Designation: F1926/F1926M – 08

Standard Test Method for Evaluation of the Environmental Stability of Calcium Phosphate Granules, Fabricated Forms, and Coatings[1]

This standard is issued under the fixed designation F1926/F1926M; the number immediately following the designation indicates the year of original adoption or, in the case of revision, the year of last revision. A number in parentheses indicates the year of last reapproval. A superscript epsilon (ε) indicates an editorial change since the last revision or reapproval.

1. Scope

1.1 This test method covers calcium phosphate materials intended for use in surgical implant applications.

1.2 Aspects of the biological response to calcium phosphate materials in soft tissue and bone have been reported from laboratory studies and clinical use (**1-10**).[2]

1.3 The requirements of this specification apply to calcium phosphate materials such as calcium hydroxyapatite (see Specification F1185), beta-tricalcium phosphate (see Specification F1088), and biphasic mixtures thereof with or without intentional addition of other minor components (<10 %).

1.4 The material(s) shall be representative of that produced for sale. It shall have been produced and processed under standard manufacturing conditions.

1.5 The materials may be in the form of powders, granules, fabricated forms or coatings; and may be porous, nonporous, textured, and other implantable topographical substrate form representative of the end-use product.

1.6 The calcium phosphate material may constitute the only material in a substrate or it may be one of multiple materials so long as all other materials present do not dissolve under the test conditions described in this test method.

1.7 This test method is limited to the laboratory evaluation of the dissolution rate of a calcium phosphate material. No correlation of the results to *in vivo* performance is implied.

1.8 The values stated in either SI units or inch-pound units are to be regarded separately as standard. The values stated in each system may not be exact equivalents; therefore, each system shall be used independently of the other. Combining values from the two systems may result in non-conformance with the standard.

1.9 *This standard does not purport to address all of the safety concerns, if any, associated with its use. It is the responsibility of the user of this standard to establish appropriate safety and health practices and determine the applicability of regulatory limitations prior to use.*

2. Referenced Documents

2.1 *ASTM Standards:*[3]

E691 Practice for Conducting an Interlaboratory Study to Determine the Precision of a Test Method

F1088 Specification for Beta-Tricalcium Phosphate for Surgical Implantation

F1185 Specification for Composition of Hydroxylapatite for Surgical Implants

3. Terminology

3.1 *Definitions of Terms Specific to This Standard:*

3.1.1 *calcium phosphate*—any one of a number of inorganic chemical compounds containing calcium and phosphate ions as its principal constituents.

3.1.2 *coating*—a layer of material mechanically or chemically adhering to the surface of a substrate.

4. Dissolution Media

4.1 Water used for preparing reagents or dissolution media shall be degassed carbon dioxide free deionized or distilled water and have less than 0.1 ppm of residual Ca^{++} ion.

4.2 *Unbuffered Water Media*—Deionized or distilled water containing 8×10^{-5} M NaCl, 8×10^{-5} M $CaCl_2$, and 5×10^{-5} M $K_3(PO_4)$.

4.3 *pH 5.5 MES Buffer Media*—1.0 M MES, [2-(N-morphplino)ethanesulfonic acid] having a pH of 5.5 at 37 ± 0.5°C and containing 8×10^{-5} M NaCl, 8×10^{-5} M $CaCl_2$, and 5×10^{-5} M $K_3(PO_4)$.

4.3.1 A buffer concentration of 1.0 M will usually provide sufficient buffer capacity to keep the solution within ±0.1 pH units of the initial value. If this is not the case, the buffer capacity should be adjusted accordingly.

[1] This test method is under the jurisdiction of ASTM Committee F04 onMedical and Surgical Materials and Devices and is the direct responsibility of Subcommittee F04.13 on Ceramic Materials.

Current edition approved Oct. 1, 2008. Published October 2008. Originally published in 1998. Last previous edition approved in 2003 as F1926 – 03. DOI: 10.1520/F1926_F1926M-08.

[2] The boldface numbers given in parentheses refer to a list of references at the end of the text.

[3] For referenced ASTM standards, visit the ASTM website, www.astm.org, or contact ASTM Customer Service at service@astm.org. For *Annual Book of ASTM Standards* volume information, refer to the standard's Document Summary page on the ASTM website.

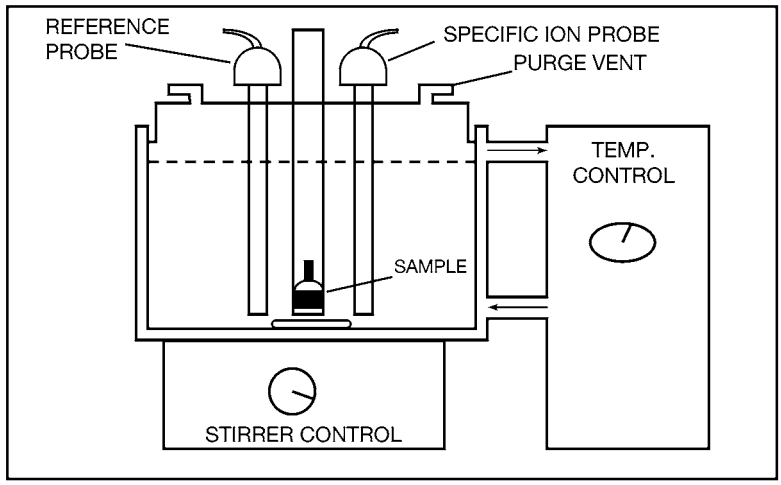

FIG. 1 Dissolution Apparatus

4.3.2 The pH must be adjusted to 5.5 at 37 ± 0.5°C using HCl or NaOH solutions.

4.4 *pH 7.4 TRIS Buffer Media*—1.0 M TRIS, [Tris(hydroxymethyl)aminomethane] having a pH of 7.4 at 37 ± 0.5°C and containing 8×10^{-5} M NaCl, 8×10^{-5} M $CaCl_2$, and 5×10^{-5} M $K_3(PO_4)$.

4.4.1 A buffer concentration of 1.0 M will usually provide sufficient buffer capacity to keep the solution within ± 0.1 pH units of the initial value. If this is not the case, the buffer capacity should be adjusted accordingly.

4.4.2 The pH must be adjusted to 7.4 at 37 ± 0.5°C using HCl or NaOH solutions.

5. Analytical Parameters

5.1 The following procedure should be performed with each of the media listed:

5.1.1 The dissolution rate shall be measured under the conditions of a constant ratio of initial material mass (mg) to total dissolution media volume (mL). The ratio of test material mass to dissolution media shall typically be between 0.005 and 0.01 mg/mL.

5.1.2 The dissolved Ca++ concentration (±1 ppm) shall be measured as soon as practical after the start of the experiment and at appropriate time intervals thereafter to allow the determination of their changes with time.

6. Analytical Procedures

6.1 Make pH measurements with an appropriately calibrated pH meter and probe.

6.2 Measure the Ca++ concentrations potentiometrically. Other methods (for example, colorimetrically, atomic absorption (AA), inductively coupled plasma (ICP) spectroscopy, or inductively coupled plasma mass spectroscopy (ICP/MS)) may be used if equivalency can be demonstrated.

6.3 An appropriate bacteriostat (for example, 0.1 v/v % Hibiclens or 0.1 w/v % sodium azide) may be added to the dissolution media prior to the start of an experiment.

7. Dissolution Apparatus

7.1 The dissolution vessel shall be of such design to easily accommodate the test specimen, the magnetic stirrer bar, and the specific ion-electrode and reference electrode assemblies. It shall also be isolated from the atmosphere by an oxygen and carbon dioxide free inert gas purge.

7.1.1 A convenient apparatus (see Fig. 1) is a 100 mL jacketed beaker with circulating water from a thermostatically controlled vessel. A flat piece of polyethylene, or other inert plastic, with appropriate holes drilled to accommodate the probes, sample holder, and purge gas tube can serve as a lid.

7.2 It shall be of appropriate dimensions to contain the required volume of dissolution media at a level to keep the test material completely submerged during the test and facilitate sufficient stirring action from the magnetic stirrer bar.

7.3 The stirrer assembly shall be capable of maintaining a constant stirring rate of 100 ± 20 rpm.

7.3.1 *Magnetic stirrer bar*—(0.31 in. (8 mm) diameter, 2 in. (51 mm) length, polytetrafluoroethylene (PTFE)-coated).

7.3.2 A different type of stirrer design and stirring rate may be used provided equivalence in experimental results can be demonstrated.

7.4 The dissolution vessel shall be thermostatically controlled at 37 ± 0.5 °C.

7.5 The dissolution apparatus may include various data recording and storage devices, strip chart recorders, computers, and so forth, to facilitate continuous monitoring throughout the duration of the experiment.

8. Preparation of Test Specimens

8.1 *Coatings*:

8.1.1 The standard test specimen for evaluating coating materials is defined in Fig. 2.

8.1.2 The test specimen shall be manufactured from the same materials and processes as substrates produced for sale.

8.1.3 The test specimen shall have the upper shank threaded so as to mate with an appropriate supporting shaft as described in Fig. 1.

8.1.4 By appropriate masking, or other techniques, the coating shall be applied only to the central 0.5 ± 0.005 in. of the test specimen.

8.1.5 The coating shall be applied to the test specimen and receive the same processing steps as the actual product.