis and the sample needs to be re-injected. If the retention time is still incorrect in the sample, refer to the analyte as an unknown.

12.3.9 A midpoint calibration check standard must be analyzed at the end of each batch of 30 samples or within 24 h after the initial calibration curve was generated, the criteria in the individual labs' quality system may be more restrictive pertaining to the number of samples. This end calibration check should come from the same calibration standard solution that was used to generate the initial curve. The results from the end calibration check standard must have a percent deviation less than 30 % from the calculated concentration for the target analyte. If the results are not within these criteria, corrective action including re-occurrence minimization is performed and either all samples in the batch are re-analyzed against a new calibration curve or the affected results are qualified with an indication that they do not fall within the performance criteria of the test method. If the analyst inspects the vial containing the end calibration check standard and notices that the sample evaporated affecting the concentration or other anomaly, a new end calibration check standard may be made and analyzed. If this new end calibration check standard has a percent deviation less than 30 % from the calculated concentration for the target analyte, the results may be reported unqualified.

12.4 If a laboratory has not performed the test before or if there has been a major change in the measurement system, for example, new analyst, new instrument, etc., an instrument qualification study including method detection limit (MDL), calibration range determination and precision and bias determination must be performed to demonstrate laboratory capability.

12.4.1 Analyze at least four replicates of a spiked water sample containing TTPC at a prepared sample concentration in the calibration range of Levels 3–6. A 2000 ng/L sample concentration was used to set the QC acceptance criteria in this method. The matrix and chemistry should be similar to the matrix used in this test method. Each replicate must be taken through the complete analytical test method including any sample manipulation and preparation steps.

12.4.2 Calculate the mean (average) percent recovery and relative standard deviation (RSD) of the four values and compare to the acceptable ranges of the QC acceptance criteria for the Initial Demonstration of Performance in [Table 6](#).

12.4.3 This study should be repeated until the single operator precision and mean recovery are within the limits in [Table 6](#). If a concentration other than the recommended concentration

is used, refer to Practice [D5847](#) for information on applying the F-test and t-test in evaluating the acceptability of the mean and standard deviation.

12.4.3.1 The QC acceptance criteria for the Initial Demonstration of Performance in [Table 6](#) were generated from the single-laboratory data shown in the Precision and Bias, Section [16](#). Data from reagent water and river water are shown in the Precision and Bias, Section [16](#). It is recommended that each laboratory determine in-house QC acceptance criteria which meet or exceed the criteria in this test method. References generating QC acceptance criteria are Practices [D2777](#), [D5847](#), and [E2554](#), and Method 8000 in EPA Publication SW-846.

12.5 *Surrogate Spiking Solution:*

12.5.1 A surrogate spiking solution containing TTPBr-D29, or similar isotopically labeled standard should be used and added to all samples. The surrogate concentration should be at a similar concentration as the target spike. At the time of standard development a labelled TTPC surrogate was not available.¹⁴ QA/QC criteria will have to be established by the laboratory and should be similar to the unlabeled TTPC target analyte since it is the isotopically labeled counterpart.

12.6 *Method Blank:*

12.6.1 A method blank for every 30 samples is prepared in 2.5 mL of reagent water, which is taken through the sample preparation Section [13](#), to investigate for contamination during sample preparation. The concentration of target analytes in the blank must be at less than half the reporting limit or the data must be qualified as having a blank issue and the reporting limit must be raised to at least three times above the blank contamination concentration.

12.7 *Reporting Limit Check Sample (RLCS):*

12.7.1 Each batch or within the 24 h analysis window a reporting limit check sample must be analyzed. The reporting limit check sample is processed like a Laboratory Control Sample just spiked at or near (1-2 times) the reporting limit. The concentration of the RLCS may be reported below the reporting limit since the spike is at or near the reporting limit. This sample is to ensure if the analyte was present at the reporting limit that it would be identified. The recovery limits for the RLCS are 35 % to 150 %, if TTPC is outside of these limits the QC exceedance is explained in a narrative accompanying the data or the batch is re-prepared and analyzed. A continued failure must be explained, investigated and should be corrected.

12.7.2 2.5 mL of reagent water is added to a 40 mL VOA vial. The sample is spiked with 25 µL of a 10 µg/L Reporting Limit Check solution in 75 % acetone/25 % water (prepared by dilution from the 100 µg/L target spike solution) is added to the reporting limit check sample to prepare a 100 ng/L sample of TTPC. The sample is then prepared as described in Section [13](#).

12.8 *Laboratory Control Sample (LCS):*

12.8.1 Analyze at least one LCS with the TTPC at a mid-level prepared sample concentration. The concentration of TTPC at a prepared sample concentration in the calibration

¹⁴ A custom synthesis of TTPBr-D29 was undertaken by Cambridge Isotope Laboratories and may be commercially available.