

SLOVENSKI STANDARD SIST EN 113-2:2021

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Nadomešča: SIST EN 113:2002 SIST EN 113:2002/A1:2004 SIST-TS CEN/TS 15083-1:2006

Trajnost lesa in lesnih proizvodov - Preskusna metoda proti glivam prostotrosnicam - 2. del: Ocenjevanje naravne ali izboljšane odpornosti

Durability of wood and wood-based products - Test method against wood destroying basidiomycetes - Part 2: Assessment of inherent or enhanced durability

Dauerhaftigkeit von Holz und Holzprodukten - Prüfverfahren in Bezug auf Holz zerstörende Basidiomyceten - Teil 2: <u>Bewertung der</u> natürlichen oder verbesserten Dauerhaftigkeit <u>https://standards.iteh.ai/catalog/standards/sist/8fe0fea3-32d9-4952-afdc-</u>

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Durabilité du bois et des matériaux dérivés du bois - Méthode d'essai vis-à-vis des champignons basidiomycètes - Partie 2 : Détermination de la durabilité inhérente ou améliorée

Ta slovenski standard je istoveten z: EN 113-2:2020

ICS:

71.100.50 Kemikalije za zaščito lesa Wood-protecting chemicals

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EUROPEAN STANDARD NORME EUROPÉENNE EUROPÄISCHE NORM

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English Version

Durability of wood and wood-based products - Test method against wood destroying basidiomycetes - Part 2: Assessment of inherent or enhanced durability

Durabilité du bois et des matériaux dérivés du bois -Méthode d'essai vis-à-vis des champignons basidiomycètes - Partie 2 : Détermination de la durabilité inhérente ou améliorée Dauerhaftigkeit von Holz und Holzprodukten -Prüfverfahren gegen Holz zerstörende Basidiomyceten - Teil 2: Bewertung der natürlichen oder verbesserten Dauerhaftigkeit

This European Standard was approved by CEN on 2 November 2020.

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EUROPEAN COMMITTEE FOR STANDARDIZATION COMITÉ EUROPÉEN DE NORMALISATION EUROPÄISCHES KOMITEE FÜR NORMUNG

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SIST EN 113-2:2021

EN 113-2:2020 (E)

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European foreword

This document (EN 113-2:2020) has been prepared by Technical Committee CEN/TC 38 "Durability of wood and wood-based products", the secretariat of which is held by AFNOR.

This European Standard shall be given the status of a national standard, either by publication of an identical text or by endorsement, at the latest by May 2021, and conflicting national standards shall be withdrawn at the latest by May 2021.

Attention is drawn to the possibility that some of the elements of this document may be the subject of patent rights. CEN shall not be held responsible for identifying any or all such patent rights.

This document supersedes CEN/TS 15083-1:2005, EN 113:1996, EN 113:1996/A1:2004.

The different parts of EN 113 deal with similar testing but relate to a different scope.

In comparison with CEN/TS 15083-1:2005 EN 113:1996, EN 113:1996/A1:2004, the following significant changes have been made:

- change of the title;
- the obligatory fungi are indicated differently;
- the methods for sterilization are updated, NDARD PREVIEW
- all annexes are informative except Annex B.

According to the CEN-CENELEC Internal Regulations, the national standards organisations of the following countries are bound to implement this European Standard: Austria, Belgium, Bulgaria, Croatia, Cyprus, Czech Republic, Denmark, Estonia, Finland, France, Germany, Greece, Hungary, Iceland, Ireland, Italy, Latvia, Lithuania, Luxembourg, Malta, Netherlands, Norway, Poland, Portugal, Republic of North Macedonia, Romania, Serbia, Slovakia, Slovenia, Spain, Sweden, Switzerland, Turkey and the United Kingdom.

Introduction

This document describes a laboratory method of test which gives a basis for the assessment of the biological durability of a sample of wood or wood product against attack by wood-destroying basidiomycetes. Specifically the natural durability of a wood species can vary depending on the conditions of growth such as climate and soil type. For this reason, the durability established using the method described in this document will relate only to the sample of timber tested. Guidance on sampling is given in Annex A.

This laboratory method provides one criterion by which the durability of the timber can be assessed. It is recommended that this information is supplemented by data from other relevant tests, and above all by practical experience.

The procedures described in this standard method are intended to be carried out by suitably trained or supervised specialists.

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1 Scope

This document specifies a method of test for determining the natural durability of a timber against wooddestroying basidiomycetes cultured on a malt extract agar medium. The method is applicable to all timber species.

Furthermore this method can be used to test modified wood. The test method described in this document can be applied to specific wood species, commercial supplies of sawn timber, wood-based materials, wood treated with preservatives and modified wood, both thermally and chemically modified wood.

However, this document is not intended to determine the effectiveness of wood preservatives used to prevent decay.

NOTE 1 Determining the efficacy of wood preservatives used to prevent decay is the scope of EN 113-1. However, in addition to this and with some amendments, it might also be possible in some cases to test treated wood using the method described here.

NOTE 2 This method can be used in conjunction with an ageing procedure, for example EN 73 or EN 84.

Annex A (informative) contains a guidance on sampling.

Annex B (normative) contains some methods of sterilization.

Annex C (informative) contains information on the culture vessels.

Annex D (informative) contains an example of a test report.

Annex E (informative) contains information on the test fungi. **PREVIEW**

Annex F (informative) contains the assessment of the results en ai)

2 Normative references

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There are no normative references in this documento/sist-en-113-2-2021

3 Terms and definitions

For the purposes of this document, the following terms and definitions apply.

ISO and IEC maintain terminological databases for use in standardization at the following addresses:

— IEC Electropedia: available at http://www.electropedia.org/

— ISO Online browsing platform: available at https://www.iso.org/obp

3.1

supplier

sponsor of the test, person or company providing the sample of wood/timber to be tested

4 Principle

Test specimens of the timber under test and reference timber test specimens are exposed to attack by pure cultures of basidiomycetes. After a prescribed period of incubation under defined conditions, the percentage loss in dry mass of the test specimens is used to establish the timber durability classification of the timber under test.

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5 Test material and apparatus

5.1 Biological material

5.1.1 Test fungi

5.1.1.1 General

The test fungi to be used are as follows and relate to corresponding virulence control specimens.

It is required that the reference timber virulence control specimens provide a median mass loss of at least 30 % with one of the test fungi (see also 5.1.2.5 and 7.4.2).

NOTE Besides obligatory fungi also other test fungi can be used (see Annex E).

5.1.1.2 Test fungi for hardwoods with beech as virulence control wood

Obligatory fungi:

— Coniophora puteana (Schumacher ex Fries) Karsten (BAM Ebw. 15).

Loss in mass of beech in 16 weeks: minimum 20 %.

— *Trametes versicolor* (L.) Lloyd (CTB 863A).

Loss in mass of beech in 16 weeks: minimum 20 %.

5.1.1.3 Test fungi for softwoods with Scots pine sapwood as virulence control wood (standards.iteh.ai)

Obligatory fungi

 Coniophora puteana (Schumacher ex Fries) Karsten (BAM Ebw. 15). https://standards.iteh.ai/catalog/standards/sist/8fe0fea3-32d9-4952-afdc-

Loss in mass of Scots pine sapwood in 16 weeks: minimum 20 %.

- Rhodonia placenta (Fr.) Niemelä, K.H. Larss. and Schigel (FPRL 280).

Loss in mass of Scots pine sapwood in 16 weeks: minimum 20 %.

5.1.1.4 Test fungi for modified wood with both beech and Scots pine sapwood as virulence control wood

Obligatory fungi:

- Coniophora puteana (Schumacher ex Fries) Karsten (BAM Ebw. 15) for both softwoods and hardwoods

Loss in mass of Scots pine sapwood in 16 weeks: minimum 20 %.

Loss in mass of beech in 16 weeks: minimum 20 %.

— Rhodonia placenta (Fr.) Niemelä, K.H. Larss. and Schigel (FPRL 280) for softwoods.

Loss in mass of Scots pine sapwood in 16 weeks: minimum 20 %.

— Trametes versicolor (L.) Lloyd (CTB 863A) for hardwoods.

Loss in mass of beech in 16 weeks: minimum 20 %.

5.1.1.5 Maintenance of fungal strains

The strains shall be maintained and treated so that its virulence is conserved and ensured (see Annex E).

The strains shall be maintained and treated (frequency of subculturing, alternation of culture media, etc.) in accordance with the instructions of their laboratory of origin (see E.2).

NOTE The parent strain is maintained in the laboratory of its origin so as to conserve and to ensure its vigour.

If tests are not undertaken regularly or if a strain shows signs of degeneration a new standard culture of the strain should be obtained from the laboratory of its origin for each test (see E.2).

When new strains are received, the virulence shall be tested to ensure the strain can achieve the minimum loss in mass (see 5.1.1.2 and 5.1.1.3).

5.1.2 Wood as virulence control

5.1.2.1 Species used for the tests

- Scots pine sapwood (*Pinus sylvestris* Linnaeus) for tests with softwoods;
- Beech (Fagus sylvatica Linnaeus) for tests with hardwoods;
- For modified wood both Scots pine sapwood and beech should be used.

5.1.2.2 Wood quality

The wood shall be free from cracks, stain, decay, insect damage or other defects. The wood shall not have been water-stored, floated, chemically treated or steamed.

Wood that has been kiln dried at temperatures not above 60 °C can be used. NOTE

The Scots pine shall be exclusively sapwood containing little resin and having between 2,5 and 8 annual growth rings per 10 mm. The proportion of latewood in the annual rings shall not exceed 30 % of the whole.

https://standards.iteh.ai/catalog/standards/sist/8fe0fea3-32d9-4952-afdc-The beech shall be even-grained, free from tyloses_discolouration_and red heart. It shall have between 2 and 6 annual growth rings per 10 mm.

5.1.2.3 Provision of virulence control specimens

Prepare planed strips having a cross-section of $(25 \pm 0,5)$ mm x $(15 \pm 0,5)$ mm. The longitudinal faces shall be parallel to the direction of the grain. The annual rings shall not be parallel to the broad faces (contact angle to be greater than 5°) but otherwise may run in any direction. Make transverse cuts, neatly to give sharp edges and a fine-sawn finish to the end-grain surfaces, to give virulence control specimens $(50 \pm 0,5)$ mm long.

The virulence control specimens shall originate from a minimum of three trees or shall be taken at random from a stock originally of more than 500 test specimens and originating from at least five planks.

5.1.2.4 Dimensions and density of virulence control specimens

The dimensions of each reference timber virulence control specimen at (12 ± 2) % moisture content or conditioned at (65 ± 5) % RH and (20 ± 2) °C for at least 2 weeks, shall be (50 ± 0.5) mm $x (25 \pm 0.5) \text{ mm } x (15 \pm 0.5) \text{ mm}.$

NOTE A two-prong electrical conductivity moisture meter is suitable for assessing moisture content.

In a batch of virulence control specimens, the density of an individual is permitted to differ from the mean value of the batch by ± 10 %.

5.1.2.5 Number and distribution of virulence control specimens

Use at least 10 reference timber virulence control specimens for each test fungus. Mark each reference timber virulence control specimen so that it can be identified throughout the test.

5.2 Products and reagents

The culture medium shall be a malt agar medium with the following composition:

- malt extract: in concentrated form: (50 ± 0.5) g; in powder form: (40 ± 0.5) g;
- agar causing no inhibition of growth of fungi: $(20 \pm 0,5)$ g to $(30 \pm 0,5)$ g;
- de-ionized water; quantity to make up to 1000 g.

Preferably use water conforming to grade 3 of ISO 3696.

Prepare this medium by warming the mixture in a boiling water bath or steam bath, stirring until completely dissolved.

Place in each culture vessel (5.3.1) a sufficient quantity of the medium to provide a minimum depth of 3 mm to 4 mm when in its in-use position. Close the vessels as specified in 5.3.1 and sterilize in an autoclave at $121 \degree$ C for 20 min. Let the vessels cool in their in-use position.

5.3 Apparatus

5.3.1 Culture vessels iTeh STANDARD PREVIEW

Kolle flasks or equivalent vessels with a capacity of between 400 ml and 650 ml providing a flat surface area of between 85 cm² and 120 cm² for the medium and close with a material that allows for air exchange.

https://standards.iteh.ai/catalog/standards/sist/8fe0fea3-32d9-4952-afdc-NOTE 1 Examples of suitable vessels/2afe/given/in/Annex/G-2-2021

NOTE 2 Kolle flasks are usually plugged with a wad of cotton wool. Other culture vessels are usually fitted with leakproof lids, the centres of which are pierced with a round hole of up to 15 mm diameter and plugged with a wad of cotton wool.

5.3.2 Drying oven

Capable of being controlled at (103 ± 2) °C.

5.3.3 Desiccators

With efficient desiccant (silica gel for example).

5.3.4 Conditioning chamber

Well ventilated and controlled at (20 ± 2) °C and (65 ± 5) % relative humidity.

5.3.5 Culture chamber

Incubator or room, dark and controlled at (22 \pm 2) °C and (70 \pm 5) % relative humidity.

5.3.6 Test specimens supports

Made of glass, stainless steel or any other inert material, that is to say, with no risk of having any effect on the culture medium, the fungus, the wood, or of modifying itself. The supports are used to prevent direct contact of the specimens with the culture medium, but shall not separate them from it by more than 3 mm. If abnormally high moisture contents (see 7.4.1) are experienced consistently, use of specimen supports of approximately 5 mm thick could help to control the problem. If thicker specimen supports are used, this should be recorded in the test report. Abnormally high moisture contents are those values of final moisture content that are a mass fraction greater than 100 %.

NOTE Supports might be capable of holding either one or two test specimens.

5.3.7 Equipment for steam sterilization or access to a radiation source

See Annex B.

5.3.8 Ordinary laboratory equipment

Including a balance capable of weighing to an accuracy of 0,01 g.

6 Test specimens

6.1 Species and source of wood

Ensure that the species of each plank or log to be tested has been identified correctly and record both the botanical and the trade name. Obtain as much information as possible on the origin and history of the sample (see Annex A). The sample of timber shall be free from penetrating wood preservative treatments, for example anti-stain products.

NOTE 1 Commercial samples of timber can contain more than one botanical species. Agreed wood species names (e.g. from ATIBT) can be mentioned as well as one can make reference to specific codes used, e.g. according to EN 13556.

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 NOTE 2
 Guidance on sampling is given in Annex A. Similar sampling as for natural durability is feasible for modified wood.

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6.2 Wood quality

The following requirements are specific for wood to be tested in realtion to natural durability and as such valid only for non-modified wood.

Record the physical characteristics of the timber sample, for example the sizes of logs/planks, the presence of resin pockets, cross-grain, knots, sapwood and where possible record the widths of annual rings and the proportion of latewood. For logs, record the position in the trunk if known.

The wood shall be free from cracks, stain, decay, insect damage or other defects. The wood shall not have been water-stored, floated, chemically treated or steamed.

NOTE 1 Wood that has been kiln dried at temperatures below 60°C can be used.

NOTE 2 There are no special requirements for modified wood.

6.3 Provision of the test specimens

Reject at least the outer 10 mm from lateral faces of planks and 50 mm from the end grain; reject at least 50 mm from the end grain of logs.

Condition the wood to a mass fraction of (12 ± 2) % moisture content or conditioned at (65 ± 5) % RH and (20 ± 2) °C for at least 2 weeks. Prepare planed strips having a cross-section of $(25 \pm 0,5)$ mm x $(15 \pm 0,5)$ mm which avoid all obvious defects and which are entirely heartwood or entirely sapwood. The longitudinal faces shall be parallel to the direction of the grain. The annual rings shall not be parallel to the broad faces (contact angle to be greater than 5°) but otherwise may run in any