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Ambient air — Determination of the mass concentration of sulfur dioxide — Tetrachloromercurate (TCM)/pararosaniline method

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Air ambiant — Détermination de la concentration en masse du dioxyde de soufre — Méthode au tétrachloromercurate (TCM) et à la pararosaniline 67:1990

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Foreword

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Draft International Standards adopted by the technical committees are circulated to the member bodies for voting. Publication as an International Standard requires approval by at least 75 % of the member bodies casting a vote.

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International Standard ISO 6767 was prepared by Technical Committee ISO/TC 146, Air quality.

Annexes A and B form an integral part of this International Standard.

Annex C is for information only.

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Introduction

This International Standard is a guideline, based on the West-Gaeke method, for the determination of the mass concentration of sulfur dioxide in ambient air. In annex A, a test is given for the purity and purification of pararosaniline hydrochloride; in annex B, a determination is given for the mass concentration of sulfur dioxide present in the sodium disulfite solution used for routine checks; in annex C, a method is given for the recovery of mercury from fresh and used solutions.

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Ambient air — Determination of the mass concentration of sulfur dioxide — Tetrachloromercurate (TCM)/pararosaniline method

1 Scope

This International Standard specifies a spectrophotometric method, known as the tetrachloromercurate (TCM)/pararosaniline method, for the determination of the mass concentration of sulfur dioxide in ambient air within the range of $20~\mu g/m^3$ to about $500~\mu g/m^3$.

The sampling period is 30 min to 60 min.

If a longer sampling period than 60 min is used, or higher concentrations of sulfur dioxide (up to about 2 000 µg/m³) are expected, care is necessary to en 67:1990 bration sure that the concentrations of sulfur dioxide in the reds/six/do/11 absorption solution given in clause 6, paragraph 2 so-6767-1990 are not exceeded. This can be achieved by a reduction of the volume flow rate during sampling. Sample solutions obtained by this procedure may be stored for up to 24 h before making measurements, provided that they are kept in a refrigerator at about 5 °C.

Substances which are known to interfere and which might be present in the air being sampled are listed in 7.5

Indications of the precision and accuracy of the method, and of the lower detection limit are given in 8.2.

Detection limit, standard deviations and interferences qualify the TCM-method for orientating field measurements in the higher concentration range. When more accurate measurements are necessary, instruments which are specially tested and calibrated should be used.

2 Normative references

The following standards contain provisions which, through reference in this text, constitute provisions of this International Standard. At the time of publi-

cation, the editions indicated were valid. All standards are subject to revision, and parties to agreements based on this International Standard are encouraged to investigate the possibility of applying the most recent editions of the standards indicated below. Members of IEC and ISO maintain registers of currently valid International Standards.

ISO 4219:1979. Air quality — Determination of gaseous sulphur compounds in ambient air — Sampling equipment.

ISO 6349:1979, Gas analysis — Preparation of calibration gas mixtures — Permeation method.

3 Principle

Absorption of sulfur dioxide present in the air sample by passage through a sodium tetrachloromercurate (TCM) solution within a specified period resulting in the formation of a dichlorosulfitomercurate complex.

Destruction of any nitrite ions formed in the sodium tetrachloromercurate solution, by nitrogen oxides present in the air sample, by adding sulfamic acid solution. Conversion of the dichlorosulfitomercurate complex into intensely violet coloured pararosaniline methyl sulfonic acid, by adding a formaldehyde solution and an acidified pararosaniline hydrochloride solution to the resultant solution.

Determination of the absorbance of the sample solution at a wavelength of about 550 nm using an appropriate spectrophotometer (or colorimeter) and calculation of the mass concentration of sulfur dioxide by means of a calibration graph prepared using calibration gas mixtures.

Depending on the the equipment available in the laboratory, it may be convenient to use sodium disulfite solutions for routine checks. However, the

solutions are only used after proper calibration using a permeation device.

Reagents

During the analysis, use only reagents of recognized analytical grade and only distilled water, preferably double-distilled water, or water of equivalent purity, free from oxidants.

WARNING — Use the reagents in accordance with the appropriate health and safety regulations.

4.1 Sodium tetrachloromercurate (TCM), absorption solution, $c(Na_2[HgCl_4]) = 0.04 \text{ mol/l.}$

Dissolve 10,9 g of mercury(II) chloride (HgCl₂), 4,7 g of sodium chloride (NaCl) and 0,07 g of ethylenedinitrilo tetraacetic acid, disodium salt dihydrate [(HOCOCH₂)₂N(CH₂)₂N(CH₂COONa)₂.2H₂O] EDTA in water in a 1 000 ml one-mark volumetric flask. Make up to the mark with water and mix well.

Store the solution in a well stoppered bottle.

The solution is stable for several months, but discard it if a precipitate is formed. iTeh STA

A procedure for recovering mercury from fresh and used solutions is given in annex C.

- 4.2 Pararosaniline hydrochloride (PRA), 0,16 g/l ISO 6 this solution contains the mass equivalent of solution.
- **4.2.1** Transfer 86 ml of approximately 38 % (m/m)hydrochloric acid (HCl) ($\rho \approx 1,19 \text{ g/ml}$) into a 1000 ml one-mark volumetric flask. Make up to the mark with water and mix well.
- **4.2.2** Transfer 205 ml of approximately 85 % (m/m)phosphoric acid H_3PO_4 ($\rho \approx 1,69 \text{ g/ml}$) into a 1000 ml one-mark volumetric flask. Make up to the mark with water and mix well.
- 4.2.3 Dissolve 0,2 g of pararosaniline hydrochloride (C₁₉H₁₇N₃.HCl), the purity of which has been tested for each new PRA batch according to annex A, in 100 ml of the hydrochloric acid solution prepared in 4.2.1.
- 4.2.4 Pipette 20 ml of the solution prepared in 4.2.3 into a 250 ml one-mark volumetric flask. Add 25 ml of the phosphoric acid solution prepared in 4.2.2. Mix well, and make up to the mark with water.

The solution is stable for several months if stored in the dark.

If the PRA solution is prepared from a PRA stock solution obtained by the purification of PRA in accordance with clause A.2, for each 1 % of difference between a degree of purity of 100 % and the degree of purity obtained, add an additional 0,2 ml of PRA stock solution before making up to the mark with water.

4.3 Formaldehyde, approximately 2 g/l solution.

Pipette commercially 5 ml of a available 36 % (m/m) to 38 % (m/m) formaldehyde (HCHO) solution into a 1000 ml one-mark volumetric flask. Make up to the mark with water and mix well.

Prepare this solution on the day of use.

4.4 Sulfamic acid, 6 g/l solution.

Dissolve 0,6 g of sulfamic acid (NH2SO3H) in 100 ml of water.

This solution is stable for a few days, if protected from air.

- **4.5 Sodium disulfite**, 0,012 g/l solution.
- 4.5.1 Dissolve 0,3 g of sodium disulfite (Na₂S₂O₅) in 500 ml of freshly distilled water which has been deaerated, for example by boiling and cooling to room temperature.

This solution is not stable.

https://standards.itch.ai/catalog/stang20s/pgt/66/400 pgsof sulfurbdioxide per millilitre. Deb015cde5549termine, as specified in annex B, the actual mass concentration of sulfur dioxide present in the solution.

> 4.5.2 Immediately after the determination of the actual mass concentration of sulfur dioxide present in the solution as specified in 4.5.1, pipette 2,0 ml of this solution into a 100 ml one-mark volumetric flask. Make up to the mark with absorption solution (4.1) and mix well.

> This solution is stable for 30 days, if stored at about 5 °C, or for one day only, if stored at room temperature.

4.6 Calibration gas mixtures.

Immediately before use, prepare zero gas and gas mixtures of sulfur dioxide and air, using the permeation technique specified in ISO 6349. The latter shall be available in at least four different mass concentration levels of sulfur dioxide covering the desired working range.

Apparatus 5

Ordinary laboratory apparatus and

- **5.1 Sampling equipment**, as specified in ISO 4219 and in 5.1.1 to 5.1.5.
- **5.1.1 Air intake** (see ISO 4219), made of polytetra-fluoroethylene or borosilicate glass washed with perchloric acid then with distilled water and dried.

5.1.2 Filter for particulate matter (see ISO 4219).

Whenever possible, avoid the use of a filter for particulate matter. However, if a filter for particulate matter is used it shall be made of a material which meets the requirements specified in ISO 4219.

5.1.3 Absorber (see ISO 4219).

The absorption efficiency of the absorption bottle for sulfur dioxide shall be determined before use and shall be at least 0,95. Examples of suitable absorption bottles are given in figure 1.

The absorption efficiency varies with the geometry of the bottle, the size of the gas bubbles and their contact time with the solution. It can be determined by inserting into the sampling train a second absorber in series with the first, and relating the amount of sulfur dioxide found in the first bottle to the total amount of sulfur dioxide found in the two absorbers. When working with midget impingers under the conditions described in clause 6, the absorber sorption efficiency was found to be better than 0.98 and the conditions described in clause 6.

The use of mixtures of sulfur dioxide and air for calibration, as described in 7.1, automatically takes into account the absorption efficiency of the system.

5.1.4 Gas meter or **air-flow regulator** (see ISO 4219).

As an alternative to the gas meter, a critical orifice in a temperature controlled box can be used. In this case, the pump shall be able to reach $p_{\rm d}/p_{\rm u} \le 0.5$, where $p_{\rm d}$ and $p_{\rm u}$ are the pressures downstream and upstream of the needle, respectively (see 5.1.5).

- **5.1.5 Two manometers**, accurate to 1 kPa, for measurement of pressure when using a critical orifice instead of a gas meter.
- **5.2 Spectrophotometer** or **colorimeter**, suitable for measuring the absorbance at about 550 nm. If a spectrophotometer is used, carry out the measurements with the instrument set at a wavelength of 548 nm. If a colorimeter is used, use a filter that has its transmittance maximum at a wavelength between 540 nm and 550 nm. Reagent blank problems may occur with apparatus having a spectral bandwith greater than 20 nm.

- **5.3 Optical cells**, plane matched pairs, with an optical path length of 1,0 cm to 5,0 cm.
- **5.4 Polyethylene bottle**, of capacity 100 ml, for transferring the exposed absorption solutions to the laboratory.

6 Sampling

Assemble a sampling train in accordance with the examples shown in figure 2 and any special requirements for the air mass under investigation.

Transfer 10 ml of the aborption solution (4.1) to an absorption bottle (5.1.3) and insert it into the sampling train. Choose a sampling period of either 30 min or 60 min and a volume flow rate between 0.5 l/min and 1 l/min.

The best results are obtained if 0,25 μ g to 2,5 μ g (0,1 μ l to 0,95 μ l at 25 °C and 101,3 kPa) of sulfur dioxide per millilitre of absorption solution is trapped.

After sampling, determine the volume of air sampled and note the atmospheric pressure (see 8.1, note 3). If the laboratory sample has to be stored before analysis, it may be kept at 5 °C for not longer than 24 h.

If the laboratory sample shows a precipitate, it is probably due to the reaction of mercury(II) with a reducing sulfur compound. Remove the precipitate by filtration or centrifugation before the analysis.

7 Procedure

7.1 Calibration

7.1.1 Preparation of a set of calibration solutions

Mixtures of sulfur dioxide and air are prepared according to ISO 6349.

In order to prepare the calibration graph, which is a plot of absorbance versus sulfur dioxide concentration, at least four different concentration levels of sulfur dioxide within the range specified in clause 2 are needed.

Sample each gas mixture under the same conditions as are used for the unknown air sample, in particular for the same sampling period and at the same flow rate. Treat the obtained sample solutions as described in 7.3 (see 4.6).

7.1.2 Zero member solution

Prepare a zero member solution from zero gas (see 4.6). This preparation is carried out according to 7.1.1.

7.1.3 Plotting the calibration graph

Correct the absorbance values to allow for the absorbance of the blank solution (7.4). Plot the net absorbance of each solution against the mass, in micrograms, of sulfur dioxide in the gas sample from which it was derived. Calculate the calibration factor f (reciprocal of the slope of the line) for use in the calculation of results (8.1).

7.2 Preparation of routine check graph

Pipette 0 ml, 1,0 ml, 2,0 ml, 3,0 ml and 4,0 ml of sodium disulfite solution (4.5.2) into a series of 25 ml one-mark volumetric flasks. Add sufficient absorption solution (4.1) to each flask to bring the volume to approximately 10 ml. Then carry out the analysis as described in 7.3.2.

Plot the absorbances of the solutions as ordinates against the mass, in micrograms, of sulfur dioxide calculated according to annex B. A linear relationship is obtained. The intercept of the line best fitting the points with the vertical axis is usually within 0,02 absorbance units of the zero member solution reading if 1 cm cells are used. Calculate the calibration factor f' (reciprocal of the slope of the line). This calibration factor can be used for routine purposes; the value obtained when dividing f' by the volume, in cubic metres, of air sampled during the calibration procedure according to 7.1, shall differ by less than 10 % from the calibration factor f ob- 150 6

7.3 **Determination**

7.3.1 Leave the sample (clause 6) for at least 20 min after sampling to allow trapped ozone to decompose. Then transfer the sample solution quantitatively to a 25 ml one-mark volumetric flask. using about 5 ml of water for rinsing.

7.3.2 Add 1 ml of the sulfamic acid solution (4.4) to each flask and allow to react for 10 min to destroy the nitrites from oxides of nitrogen. Then pipette 2,0 ml of the formaldehyde solution (4.3) followed by 5 ml of the pararosaniline reagent (4.2.4) into the flasks. Make up to the mark with freshly boiled and cooled distilled water and store at 20 °C \pm 1 °C. Using the spectrophotometer or colorimeter (5.2) measure the absorbance of the sample solution and the zero member solution, against distilled water in the reference cell, between 30 min and 60 min after the addition of the reagents and immediately after filling the cells.

Do not allow the coloured solutions to remain in the cell as a coloured film will be deposited on the inside walls.

NOTE 2 Fixed time intervals between the addition of each reagent, for example 1 min, ensure a better reproducibility of the formation of the absorbing compound.

Preparation of the blank solution

Prepare a blank by pipetting 10 ml of unexposed absorption solution (4.1) into a 25 ml one-mark volumetric flask, add the reagents as before, treat the solution as described in 7.3.2 and read the absorbance against distilled water using 1 cm cells. Compare this value with the one recorded for the zero member solution obtained when preparing the calibration graph (7.1). Differences of more than 10 % between the two values indicate contamination of the distilled water or the reagents or decomposition of the latter, in which case fresh reagents shall be prepared.

7.5 Interference

The amount of EDTA added eliminates interference from heavy metals: up to 30 µg of manganese(II), 10 μg of chromium(III), 10 μg of copper(II) and 22 μg of vanadium(V) in 10 ml of the absorption solution.

If sulfamic acid (4.4) is used with the described procedure, up to 50 µg of nitrogen dioxide per 10 ml of absorption solution can be tolerated.

Interference from oxides of nitrogen, ozone and reducing sulfur compounds (for example hydrogen sulfide and mercaptans) are eliminated or minitained with calibration gas mixturestandards.iteh.ai/catalog/standmizedt/dsulfurid-acid and sulfates do not interfere. b015cde5549No interference by sulfur trioxide has been proved experimentally, since this presumably becomes hydrolysed to sulfuric acid in the absorption solution.

Expression of results

8.1 Calculation

Calculate the mass concentration of sulfur dioxide, $\rho(SO_2)$, expressed in micrograms per cubic metre. by using the equations

$$\rho(\mathrm{SO_2}) = \frac{f(A_\mathrm{s} - A_\mathrm{b})}{V_\mathrm{1}}$$

$$\rho(\mathrm{SO_2}) = \frac{f'(A_\mathrm{s} - A_\mathrm{b})}{V_\mathrm{1}}$$

where

f and f' are calibration factors (see 7.1 and 7.2):

 A_{ς} is the absorbance of the sample solution;

is the absorbance of the blank; A_{b}

 V_1 is the volume, in cubic metres, of the air sample.

NOTE 3 If the sulfur dioxide concentration at reference conditions (298 K, 101,3 kPa $\approx 10^2$ kPa) is needed, replace the volume V_1 of air sampled by the corresponding value V_2 of the volume under the reference conditions:

$$V_2 = \frac{298 \, V_1 p}{(273 + t)10^2}$$

where

p is the barometric pressure, in kilopascals;

t is the temperature, in degrees Celsius, of the air sample.

8.2 Performance characteristics

8.2.1 Precision and accuracy

The method is not known to have any inherent bias or inaccuracy. The accuracy achieved in practice will depend on the care used in performing the various calibrations and measurements.

As an indication of performance, a relative standard deviation of the order of \pm 10 % should be achievable for a measured concentration between 80 μg SO₂/m³ and 200 μg SO₂/m³ in ambient ar and ard S

8.2.2 Lower detection limit

The lower detection limit of sulfur dioxide in 10 ml of TCM sample solution is between 0,2 μ g and 1,0 μ g, based on twice the standard deviation of the zero member solution, (7.1.2). This corresponds to a concentration between 7 μ g SO₂/m³ and 33 μ g SO₂/m³ [0,002 ppm (V/V) to 0,011 ppm (V/V) in an air sample of 30 litres (for example 1 h sampling at 0,5 Vmin)].

9 Test report

The test report shall include the following information:

- a) a complete identification of the air sample;
- b) a reference to this International Standard;
- c) the results obtained;
- d) any unusual features noted during the determination;

e) any operation not specified in this International Standard or in the International Standards to which reference is made, or regarded as op-

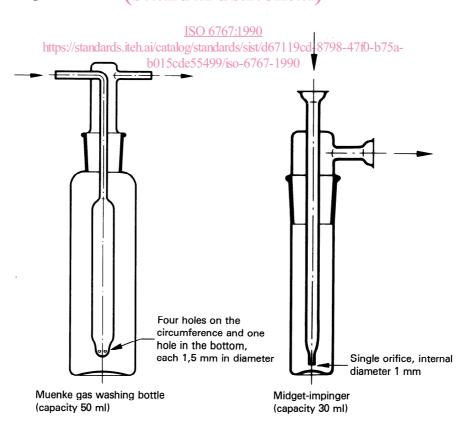


Figure 1 — Examples of absorption bottles suitable for sampling sulfur dioxide in air