



Standard Test Method for Determining Aerobic Aquatic Biodegradation of Lubricants or Their Components¹

This standard is issued under the fixed designation D 5864; the number immediately following the designation indicates the year of original adoption or, in the case of revision, the year of last revision. A number in parentheses indicates the year of last reapproval. A superscript epsilon (ϵ) indicates an editorial change since the last revision or reapproval.

1. Scope

1.1 This test method covers the determination of the degree of aerobic aquatic biodegradation of fully formulated lubricants or their components on exposure to an inoculum under laboratory conditions.

1.2 This test method is intended to specifically address the difficulties associated with testing water insoluble materials and complex mixtures such as are found in many lubricants.

1.3 This test method is designed to be applicable to all lubricants that are not volatile and are not inhibitory at the test concentration to the organisms present in the inoculum.

1.4 *This standard does not purport to address all of the safety concerns, if any, associated with its use. It is the responsibility of the user of this standard to establish appropriate safety and health practices and determine the applicability of regulatory limitations prior to use.* Specific hazards are discussed in Section 10.

2. Referenced Documents

2.1 ASTM Standards:

- D 1129 Terminology Relating to Water²
- D 1193 Specification for Reagent Water²
- D 1293 Test Methods for pH of Water²
- D 4447 Guide for Disposal of Laboratory Chemicals and Samples³
- D 5291 Test Methods for Instrumental Determination of Carbon, Hydrogen, and Nitrogen in Petroleum Products and Lubricants⁴
- E 943 Terminology Relating to Biological Effects and Environmental Fate⁵

2.2 ISO Standard:⁵

- 4259:1992(E) Petroleum Products—Determination and Application of Precision Data in Relation to Methods of Test

¹ This test method is under the jurisdiction of ASTM Committee D-2 on Petroleum Products and Lubricants and is the direct responsibility of Subcommittee D02.12 on Environmental Standards for Lubricants.

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² *Annual Book of ASTM Standards*, Vol 11.01.

³ *Annual Book of ASTM Standards*, Vol 11.04.

⁴ *Annual Book of ASTM Standards*, Vol 05.03.

⁵ Available from American National Standards Institute, 11 West 42nd St., New York, NY, 10036.

2.3 APHA Standard:⁶

- 2540B Total Solids Dried at 103–105°C
- 9215 Heterotrophic Plate Count

3. Terminology

3.1 Definitions:

3.1.1 Definitions of terms applicable to this test method that are not described herein appear in the *Compilation of ASTM Standard Definitions*, 1990⁷ or Terminology E 943.

3.1.2 *aerobic*, *adj*—(1) taking place in the presence of oxygen, (2) living or active in the presence of oxygen.

3.1.3 *biodegradation*, *n*—the process of chemical breakdown or transformation of a substance caused by organisms or their enzymes.

3.1.4 *biomass*, *n*—any material, excluding fossil fuels, which is or was a living organism or component of a living organism.

3.1.5 *blank*, *n*—a flask containing the test medium and the inoculum with no additional carbon source added.

3.1.6 *inoculum*, *n*—spores, bacteria, single celled organisms, or other live materials, that are introduced into a test medium.

3.1.7 *lag phase*, *n*—the period of physiological activity and diminished cell division following the addition of microorganisms to a new culture medium.⁸

3.1.8 *log phase*, *n*—the period of growth of microorganisms during which cells divide at a constant rate.⁸

3.1.9 *mixed liquor*, *n*—the contents of an aeration tank including the activated sludge mixed with primary effluent or the raw wastewater and return sludge.

3.1.10 *pre-adaptation*, *n*—the incubation of an inoculum in the presence of the test substance which is done prior to the initiation of the test and under conditions similar to the test conditions.

3.1.10.1 *Discussion*—The aim of pre-adaptation is to improve the precision of the test method by decreasing variability in the rate of biodegradation produced by the inoculum. Pre-adaptation may mimic the natural processes which cause

⁶ From *Standard Methods for the Examination of Water and Wastewater*, latest edition. Available from the American Public Health Association, 1015 18th St., N.W., Washington, DC 20036.

⁷ Available from ASTM Headquarters.

⁸ Adapted from *McGraw-Hill Dictionary of Scientific and Technical Terms*, 4th ed., 1989.

changes in the microbial population of the inoculum leading to more rapid biodegradation of the test substance.

3.1.11 *supernatant, n*—the liquid above settled solids.

3.1.12 *theoretical CO₂, n*—the amount of CO₂ which could hypothetically be produced from the complete biological oxidation of all of the carbon in a substance.

3.1.13 *ultimate biodegradation, n*—degradation achieved when the test substance is totally utilized by microorganisms resulting in the production of CO₂, (and possibly methane in the case of anaerobic biodegradation), water, inorganic compounds, and new microbial cellular constituents (biomass or secretions, or both).

3.1.14 *ultimate biodegradation test, n*—a test that estimates the extent to which the carbon in a product has been converted to CO₂ or methane, either directly, by measuring the production of CO₂ or methane, or indirectly, by measuring the consumption of O₂.

3.1.14.1 *Discussion*—The measurement of new biomass is not attempted.

4. Summary of Test Method

4.1 Biodegradation of a lubricant or the component(s) of a lubricant is measured by collecting and measuring the CO₂ produced when the lubricant or component is exposed to microorganisms under controlled aerobic aquatic conditions. This value is then compared to the theoretical amount of CO₂ which could be generated if all of the carbon in the test material were converted to CO₂. CO₂ is a product of aerobic microbial metabolism of carbon-containing substances and so is a direct measure of the test substance's ultimate biodegradation. CO₂ production is quantified by trapping it in a Ba(OH)₂ solution and titrating the solution to calculate the amount of CO₂ absorbed.

4.2 The carbon content of the test substance is determined by Test Method D 5291 or an equivalent method and the theoretical CO₂ is calculated from that measurement. It is necessary to directly measure the carbon content of the test substance instead of calculating this number, because of the complexity of the mixture of compounds present in lubricants.

4.3 Biodegradability is expressed as a percentage of theoretical CO₂ production.

5. Significance and Use

5.1 Results from the test method suggest, within the confines of a controlled laboratory setting, the degree of aerobic aquatic biodegradation of a lubricant or components of a lubricant by measuring the evolved carbon dioxide upon exposure of the test material to an inoculum. The plateau level of CO₂ evolution in this test method will suggest the degree of biodegradability of the lubricant. Test substances that achieve a high degree of biodegradation in this test may be assumed to easily biodegrade in many aerobic aquatic environments.

5.2 Because of the stringency of this test, a low yield of CO₂ does not necessarily mean that the test substance is not biodegradable under environmental conditions, but indicates that further testing is necessary to establish biodegradability.

5.3 Information on toxicity to the inoculum of the test substance may be useful in the interpretation of low biodegradation results.

5.4 Activated sewage-sludge from a sewage-treatment plant that principally treats domestic waste is considered an acceptable active aerobic inoculum available over a wide geographical area in which to test a broad range of lubricants. An inoculum derived from soil or natural surface waters, or both, or any combination of the three sources, is also appropriate for this test method.

NOTE 1—Allowance for various and multiple inoculum sources provides access to a greater diversity of biochemical competency and potentially represents more accurately the capacity for biodegradation.

5.5 A reference or control substance known to biodegrade is necessary in order to verify the activity of the inoculum. The test must be regarded as invalid and should be repeated using a fresh inoculum if the reference does not demonstrate a biodegradation of > 60 % of the theoretical CO₂ evolution within 28 days.

5.6 A total CO₂ evolution in the blank at the end of the test exceeding 50 mg CO₂ per 3 L of medium should be considered as invalidating the test.

5.7 The water solubility or dispersibility of the lubricant or component may influence the results obtained and hence the procedure may be limited to comparing lubricants or components with similar solubilities.

5.8 The ratio of carbon incorporated into cellular material to carbon released as CO₂ will vary depending on the organic substrate, on the particular microorganisms carrying out the conversion, and on the environmental conditions under which the conversion takes place. In principle, this variability complicates the interpretation of the results from this test method.

6. Apparatus

6.1 *Carbon Dioxide Scrubbing Apparatus*—(see Fig. 1):

6.1.1 The following are required to produce a stream of CO₂-free air of sufficient volume to test up to three materials and the accompanying reference and blank controls in triplicate:

6.1.1.1 *Five 1-L plastic bottles*, containing 700 mL of 10-M sodium hydroxide (NaOH),

6.1.1.2 *Two empty 1-L Erlenmeyer flasks*, to prevent liquid carryover, and

6.1.1.3 *One 1-L Erlenmeyer flask*, containing 700 mL of 0.0125 M barium hydroxide [Ba(OH)₂] solution.

6.1.2 Connect the bottles in series, as shown in Fig. 1, using vinyl, or other suitable non gas-permeable tubing, to a pressurized air system, and purge air through the scrubbing solution at a constant rate.

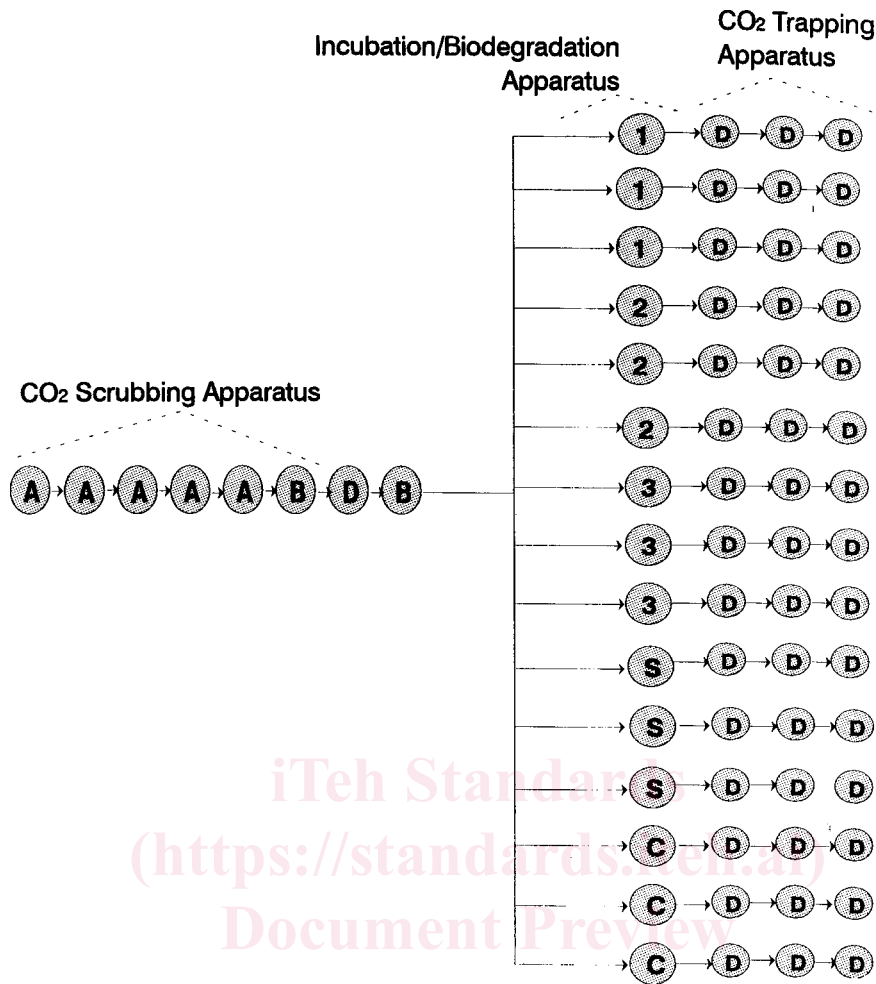
6.1.3 For each additional test substance to be tested, add one additional 1-L plastic bottle filled with 700 mL of 10 M sodium hydroxide.

6.1.4 The CO₂ scrubbing apparatus upstream of the Erlenmeyer flask containing the Ba(OH)₂ solution may be replaced by an alternative system which effectively and consistently produces CO₂ free air (that is, containing less than 1 ppm CO₂).

6.2 *Incubation/Biodegradation Apparatus*—Each test material, reference, or control requires the following:

6.2.1 *Three 4-L Erlenmeyer flasks*,

6.2.2 *Stoppers*, which are non-permeable to CO₂.



- A = NaOH
- B = Empty
- C = Blank
- S = Standard
- D = Ba(OH)₂
- 1 = Test substance 1
- 2 = Test substance 2
- 3 = Test substance 3

FIG. 1 Aerobic Aquatic Biodegradation Testing Schematic

6.2.3 *Flexible Plastic Tubing*, which is non-permeable to CO₂.

6.2.4 *Agitators or Stirrers*, for each 4-L Erlenmeyer flask.

6.3 *Analytical Balance*, to weigh out test material or reference material before or as adding to the test flask,

6.4 *Trapping Apparatus for Measuring Production of CO₂*—For each incubation apparatus, the following are required:

6.4.1 *Several 200-mL Bottles*, fitted with gas bubblers and containing 100 mL 0.0125 M Ba(OH)₂ carbon dioxide scrubbing solution.

6.5 *Titration Apparatus for Measuring Production of CO₂*:

6.5.1 *100-mL burette*.

6.6 *Glass Wool*, for filtering the inoculum.

7. Reagents and Materials

7.1 *Purity of Reagents*—Reagent grade chemicals shall be used in all tests. Unless otherwise indicated, it is intended that

all reagents conform to the specifications of the Committee on Analytical Reagents of the American Chemical Society where such specifications are available.⁹ Other grades may be used, provided it is first ascertained that the reagent is of sufficiently high purity to permit its use without lessening the accuracy of the determination.

7.2 *Purity of Water*—Unless otherwise indicated, references to water shall be understood to mean reagent water as defined by Type II of Specification D 1193.

7.3 Prepare the following stock solutions:

7.3.1 *Ammonium Sulfate Solution (40 g/L)*—Dissolve 40 g of ammonium sulfate ((NH₄)₂SO₄) in water and dilute to 1 L.

⁹ *Reagent Chemicals, American Chemical Society Specifications*, American Chemical Society, Washington, DC. For suggestions on the testing of reagents not listed by the American Chemical Society, see *Analar Standards for Laboratory Chemicals*, BDH Ltd., Poole, Dorset, U.K., and the *United States Pharmacopeia and National Formulary*, U.S. Pharmaceutical Convention, Inc. (USPC), Rockville, MD.