INTERNATIONAL STANDARD

ISO 7899-1

Second edition
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Water quality — Detection and enumeration of intestinal enterococci in surface and waste water —

Part 1:
Miniaturized method (Most Probable Number) by inoculation in liquid medium

Qualité de l'eau — Recherche et dénombrement des entérocoques intestinaux dans les eaux de surface et résiduaires —

Partie 1: Méthode miniaturisée (nombre le plus probable) par ensemencement en milieu liquide

Reference numbe
ISO 7899-1:1998(E)

Contents

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Foreword

ISO (the International Organization for Standardization) is a worldwide federation of national standards bodies (ISO member bodies). The work of preparing International Standards is normally carried out through ISO technical committees. Each member body interested in a subject for which a technical committee has been established has the right to be represented on that committee. International organizations, governmental and non-governmental, in liaison with ISO, also take part in the work. ISO collaborates closely with the International Electrotechnical Commission (IEC) on all matters of electrotechnical standardization.

Draft International Standards adopted by the technical committees are circulated to the member bodies for voting. Publication as an International Standard requires approval by at least 75 % of the member bodies casting a vote.

International Standard ISO 7899-1 was prepared by Technical Committee ISO/TC 147, *Water quality*, Subcommittee SC 4, *Biological methods.*

This second edition cancels and replaces the first edition (ISO 7899-1:1984), which has been technically revised.

ISO 7899 consists of the following parts, under the general title *Water quality — Detection and enumeration of intestinal enterococci in surface and waste water*:

— *Part 1: Miniaturized method (Most Probable Number) by inoculation in liquid medium*

— *Part 2: Method by membrane filtration*

Annexes E and F form an integral part of this part of ISO 7899. Annexes A, B, C, D and G are for information only.

Introduction

The aim of this part of ISO 7899 is to enumerate the major intestinal enterococci, namely *E. faecalis*, *E. faecium*, *E. durans* and *E. hirae*, which occur frequently in faeces of humans and homeothermic animals. Other faecal *Enterococcus* species, namely *E. avium*, *E. cecorum*, *E. columbae* and *E. gallinarum*, and *Streptococcus bovis*/*equinus* strains may occasionally be included, but they occur rarely in the environmental samples. Their recovery tends to be low. *Enterococcus casseliflavus* and *E. mundtii* are non-faecal species which, when present in water samples (e.g. because of influence of plant material and some industrial effluents), are enumerated as faecal enterococci. These species and other rare non-faecal species tend to produce yellow pigment on a non-selective medium. The possible interference of non-faecal *Enterococcus* species should therefore be considered in the interpretation of results.

Water quality — Detection and enumeration of intestinal enterococci in surface and waste water —

Part 1:
Miniaturized method (Most Probable Number) by inoculation in liquid medium

1 Scope

This part of ISO 7899 specifies a miniaturized method for the detection and enumeration of major intestinal enterococci in surface and waste water by inoculation in a liquid medium. The method is applicable to all types of surface and waste waters, particularly those rich in suspended matter.

This method is not suitable for drinking water and any other type of water for which the guideline count is less than 15 per 100 ml.

2 Normative references

The following standards contain provisions which, through reference in this text, constitute provisions of this part of ISO 7899. At the time of publication, the editions indicated were valid. All standards are subject to revision, and parties to agreements based on this part of ISO 7899 are encouraged to investigate the possibility of applying the most recent editions of the standards indicated below. Members of IEC and ISO maintain registers of currently valid International Standards.

ISO 3951:1989, *Sampling procedures and charts for inspection by variables for percent nonconforming*.

ISO 5667-1:1980, *Water quality — Sampling — Part 1: Guidance on the design of sampling programmes.*

ISO 5667-2:1991, *Water quality — Sampling — Part 2: Guidance on sampling techniques.*

ISO 5667-3:1994, *Water quality — Sampling — Part 3: Guidance on the preservation and handling of samples.*

ISO 8199:1988, *Water quality — General guide to the enumeration of microorganisms by culture*.

ISO/IEC Guide 2:1996, *Standardization and related activities — Vocabulary.*

3 Definitions

For the purposes of this part of ISO 7899, the definitions given in ISO/IEC Guide 2 and the following definition apply.

3.1
intestinal enterococci
microorganisms capable of aerobic growth at 44 °C and of hydrolysing the 4-methylumbelliferyl-β-D-glucoside (MUD), in the presence of thallium acetate, nalidixic acid and 2,3,5-triphenyltetrazolium chloride (TTC), in the liquid medium specified

4 Principle

The diluted sample is inoculated in a row of microtitre plate wells containing dehydrated culture medium.

The microtitre plates are examined under ultraviolet light at 366 nm in the dark after an incubation period of between 36 h and 72 h at 44 °C ± 0,5 °C. The presence of enterococci is indicated by fluorescence resulting from the hydrolysis of MUD. The results are given as Most Probable Number (MPN) per 100 ml.

5 Apparatus

With the exception of equipment supplied sterile, the glassware shall be sterilized in accordance with the instructions given in ISO 8199.

Usual microbiological laboratory equipment, and in particular:

5.1 Apparatus for sterilization by dry heat (oven) or by steam (autoclave).

5.2 Thermostatic incubator, regulated at 44 °C ± 0,5 °C.

5.3 Tunnel drier or **vertical laminar air flow cabinet** (preferably class II).

5.4 UV observation chamber (Wood's Lamp 366 nm).

WARNING — UV light can cause irritation of skin and eyes. Use protective gloves and glasses.

5.5 Portable refractometer (optional).

5.6 pH meter, with an accuracy of ± 0,1.

5.7 Test tubes, 16 mm x 160 mm and 20 mm x 200 mm**, or flasks** with similar capacity.

5.8 Adjustable or pre-set 8-channel multipipette, or any system suitable for measuring and distributing 200 µl per well.

5.9 Sterile tips for multipipette.

5.10 Equipment for membrane filtration, in accordance with ISO 8199, including membrane filters with a nominal pore size of 0,2 µm, for sterilization of liquid media.

5.11 Sterile microtitre plates, 96-well, 350 µl, flat-bottomed, nonfluorescent.

5.12 Sterile adhesive cover strips for sealing microtitre plates.

5.13 Sterile Petri dishes, 90 mm in diameter.

6 Sampling

Take the samples and deliver them to the laboratory in accordance with ISO 8199 and ISO 5667-1, ISO 5667-2 and ISO 5667-3.

7 Culture media and diluents

7.1 General instructions

To ensure reproducible results, prepare culture medium and diluents, using either constituents of uniform quality and chemicals of recognized analytical or a dehydrated diluent or complete medium prepared following the manufacturer's instructions. Prepare them with distilled or demineralized water, free from substances capable of inhibiting or promoting growth under the test conditions. If the media are not used immediately, preserve them in the dark at (5 ± 3) °C, for up to one month in conditions avoiding any alterations to their composition.

NOTE The use of chemicals of other grades is permissible providing they are shown to be of equivalent performance in the test.

7.2 Diluent

7.2.1 Special Diluent (SD)

Synthetic sea salt[1)]	22,5 g
Bromophenol blue solution (optional)	10 ml
Demineralized or distilled water (7.2.2)	1000 ml

Sterilize in the autoclave (5.1) at 121 °C ± 3 °C for 15 min to 20 min.

The bromophenol blue solution is prepared by adding 0,04 g in 100 ml of 50 % ethanol. It is used only to colour the SD blue and avoid confusing it with demineralized or distilled water.

7.2.2 Demineralized or distilled water

Water used for dilution shall be demineralized or distilled water free from substances inhibiting growth under the test conditions.

Sterilize in the autoclave (5.1) before use at 121 °C ± 3 °C for 15 min to 20 min.

7.3 Culture medium: MUD/SF medium

7.3.1 Composition

7.3.1.1 Solution A

Tryptose	40 g
KH_2PO_4	10 g
D(+)-galactose	2 g
Polyoxyethylenesorbitan monooleate (Tween® 80[2)])	1,5 ml
Demineralized or distilled water (7.2.2)	900 ml

Add tryptose, KH_2PO_4, galactose and Tween® 80 to 900 ml of water, whilst maintaining gentle heat and magnetic stirring, then bring to the boil until completely dissolved. Allow to cool.

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1) A typical analysis of a commercially available and suitable synthetic sea salt is given in annex C. Pure NaCl solutions are not suitable, as they lead to marked inhibition.

2) Tween® 80 is an example of a suitable product available commercially. This information is given for the convenience of users of this part of ISO 7899 and does not constitute an endorsement by ISO of this product.

7.3.1.2 Solution B

$NaHCO_3$	4 g
Nalidixic acid	250 mg
Demineralized or distilled water (7.2.2)	50 ml

Add both chemicals to 50 ml of water, whilst maintaining gentle heat and magnetic stirring. Allow to cool.

7.3.1.3 Solution C

Thallium(I) acetate	2 g
2,3,5-triphenyltetrazolium chloride	0,1 g
Demineralized or distilled water (7.2.2)	50 ml

Add both chemicals to 50 ml of water, whilst maintaining gentle heat and magnetic stirring. Allow to cool.

7.3.1.4 Solution D

MUD (4-methylumbelliferyl-β-D-glucoside)	150 mg
N,N-dimethylformamide	2 ml

WARNING — Thallium acetate and *N,N*-dimethylformamide are toxic. Use in a chemicals fume hood.

7.3.2 Preparation

Mix together solutions A+B+C+D.

Adjust the pH to 7,5 ± 0,2.

Sterilize by filtration through a membrane of average pore size 0,2 µm (5.10).

Distribute in 96-well microtitre plates (5.11) with a volume of 100 µl of media in each well (minimum capacity 350 µl) and dehydrate immediately in a tunnel drier or laminar air-flow cabinet (5.3).

The manufacturing of the medium shall meet the quality criteria given in annex E.

8 Procedure

8.1 Choice of dilutions

The number of dilutions to inoculate varies according to the presumed level of contamination of the water to be tested. Table 1 gives some examples.

Table 1

Origin of sample	No. of dilutions	No. of wells /dilution	Measurement limits bacteria / 100 ml
Bathing water	2	64 wells to 1/2 32 wells to 1/20	15 to $3,5x10^4$
Other surface water	4	24 wells to 1/2 24 wells to 1/20 24 wells to 1/200 24 wells to 1/2 000	40 to $3,2x10^6$
Waste water and treatment plants	6	16 wells to 1/2 Up to 16 wells to 1/200 000	60 to $6,7x10^8$

8.2 Preparation of dilutions

NOTE These procedures should be performed in a biological safety cabinet, as aerosols may be created by diluting and pipetting.

8.2.1 Fresh and brackish (waste) water [salinity < 30 g/kg, measured with refractometer (5.5) or equivalent method]

Prepare the relevant number of sterile tubes (5.7) in a rack, according to the number of selected dilutions; add 9 ml of the special diluent (7.2.1) to each tube.

Vigorously stir the sample (see clause 6) in order to obtain a homogeneous distribution of the microorganisms and, using a sterile pipette, immediately transfer 9 ml of this homogenized sample to the first tube containing 9 ml of diluent (7.2.1) (1/2 dilution).

Using a fresh pipette, transfer 1 ml of this dilution (homogenized) to the second tube (1/20 dilution).

From the second tube (dilution 1/20 carefully homogenized) proceed, if necessary, to another 1/10 dilution giving the dilution 1/200.

Continue as above until all the dilutions have been prepared.

8.2.2 Sea water (salinity ⩾ 30 g/kg)

Prepare the relevant number of sterile tubes (5.7) in a rack, according to the number of selected dilutions, add 9 ml of demineralized or distilled water (7.2.2) to the first tube and 9 ml of the special diluent (7.2.1) to the other tubes.

Vigorously stir the sample (see clause 6) in order to obtain a homogeneous distribution of the microorganisms and, using a sterile pipette, immediately transfer 9 ml of this homogenized sample to the first tube containing 9 ml water (7.2.2) (1/2 dilution).

Using a fresh sterile pipette, transfer 1 ml of this dilution (homogenized) to the second tube (1/20 dilution).

From the second tube (dilution 1/20 carefully homogenized) proceed, if necessary, to another 1/10 dilution giving the following dilution (1/200).

Continue as above until all the dilutions have been prepared.

8.3 Inoculation and incubation of microtitre plates

8.3.1 Inoculation

Transfer the contents of the first tube of dilution to an empty, sterile Petri dish, of minimum diameter 90 mm.

Using a multichannel pipette (5.8) with 8 sterile tips (5.9), distribute 200 µl into each well of a microtitre plate (5.11) corresponding to this first dilution.

For subsequent dilutions (1/20, 1/200, etc.) operate in an identical manner, changing the Petri dish and the row of 8 sterile tips between each dilution.

Alternatively, any other suitable system (5.8) may be used to distribute 200 µl of each dilution per well in accordance with table 1.

CAUTION — Beware of contamination via overflow from one well to another.

8.3.2 Incubation

Once the microtitre plate is inoculated, cover with the disposable sterile adhesive tape (5.12) provided for this purpose.

Incubate the microtitre plate (5.2) at 44 °C ± 0,5 °C for a minimum of 36 h and a maximum of 72 h.

NOTE The microtitre plates should be handled with care, without tilting.

8.4 Reading of results

Place each microtitre plate, including adhesive, in the UV observation chamber (5.4).

Consider all wells in which a blue fluorescence is observed as being positive.

NOTE The reading may be carried out any time after 36 h, as the fluorescence does not alter with time.

9 Expression of results

9.1 Determination of characteristic number

For each chosen dilution, note the number of positive (+) wells.

EXAMPLE 1 : Bathing water

1/2	32 + out of 64
1/20	5 + out of 32

Record 32/5 as characteristic number

EXAMPLE 2 : Other surface water

1/2	24 + out of 24
1/20	18 + out of 24
1/200	5 + out of 24
1/2 000	1 + out of 24

Record 18/5/1 as characteristic number

EXAMPLE 3 : Waste water

1/2	16 + out of 16
1/20	16 + out of 16
1/200	12 + out of 16
1/2 000	5 + out of 16
1/20 000	0 + out of 16
1/200 000	0 + out of 16

Record 12/5/0 as characteristic number

Where three or more dilutions have been inoculated, a characteristic number of three digits, ending in 0 where possible, shall be recorded, in accordance with ISO 8199.

9.2 Calculation of the MPN and its confidence interval

The MPN is a statistical estimation of the density of microorganisms, assumed to correspond to a Poisson distribution in the volumes inoculated. Confidence intervals are attached to this MPN.

Software shown in annex A or B enable the calculation of the MPN of intestinal enterococci per millilitre of water for each configuration of inoculations and the confidence interval at 95 %.

EXAMPLE 1: Assuming CN is the Characteristic Number, LO the Lower Limit and UP the Upper Limit:

If CN = 32/5, the software in annex A gives 7,56 enterococci per millilitre,

[LO = 5,42 − UP = 10,54],

i.e. 756/100 ml (542 to 1054)

EXAMPLE 2 :

If CN = 18/5/1, the software in annex A gives 159,08/ml,

[LO = 101,99 − UP = 248,11]

EXAMPLE 3 :

If CN = 12/5/0, the software in annex A gives 1 724,61/ml

[LO = 1 003,98 − UP = 2 962,50]

If none of the wells is positive, express the result in the following form:

< *n*/100 ml

where *n* is the MPN for 1 positive well under the dilution conditions employed.

10 Test report

The test report shall include all details necessary for the complete identification of the sample, referring to the method used and the results.

The test report shall also mention any special phenomena observed during the test and any non-specified or optional operations used in the method which may have altered the results.

11 Performance data

Information concerning the repeatability and reproducibility of the procedure, obtained from interlaboratory tests, is given in annex D.