



Designation: D 3686 – 95 (Reapproved 2001)<sup>ε1</sup>

## Standard Practice for Sampling Atmospheres to Collect Organic Compound Vapors (Activated Charcoal Tube Adsorption Method)<sup>1</sup>

This standard is issued under the fixed designation D 3686; the number immediately following the designation indicates the year of original adoption or, in the case of revision, the year of last revision. A number in parentheses indicates the year of last reapproval. A superscript epsilon (ε) indicates an editorial change since the last revision or reapproval.

<sup>ε1</sup> NOTE—SI statement was added and part of Footnote F of Table A1.1 was deleted editorially in October 2001.

### 1. Scope

1.1 This practice covers a method for the sampling of atmospheres for determining the presence of certain organic vapors by means of adsorption on activated charcoal using a charcoal tube and a small portable sampling pump worn by a worker. A list of some of the organic chemical vapors that can be sampled by this practice is provided in **Annex A1**. This list is presented as a guide and should not be considered as absolute or complete.

1.2 This practice does not cover any method of sampling that requires special impregnation of activated charcoal or other adsorption media.

1.3 The values stated in SI units are to be regarded as the standard.

1.4 *This standard does not purport to address all of the safety concerns, if any, associated with its use. It is the responsibility of the user of this standard to establish appropriate safety and health practices and determine the applicability of regulatory limitations prior to use.* A specific safety precaution is given in **9.4**.

### 2. Referenced Documents

#### 2.1 ASTM Standards:

**D 1356** Terminology Relating to Sampling and Analysis of Atmospheres<sup>2</sup>

**D 3687** Practice for Analysis of Organic Compound Vapors Collected by the Activated Charcoal Tube Adsorption Method<sup>2</sup>

#### 2.2 NIOSH Standard:

**CDC-99-74-45** Documentation of NIOSH Validation Tests<sup>3</sup>

**HSM-99-71-31** Personnel Sampler Pump for Charcoal

Tubes; Final Report<sup>3</sup>

#### 2.3 OSHA Standard:

**CFR 1910** General Industrial OSHA Safety and Health Standard<sup>4</sup>

### 3. Terminology

3.1 For definitions of terms used in this method, refer to Terminology **D 1356**.

3.2 Activated charcoal refers to properly conditioned coconut-shell charcoal.

### 4. Summary of Practice

4.1 Air samples are collected for organic vapor analysis by aspirating air at a known rate through sampling tubes containing activated charcoal, which adsorbs the vapors.

4.2 Instructions are given to enable the laboratory personnel to assemble charcoal tubes suitable for sampling purposes.

4.3 Instructions are given for calibration of the low flow-rate sampling pumps required in this practice.

4.4 Information on the correct use of sampling devices is presented.

4.5 Practice **D 3687** describes a practice for the analysis of these samples.

### 5. Significance and Use

5.1 Promulgations by the Federal Occupational Safety and Health Administration (OSHA) in 29 **CFR 1910.1000** designate that certain organic compounds must not be present in workplace atmospheres at concentrations above specific values.

5.2 This practice, when used in conjunction with Practice **D 3687**, will provide the needed accuracy and precision in the determination of airborne time-weighted average concentrations of many of the organic chemicals given in 29 **CFR 1910.1000**, **CDC-99-74-45** and **HSM-99-71-31**.

<sup>1</sup> This practice is under the jurisdiction of ASTM Committee D22 on Air Quality and is the direct responsibility of Subcommittee D22.04 on Workplace Atmospheres. Current edition approved January 15, 1995. Published March 1995. Originally published as D 3686 – 78. Last previous edition D 3686 – 89.

<sup>2</sup> *Annual Book of ASTM Standards*, Vol 11.03.

<sup>3</sup> Available from the U.S. Department of Commerce, National Technical Information Service, Port Royal Road, Springfield, VA 22161.

<sup>4</sup> Available from Superintendent of Documents, U.S. Government Printing Office, Washington, DC 20402.

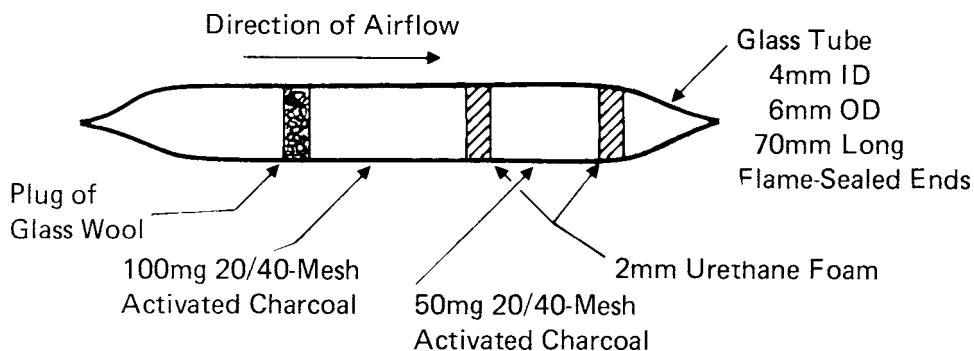


FIG. 1 Activated Charcoal Adsorption Sampling Tube

5.3 A partial list of chemicals for which this method is applicable is given in **Annex A1**, along with their OSHA permissible exposure limits.

## 6. Interferences

6.1 Water mist and vapor can interfere with the collection of organic compound vapors. Humidity greater than 60 % can reduce the adsorptive capacity of activated charcoal to 50 % for some chemicals (1).<sup>5</sup> Presence of condensed water droplets in the sample tube will indicate a suspect sample.

## 7. Apparatus

### 7.1 Charcoal Tube:

7.1.1 A sampling tube consists of a length of glass tubing containing two sections of activated charcoal which are held in place by nonadsorbent material and sealed at each end.

7.1.1.1 Sampling tubes are commercially available. The tubes range in size from 100/50 to 800/400 mg, which means the tubes are divided into two sections with the front section containing 100 to 800 mg of activated charcoal and the back section containing 50 to 400 mg of activated charcoal. The 100/50-mg tube ((2-4) and Fig. 1) which is the one most frequently used, consists of a glass tube, 70-mm long, 6-mm outside diameter, 4-mm inside diameter, and contains two sections of 20/40 mesh-activated charcoal but separated by a 2-mm section of urethane foam. The front section of 100 mg is retained by a plug of glass wool, and the back section of 50 mg is retained by either a second 2-mm portion of urethane foam or a plug of glass wool. Both ends of the tube are flame-sealed.

NOTE 1—Urethane foam is known to adsorb certain pesticides (5), for which this practice is contraindicated.

7.1.1.2 When it is desirable to sample highly volatile compounds for extended periods, or at a high volume flow rate, a larger device capable of efficient collection can be used, provided the proportions of the tube and its charcoal contents are scaled similarly to the base dimensions, to provide nominally the same linear flow rate and contact time with the charcoal bed.

7.1.2 The back portion of the sampler tube, which may contain between 25 and 100 % of the mass of activated charcoal present in the front section, adsorbs vapors that

penetrate the front section and serves as a warning that breakthrough may have occurred. (**Annex A1** gives recommended maximum tube loading information for many chemicals.)

7.1.2.1 Should analysis of the back portion show it to contain more than 10 % of the amount found in the front section, the possibility exists that solvent vapor penetrated both sections of charcoal, and the sample must be considered suspect. These percentages apply to 100/50-mg tubes. For other size tubes having disproportionate amounts of charcoal in the front and back sections, the percentages used to indicate potential breakthrough must be adjusted to take into account different ratios of charcoal. If results from the analysis of suspect samples are used to calculate vapor concentrations, the results must be reported as equal to or greater than the calculated concentrations. In such cases, the test must be repeated for confirmation of vapor concentration.

NOTE 2—Reportings from suspect samples would have significance when health standards are clearly exceeded and the amount by which they are exceeded is academic. (See 9.5.)

7.1.3 The adsorptive capacity and desorption efficiency of different batches of activated charcoal may vary. Commercial tubes, if used, should be purchased from the same batch and in sufficient number to provide sampling capacity for a definite period of time. *Care must be taken to have enough tubes from the same batch for a given study.*

7.1.3.1 The desorption efficiency and contamination level of a batch of tubes should be determined, following the procedure outlined in Practice D 3687 for activated charcoal. A random selection of at least five charcoal tubes from a specified lot should be taken for these checks.

7.1.4 Pressure drop across the sampling tube should be less than 25 mm Hg (3.3 kPa) at a flow rate of 1000 mL/min and less than 4.6 mm Hg (0.61 kPa) at a flow rate of 200 mL/min.

7.1.5 Charcoal sampling tubes prepared in accordance with this practice and with sealed glass ends may be stored indefinitely.

### 7.2 Sampling Pumps:

7.2.1 Any pump whose flow rate can be accurately determined and be set at the desired sampling rate is suitable. Primarily though, this practice is intended for use with small personal sampling pumps.

7.2.2 Pumps having stable low flow rates (10 to 200 mL/min) are preferable for long period sampling (up to 8 h) or

<sup>5</sup> The boldface numbers in parentheses refer to the list of references at the end of this standard.

when the concentration of organic vapors is expected to be high. Reduced sample volumes will prevent exceeding the adsorptive capacity of the charcoal tubes. (Suggested flow rates and sampling times are given in **Annex A1** for anticipated concentration ranges. Sample volumes are also discussed in 9.5.)

7.2.3 Pumps are available that will provide stable flow rates between  $\pm 5\%$ . Pumps should be calibrated before and after sampling. If possible, flow rates should be checked during the course of the sampling procedure.

7.2.4 All sampling pumps must be carefully calibrated with the charcoal tube device in the proper sampling position. (See **Annex A2** for calibration procedure.)

## 8. Reagents

8.1 *Activated Coconut-Shell Charcoal*—Prior to being used to make sampling devices the charcoal should be heated in an inert gas to 600°C and held there for 1 h. Commercially available coconut charcoal (20/40 mesh) has been found to have adequate adsorption capacity. Other charcoals can be used for special applications.

## 9. Sampling with Activated Charcoal Samplers

9.1 *Calibration of the Sampling System*—Calibrate the sampling system, including pump, flow regulator, tubing to be used, and a representative charcoal tube (or an equivalent induced resistance) with a primary or calibrated secondary flow-rate standard to  $\pm 5\%$ .

9.1.1 A primary standard practice is given for the calibration of low flow-rate pumps in **Annex A2** and **Fig. A2.1**.

9.2 Break open both ends of the charcoal tube to be used for sampling, ensuring that each opening is at least one half the inside diameter of the tube.

9.3 Insert the charcoal tube into the sampling line, placing the back-up section nearest to the pump. At no time should there be any tubing ahead of the sampling tubes.

9.4 For a breathing zone sample, fasten the sampling pump to the worker, and attach the sampling tube as close to the worker's breathing zone as possible. Position the tube in a vertical position to avoid channeling of air through the adsorber sections.

**NOTE 3—Warning:** Assure that the presence of the sampling equipment is not a safety hazard to the worker.

9.4.1 Turn on the pump and adjust the flow rate to the recommended sampling rate.

9.4.2 Record the flow rate and starting time or, depending on the make of pump used, the register reading.

9.5 *Sampling Volumes*—The minimum sample volume will be governed by the detection limit of the analytical method, and the maximum sample volume will be determined by either the adsorptive capacity of the charcoal or limitations of the pump battery.

9.5.1 One method of calculating required sample volumes is to determine first the concentration range, over which it is important to report an exact number, for example from 0.2 to 2 times the permissible exposure concentration, and then calculate the sample volumes as follows:

$$\begin{aligned} & \text{Minimum sample volume, m}^3 & (1) \\ & = \frac{\text{minimum detection limit, mg}}{0.2 \times \text{permissible exposure limit, mg/m}^3} \end{aligned}$$

$$\begin{aligned} & \text{Maximum sample volume, m}^3 & (2) \\ & = \frac{\text{tube capacity for vapors, mg}}{2 \times \text{permissible exposure limit, mg/m}^3} \end{aligned}$$

9.5.2 Select a sampling rate that, in the sampling time desired, will result in a sample volume between the minimum and maximum calculated in 9.5.1.

9.5.2.1 Generally a long sampling time at a low flow rate is preferable to short-term high-volume sampling. This is consistent with the fact that most health standards are based on 8-h/day time-weighted averages of exposure concentrations.

9.5.2.2 A sample flow rate of less than 10 mL/min, however, should not be used. Calculations based upon diffusion coefficients for several representative compounds indicate that sampling at less than 10 mL/min may not give accurate results.<sup>6</sup>

9.5.2.3 Approximate sample volumes and sample times are given in **Annex A1**.

9.5.3 When spot checks are being made of an environment, a sample volume of 10 L is adequate for determining vapor concentrations in accordance with exposure guidelines.

9.6 At the end of the sampling period recheck the flow rate, turn off the pump, and record all pertinent information: time, register reading, and if pertinent, temperature, barometric pressure, and relative humidity.

9.6.1 Seal the charcoal tube with the plastic caps provided.

9.6.2 Label the tube with the appropriate information to identify it.

9.7 At least one charcoal sampling tube should be presented for analysis as a field blank with every 10 or 15 samples, or for each specific inspection or field study.

9.7.1 Break the sealed ends off the tube and cap it with the plastic caps. Do not draw air through the tube, but in all other ways treat it as an air sample.

9.7.2 The purpose of the field blank is to assure that if the sampling tubes adsorb vapors extraneous to the sampling atmosphere, the presence of the contaminant will be detected.

9.7.3 Results from the field blanks shall not be used to correct sample results. If a field blank shows contamination, the samples taken during the test must be assumed to be contaminated.

9.8 *Calculation of Sample Volume:*

9.8.1 For sample pumps with flow-rate meters:

$$\text{Sample volume, mL} = f \times t \left( \sqrt{\frac{P_1}{P_2} \times \frac{T_2}{T_1}} \right) \quad (3)$$

where:

$f$  = flow rate sampled, mL/min,

$t$  = sample time, min,

$P_1$  = pressure during calibration of sampling pump (mm Hg or kPa)

<sup>6</sup> Heitbrink, W. A., "Diffusion Effects Under Low Flow Conditions," American Industrial Hygiene Association Journal, Vol 44, No. 6, 1983, pp. 453-462.

$P_2$  = pressure of air sampled (mm Hg or kPa)  
 $T_1$  = temperature during calibration of sampling pump (K),  
 and

$T_2$  = temperature of air sampled (K).

9.8.2 For sample pumps with counters:

$$V = \frac{(R_2 - R_1) \times V}{I} \times \frac{P_1}{760} \times \frac{298}{T_1 + 273} \quad (4)$$

where:

$R_2$  = final counter reading,

$R_1$  = beginning counter reading,

$V$  = volume, (1) mL-count (1)

$P_1$  = barometric pressure, mm Hg,

$T_1$  = temperature, °C, and

$V$  = total sample volume, mL.

## 10. Handling and Shipping of Samples Collected on Charcoal Sampling Tubes

10.1 There is a paucity of information on the possible fate of the many different chemical species that can be collected in activated charcoal and the variety of conditions to which these samples may be exposed. Good practice suggests the following:<sup>7</sup>

<sup>7</sup> Two studies that present information pertinent to this section are:  
 Saalwaechter, A. T., et al, "Performance Testing of the NIOSH Charcoal Tube Technique for the Determination of Air Concentrations of Organic Vapors," *American Industrial Hygiene Association Journal*, Vol 38, No. 9, September 1977, pp. 476-486.  
 Hill, R. H., Jr., et al, "Gas Chromatographic Determination of Vinyl Chloride in Air Samples Collected on Charcoal," *Analytical Chemistry*, Vol 48, No. 9, August 1976, pp. 1395-1398.

10.1.1 Samples should be capped securely and identified clearly.

10.1.2 Samples collected in charcoal tubes should not be kept in warm places or exposed to direct sunlight.

10.1.3 Samples of highly vaporous or low-boiling materials, such as vinyl chloride, should be stored and transported in dry ice.

10.1.4 At present there are no published test data on the effect of conditions in aircraft cargo holds on capped samples. The preferred procedure is to carry the samples on board.

10.1.5 Samples should be shipped as soon as possible, stored under refrigeration until they are analyzed, and analyzed if possible within five working days.

10.1.6 Migration or equilibration of the sampled material within the sampling tube during prolonged or adverse storage or handling could be interpreted as break-through. This can be prevented by separating the front and back sections immediately after sampling, by having each section in a separate tube and capping them separately.

10.1.7 In some situations, circumstances and facilities may permit making up calibration standards at the facility where the study is being made and submitting these standards as quality control checks. (See Practice D 3687 for recommended procedure for making up standards.)

10.1.8 Bulk solvent samples should never be shipped or stored with the collected air samples.

## 11. Keywords

11.1 activated charcoal tube; air monitoring; charcoal tube; organic vapors; sampling and analysis; workplace atmospheres

## ASTM D3686-95 (2001)e1 ANNEXES

<https://standards.iteh.ai/catalog/standards/sist/7c464225-806b-4b72-b945-f89eaa46ccb2/astm-d3686-952001e1>  
 (Mandatory Information)

### A1. INFORMATION OF SOME ORGANIC COMPOUND VAPORS THAT CAN BE COLLECTED ON COCONUT-SHELL CHARCOAL (100/50 mg tubes)

**TABLE A1.1 INFORMATION OF SOME ORGANIC COMPOUND VAPORS THAT CAN BE COLLECTED ON COCONUT-SHELL CHARCOAL (100/50 mg tubes)**

Substance PEL ppm-mg/m <sup>3A</sup>	Recommended Sampling Rate, mL/min to Detect Ap- proximately 15 to 200 % of PEL in Time Given <sup>B</sup>			Recommended Maximum Tube Load- ing, mg <sup>C</sup>	Approximate Desorption Ef- ficiency % <sup>D</sup>	Eluent	GC Column <sup>E</sup>	CV <sub>T</sub> <sup>F</sup>
	2h	4h	8h					
Acetone, 1000-2400	10	<sup>G</sup>	<sup>G</sup>	9	86 ± 10	CS <sub>2</sub>	3	0.082
Acetonitrile, 40-70	50	25	25	2.7				0.072
Allyl alcohol, 2-4.8	200	100	50	<0.4	89 ± 5	CS <sub>2</sub> + 5 % 2-propanol	2	0.11
<i>n</i> -Amyl acetate, 100-525	50	25	10	15	86 ± 5	CS <sub>2</sub>	4	0.051
<i>sec</i> -Amyl acetate, 125-650	50	25	10	15.5	91 ± 10	CS <sub>2</sub>	4	0.071
Isoamyl alcohol, 100-360	50	25	10	10		CS <sub>2</sub> + 5 % 2-propanol	2	0.077
Benzene, 10-31.3	100	100	50		96	CS <sub>2</sub>	1	0.060
Benzyl chloride, 1-5	—	200	200	<0.4	90 ± 5	CS <sub>2</sub>	2	0.096
Butadiene, 1000-2200	10	<sup>G</sup>	<sup>G</sup>	4		CS <sub>2</sub>	1	0.058