
**Starch and derived products — Heavy
metals content —**

Part 3:

Determination of lead content by atomic
absorption spectrometry with electrothermal
atomization

iTeh STANDARD PREVIEW

Amidons, féculés et produits dérivés — Teneur en métaux lourds —

*Partie 3: Détermination de la teneur en plomb par spectrométrie
d'absorption atomique avec atomisation électrothermique*

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Foreword

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Draft International Standards adopted by the technical committees are circulated to the member bodies for voting. Publication as an International Standard requires approval by at least 75 % of the member bodies casting a vote.

International Standard ISO 11212-3 was prepared by Technical Committee ISO/TC 93, *Starch (including derivatives and by-products)*.

ISO 11212 consists of the following parts, under the general title *Starch and derived products — Heavy metals content*: [ISO 11212-3:1997](https://standards.iteh.ai/catalog/standards/sist/d1a2138c-c9c9-4072-9bec-20923510b426/iso-11212-3-1997)

- *Part 1: Determination of arsenic content by atomic absorption spectrometry*
- *Part 2: Determination of mercury content by atomic absorption spectrometry*
- *Part 3: Determination of lead content by atomic absorption spectrometry with electrothermal atomization*
- *Part 4: Determination of cadmium content by atomic absorption spectrometry with electrothermal atomization*

Annex A of this part of ISO 11212 is for information only.

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Starch and derived products — Heavy metals content —

Part 3:

Determination of lead content by atomic absorption spectrometry with electrothermal atomization

1 Scope

This part of ISO 11212 specifies a method for the determination of the lead content of starch, including derivatives and by-products, by atomic absorption spectrometry with electrothermal atomization.

The number of parameters for the procedure involved in the electrothermal atomization is far larger than in flame atomization; it is thus impossible to propose a comprehensive method likely to ensure the attainment of satisfactory results on all types of apparatus currently available. Each analyst should therefore optimize the conditions of use of his/her own apparatus on the basis of general or particular instructions.

2 Definition

ISO 11212-3:1997

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For the purposes of this part of ISO 11212, the following definition applies.

2.1 lead content: Quantity of lead determined in accordance with the conditions specified in this method and expressed as lead (Pb), in micrograms per kilogram of the product as received.

3 Principle

Wet digestion of the organic matrix. Injection of an aliquot portion of digested sample, in the presence of a matrix modifier, into the furnace of an electrothermal atomization atomic absorption spectrometer.

Measurement of the absorbance at a wavelength of 283,3 nm.

Determination of the concentration of lead in the sample by means of a calibration curve.

4 Reagents

Use only reagents of recognized analytical grade and distilled water or water of equivalent purity.

4.1 Nitric acid ($\rho_{20} = 1,38$ g/ml).

4.2 Hydrogen peroxide, 30 % (V/V) solution.

4.3 Matrix modifier, of the following composition:

Ammonium dihydrogen phosphate [(NH ₄)H ₂ PO ₄]	10 g
Distilled water to make up to	1 000 ml

4.4 Lead standard solution, 1 g/l.

Standard solutions are commercially available at this concentration. These solutions may be prepared by weighing and dissolving the salt or metal of known purity.

4.5 Calibration solutions

Before each series of measurements, prepare from the standard lead solution (4.4) at least five calibration solutions covering the range of concentrations to be determined. 100 ml of each calibration solution shall contain 7,5 ml of nitric acid (4.1) and 20 ml of the matrix modifier solution (4.3) if the latter is not distributed by the automatic injection device.

5 Apparatus

All the glassware used shall be previously washed by means of suitable products (such as nitric acid) and rinsed with distilled water to eliminate any trace of lead.

Use ordinary laboratory apparatus and, in particular, the following.

5.1 Digestion apparatus (see figure 1), made of borosilicate glass and consisting of three elements terminating with conical ground joints (5.1.1 to 5.1.3).

5.1.1 Soxhlet extraction tube, of capacity 200 ml, equipped with a stopcock and a lateral tube connected directly to the flask (5.1.3).

5.1.2 Cooling apparatus, 35 cm long, connected to the top of the Soxhlet extraction tube (5.1.1).

5.1.3 Round-bottom flask, of capacity 250 ml, connected to the lower part of the Soxhlet extraction tube (5.1.1).

When the stopcock is open, the device is under reflux; when it is closed, the Soxhlet extraction tube (5.1.1) retains the condensed water and acid vapours.

5.2 Atomic absorption spectrometer, consisting of five elements (5.2.1 to 5.2.5).

5.2.1 High-resolution monochromator, allowing a 0,2 nm bandwidth slit.

5.2.2 Correcting device for non-specific absorption.

5.2.3 Measuring and photoelectric reception device, with a response time not exceeding about 10 ms.

5.2.4 Detector and signal processing system, allowing recording of the maximum and/or integrated absorbance signal.

5.2.5 Lead discharge lamp or lead hollow cathode lamp.

5.3 Electrothermic atomizer

The most widely used atomizer, for which the general conditions of use are suggested, is a graphite tubular furnace placed in the optical axis of the spectrometer, heated by the Joule effect. The furnace shall be maintained in an inert atmosphere to avoid its destruction by oxidation when heated at a high temperature, and shall be equipped with an automatic injection device which is necessary to obtain good repeatability and to reduce the risk of contamination.

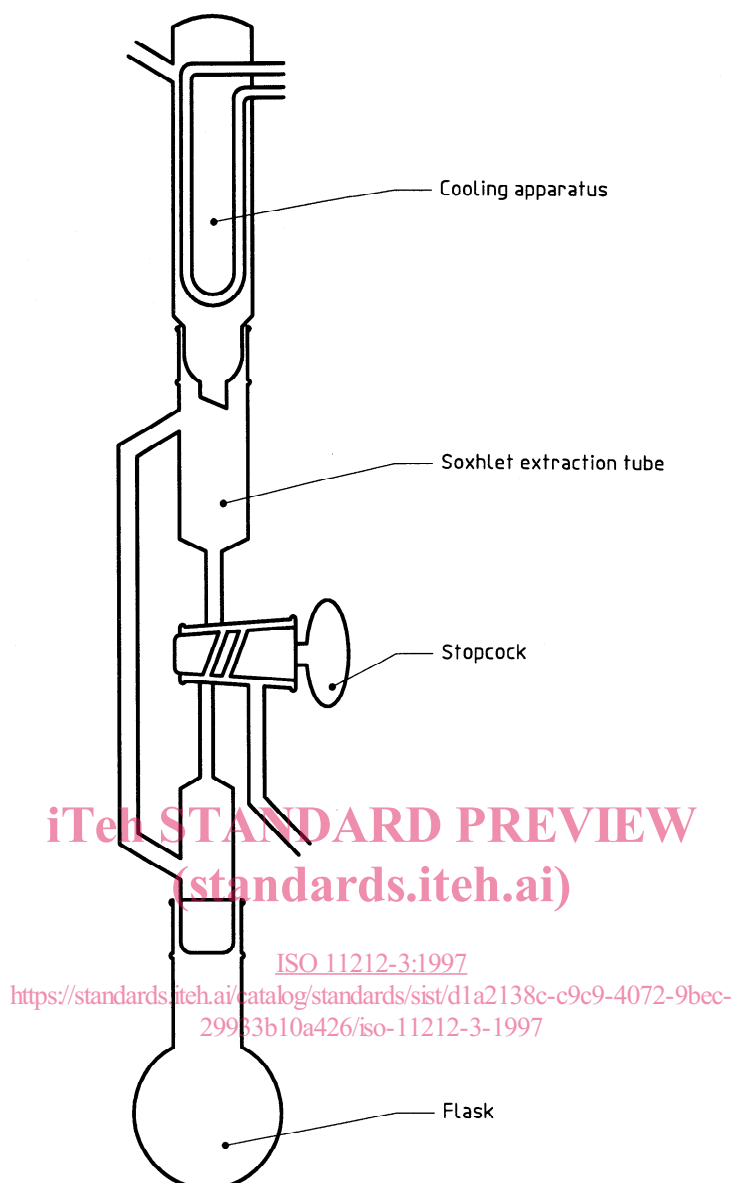


Figure 1 — Digestion apparatus

5.4 Pyrocoated graphite tube, with Lvov platform.

5.5 Pipettes and micropipettes, of suitable capacity.

5.6 Analytical balance.

6 Procedure

6.1 General

To avoid too high a result, it is important to decontaminate the glassware with nitric acid, to rinse it correctly, to prevent any external contamination induced by handling and by the laboratory atmosphere, and to check the purity of the reagents by means of the blank tests described in 6.4.

6.2 Preparation of test sample

Thoroughly homogenize the sample.

6.3 Digestion

Use the digestion apparatus described in 5.1.

Weigh, to the nearest 1 mg, about 5 g of the test sample into the flask (5.1.3). Add 27,5 ml of nitric acid (4.1) and 1 ml of hydrogen peroxide (4.2). Distil under reflux for 4 h leaving the stopcock open. Turn the stopcock off, continue heating and distil until about 20 ml \pm 1 ml of liquid are recovered in the extraction tube (5.1.1). Stop heating and allow the flask to cool. Separate the flask from the extraction tube. Add 20 ml of water to the digested residue in the flask, bring to the boil for a few minutes, stop heating and allow to cool. Transfer the solution to a 100 ml volumetric flask, add 20 ml matrix modifier (4.3) (if it is not distributed by the automatic injection device), dilute to the mark with distilled water and stir.

6.4 Blank test

Perform digestion under the same conditions as in 6.3, replacing the test portion by 5 ml of water.

6.5 Electrothermal atomization programme

The heating programme of the furnace mainly depends on the chemical properties of the substance to be analysed, on the matrix and on the method for approaching the isothermal conditions chosen. It is composed of four stages (6.5.1 to 6.5.4) which shall be optimized by each laboratory.

6.5.1 Drying

It is advisable to increase the temperature slowly up to a final temperature slightly higher than the boiling temperature of the solvent and to maintain this for at least 5 s.

6.5.2 Thermal pretreatment

The temperature for this stage, during which the organic matrix is eliminated and the mineral matrix is modified, shall be adapted by adding a matrix modifier (ammonium dihydrogen phosphate) (4.3) capable of stabilizing the substance by heat.

6.5.3 Atomization

This stage is generally performed with a quick increase in temperature together with no or a reduced flow of gas to ensure a maximum concentration of atoms in the optical path length.

6.5.4 Cleaning of the furnace

Memory effects are possible, therefore the furnace (5.3) shall be cleaned after each injection. Cleaning is generally performed for a few seconds at maximum temperature and gas flow rate.

6.6 Determination of the calibration curve

Inject to the programmed furnace (5.3), 10 μ l of the diluted calibration solution (4.5) and 2 μ l of the matrix modifier (4.3) if the latter is not delivered by the automatic injection device. Measure the absorbance of each calibration solution at a wavelength of 283,3 nm using the spectrometer (5.2).

Draw the calibration curve by plotting the lead concentrations of the calibration solutions, expressed in micrograms per litre, as the abscissa against the corresponding values of the signal, read either in maximum absorbance or in integrated absorbance, as the ordinate. The calibration curve shall be periodically checked depending on the length of the series of analyses.

6.7 Determination

Measure the absorbance of the test samples under the same conditions as the calibration solutions and compare the results with the previously plotted calibration curve.

7 Expression of results

With reference to the calibration curve, determine the concentrations corresponding to the signals of the test portion and the blank. The lead concentration of the sample, w , expressed in micrograms per kilogram of the product as received, is given by the equation:

$$w = \frac{(\rho_1 - \rho_0) \times 100}{m}$$

where

ρ_1 is the numerical value of the lead concentration, in micrograms per litre, of the test solution (6.3) read from the calibration curve (6.6);

ρ_0 is the numerical value of the lead concentration, in micrograms per litre, of the blank test solution (6.4) read from the calibration curve (6.6);

m is the numerical value of the mass, in grams, of the test portion (6.3).

NOTE — When strictly following this method, the quantification limit can reach 50 µg/kg.

8 Precision

The details of an interlaboratory test on the precision of the method are summarized in annex A. The values derived from the interlaboratory test may not be applicable to analyte concentration ranges and matrices other than those given in annex A.

8.1 Repeatability

The absolute difference between two independent single test results, obtained using the same method on identical test material in the same laboratory by the same operator using the same equipment within a short interval of time, will not exceed the repeatability limit r deduced from table A.1 in more than 5 % of cases.

8.2 Reproducibility

The absolute difference between two single test results, obtained using the same method on identical test material in different laboratories with different operators using different equipment, will not exceed the reproducibility limit R deduced from table A.1 in more than 5 % of cases.

9 Test report

The test report shall specify:

- the method in accordance with which sampling was carried out, if known;
- the method used;
- the test result(s) obtained; and
- if the repeatability has been checked, the final quoted result obtained.

It shall also mention all operating details not specified in this part of ISO 11212, or regarded as optional together with details of any incidents which may have influenced the test result(s).

The test report shall include all information necessary for the complete identification of the sample.

Annex A (informative)

Results of interlaboratory test

An interlaboratory test at the international level was carried out by 12 laboratories in 1993. The statistical results indicated in table A.1 were determined in accordance with ISO 5725¹⁾.

Table A.1 — Interlaboratory test on corn starch

Parameter	Sample ¹⁾		
	LC	HC	VHC
No. of laboratories retained after eliminating outliers	8	10	9
No. of outliers (laboratories)	6	5	6
No. of accepted results	32	39	36
Mean lead content (µg/kg)	108,7	169,4	289,2
Repeatability standard deviation, s_r (µg/kg)	12,6	14,1	24,9
Repeatability limit, $r = 2,8 \times s_r$ (µg/kg)	35,6	39,9	70,3
Reproducibility standard deviation, s_R (µg/kg)	64,8	53,0	72,3
Reproducibility limit, $R = 2,8 \times s_R$ (µg/kg)	183,4	150,0	204,6
1) Corn starch with LC: low content HC: high content VHC: very high content			

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1) ISO 5725:1986, *Precision of test methods — Determination of repeatability and reproducibility for a standard test method by inter-laboratory tests* (now withdrawn), was used to obtain the precision data.

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