
**Plastics — Polyurethane raw materials —
Determination of isocyanate content**

*Plastiques — Matières premières des polyuréthanes — Détermination
de la teneur en isocyanate*

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ISO copyright office
Case postale 56 • CH-1211 Geneva 20
Tel. + 41 22 749 01 11
Fax + 41 22 749 09 47
E-mail copyright@iso.org
Web www.iso.org

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Foreword

ISO (the International Organization for Standardization) is a worldwide federation of national standards bodies (ISO member bodies). The work of preparing International Standards is normally carried out through ISO technical committees. Each member body interested in a subject for which a technical committee has been established has the right to be represented on that committee. International organizations, governmental and non-governmental, in liaison with ISO, also take part in the work. ISO collaborates closely with the International Electrotechnical Commission (IEC) on all matters of electrotechnical standardization.

International Standards are drafted in accordance with the rules given in the ISO/IEC Directives, Part 2.

The main task of technical committees is to prepare International Standards. Draft International Standards adopted by the technical committees are circulated to the member bodies for voting. Publication as an International Standard requires approval by at least 75 % of the member bodies casting a vote.

Attention is drawn to the possibility that some of the elements of this document may be the subject of patent rights. ISO shall not be held responsible for identifying any or all such patent rights.

ISO 14896 was prepared by Technical Committee ISO/TC 61, *Plastics*, Subcommittee SC 12, *Thermosetting materials*.

This third edition cancels and replaces the second edition (ISO 14896:2006), of which it constitutes a minor revision, the main purpose of which was to combine the standard with its amendment (ISO 14896:2006/Amd.1:2007), thereby adding a new subclause (12.1.8).

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Plastics — Polyurethane raw materials — Determination of isocyanate content

SAFETY STATEMENT — Persons using this document should be familiar with normal laboratory practice, if applicable. This document does not purport to address all of the safety concerns, if any, associated with its use. It is the responsibility of the user to establish appropriate safety and health practices and to ensure compliance with any regulatory requirements.

1 Scope

This International Standard specifies two methods for the measurement of the isocyanate content of aromatic isocyanates used as polyurethane raw materials. Method A is primarily applicable to refined toluene diisocyanate (TDI), methylene-bis-(4-phenylisocyanate) (MDI) and their prepolymers. Method B is applicable to refined, crude or modified isocyanates derived from toluene diisocyanate, methylene-bis-(4-phenylisocyanate) and polymethylene polyphenylisocyanate. It can also be used for isomer mixtures of toluene diisocyanate, methylene-bis-(4-phenylisocyanate) and polymethylene polyphenylisocyanate. Other aromatic isocyanates may be analysed by this method if precautions are taken to verify suitability. It is not applicable to blocked isocyanates.

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2 Normative references

ISO 14896:2009

The following referenced documents are indispensable for the application of this document. For dated references, only the edition cited applies. For undated references, the latest edition of the referenced document (including any amendments) applies.

ISO 385, *Laboratory glassware — Burettes*

ISO 648, *Laboratory glassware — Single-volume pipettes*

ISO 3696, *Water for analytical laboratory use — Specification and test methods*

ISO 4787, *Laboratory glassware — Volumetric glassware — Methods for use and testing of capacity*

ISO 4788, *Laboratory glassware — Graduated measuring cylinders*

ISO 6353-1, *Reagents for chemical analysis — Part 1: General test methods*

ISO 6353-2, *Reagents for chemical analysis — Part 2: Specifications — First series*

ISO 6353-3, *Reagents for chemical analysis — Part 3: Specifications — Second series*

ISO 14898:1999, *Plastics — Aromatic isocyanates for use in the production of polyurethane — Determination of acidity*

3 Terms and definitions

For the purposes of this document, the following terms and definitions apply.

3.1 polyurethane
polymer prepared by the reaction of an organic di- or polyisocyanate with compounds containing two or more hydroxyl groups

NOTE Polyurethanes may be thermosetting, thermoplastic, rigid or soft and flexible, cellular or non-cellular.

3.2 assay
percent by mass of a specific isocyanate present in a sample

**3.3 isocyanate content
NCO content**
percent by mass of the NCO groups present in a sample

3.4 amine equivalent
mass of sample that will combine with 1 gram-equivalent of dibutylamine

4 Principle

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4.1 Method A

The isocyanate sample is reacted with an excess of dibutylamine in toluene to form the corresponding substituted urea. After cooling to room temperature, acetone is added as a co-solvent, then the reaction mixture is back-titrated with standardized aqueous HCl using potentiometric or colorimetric end point determination.

4.2 Method B

The isocyanate sample is reacted with an excess of dibutylamine in a toluene/trichlorobenzene solvent to form the corresponding substituted urea. After cooling to room temperature, the reaction mixture is diluted with methanol and back-titrated potentiometrically or colorimetrically with methanolic hydrochloric acid. See also 12.1.8.

5 Application

These test methods can be used for research or for quality control purposes to characterize isocyanates used in polyurethane products.

6 Interferences

Phosgene, the carbamyl chloride of the isocyanate, hydrogen chloride and any other acidic or basic compounds will interfere. In refined isocyanates, these impurities are usually present in such small amounts that they do not affect the determination; however, some crude or modified isocyanates may contain acidities of up to approximately 0,3 %, so the value reported for the NCO content of unrefined isocyanates should preferably be designated as “corrected” or “uncorrected” for acidity.

7 Sampling

Since organic isocyanates react with atmospheric moisture, take special precautions in sampling (see warning). Usual sampling methods (for example, sampling an open drum with a thief), even when conducted rapidly, can cause contamination of the sample with insoluble ureas; therefore, blanket the sample with a dry inert gas (e.g. nitrogen, argon or dried air) at all times.

WARNING — Organic isocyanates are hazardous when absorbed through the skin, or when the vapours are breathed. Provide adequate ventilation and wear protective gloves and eyeglasses.

8 Test conditions

Since isocyanates react with moisture, keep the laboratory humidity low, preferably below 50 % relative humidity.

9 Reagents

Use reagent-grade chemicals in all tests. Unless otherwise indicated, it is intended that all reagents shall conform to the specifications of ISO 6353-1, ISO 6353-2 and ISO 6353-3. Other grades may be used, provided that it is first determined that the reagent is of sufficiently high purity to permit its use without lessening the accuracy of the determination. Unless otherwise indicated, references to water shall be understood to mean grade 3 water as defined in ISO 3696.

9.1 Acetone (method A).

9.2 Toluene, dried over type 4A molecular sieve.

9.3 di-*n*-butylamine solution, 1 mol/l (method A).

Dilute 129 g of di-*n*-butylamine to 1 litre with toluene.

9.4 di-*n*-butylamine solution, 2 mol/l (method B).

Dilute 258 g of di-*n*-butylamine to 1 litre with toluene.

9.5 Aqueous hydrochloric acid, 1 mol/l (method A).

Prepare 1 mol/l aqueous hydrochloric acid and standardize frequently enough to detect changes of 0,001 mol/l.

9.6 Methanolic hydrochloric acid, 1 mol/l (method B).

Prepare 1 mol/l hydrochloric acid in methanol and standardize frequently enough to detect changes of 0,001 mol/l.

NOTE In order to have homogeneous solutions, it is recommended that methanolic HCl be used in this procedure. If desired, aqueous HCl can be used; however, turbidity will be encountered in some titrations. It is recommended that 200 ml to 250 ml of methanol be added to the reacted product to minimize the formation of two layers. Experience has shown that, if the mixtures are agitated vigorously, inhomogeneity can be tolerated without adversely affecting the results.

9.7 Bromophenol blue indicator solution, for colorimetric titration: 0,04 % aqueous solution of bromophenol blue sodium salt, reagent grade, or 0,04 % solution of bromophenol blue in acetone.

9.8 1,2,4-Trichlorobenzene (TCB), dried over type 4A molecular sieve (method B).

9.9 Methanol (method B).

10 Apparatus

10.1 Potentiometric titrator or pH-meter, accurate to 0,1 mV or better, equipped with a pair of electrodes or a combination glass-calomel electrode (filled with a 1 mol/l lithium chloride solution in methanol, or an equivalent solution) and a piston burette having a 20 ml capacity.

10.2 Syringes, capacity 2 ml and 5 ml, and **syringes with a large orifice** suitable for weighing viscous prepolymers by difference to the nearest 1 mg.

10.3 Magnetic stirrer.

10.4 Analytical balance, accurate to 0,1 mg.

10.5 Iodine flask, capacity 500 ml, with ground-glass neck (method A).

10.6 Conical flask, capacity 250 ml, with a wide mouth (method B).

10.7 Volumetric pipettes, capacity 25 ml (method A) and 20 ml (method B), conforming to ISO 4787.

10.8 Measuring pipette, capacity 1 ml, conforming to ISO 648.

10.9 Graduated cylinders, capacity 250 ml (method A) and 100 ml (method B), conforming to ISO 4788.

10.10 Beaker, capacity 500 ml.

10.11 Burette, capacity 50 ml, for colorimetric titration, conforming to ISO 385.

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11 Method A — Toluene/dibutylamine with aqueous HCl

11.1 Procedure

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11.1.1 Using a volumetric pipette (10.7), add 25 ml of 1 mol/l dibutylamine solution (9.3) to an iodine flask (10.5). Rinse the walls of the flask with 10 ml of toluene (9.2).

11.1.2 Weigh a sample of the product to be analysed to the nearest 0,1 mg using a suitable syringe (10.2). The sample used for analysis shall be completely liquid; if it contains crystallized isocyanates, heat it carefully until a homogeneous liquid phase is obtained. Add m_0 grams of the product to the dibutylamine solution in the iodine flask.

The mass (m_0) in grams of the product to be analysed shall contain (15 ± 5) milliequivalents of isocyanate or, in the case of TDI, about 1,5 g or, in the case of MDI, about 2,5 g.

In the event that the isocyanate equivalent is not known, a preliminary test should be run to determine the proper sample size to be used.

11.1.3 After complete dissolution, allow to react for 15 min at ambient temperature. The reaction will cause some warming of the solution. Let the solution stand until it reaches room temperature (an additional 5 min to 10 min).

11.1.4 Using a graduated cylinder (10.9), add 150 ml of acetone (9.1), taking care to rinse the flask walls and stopper.

11.1.5 Titrate the excess dibutylamine using one of the two following procedures:

11.1.5.1 Potentiometric titration (recommended)

Pour the contents of the iodine flask into the titration beaker (10.10), rinsing with 25 ml of acetone (9.1). Place the beaker on the magnetic stirrer (10.3) and stir the contents.

Immerse the electrodes in the reaction mixture and titrate with 1 mol/l aqueous hydrochloric acid (9.5), using the potentiometer (10.1) to determine the equivalence point.

11.1.5.2 Colorimetric titration

Place the iodine flask on the magnetic stirrer and stir the reaction mixture.

Using a graduated 1 ml pipette (10.8), add 0,8 ml of bromophenol blue solution (9.7).

Titrate using the burette containing 1 mol/l aqueous hydrochloric acid until the indicator changes from blue to yellow and remains stable for 15 s. The solution will change from a blue colour at the start of the titration to a bluish-green intermediate colour and to a yellow colour at the end point. Recognition of the end point is a matter of experience, but better defined colour changes are obtained when the acid is titrated rapidly into the solution until the first flash of yellow colour is observed. This flash of colour appears within a few tenths of a millilitre of the end point.

11.1.6 Conduct a blank determination under identical conditions, but omit the test portion.

11.2 Expression of results

11.2.1 Calculate the % NCO as follows:

$$\% \text{ NCO} = \frac{4,202 \times (V_1 - V_2) \times c}{m_0}$$

where

V_1 is the volume of HCl required for titration of the blank, in ml, to the nearest 0,01 ml;

V_2 is the volume of HCl required for titration of the test portion, in ml, to the nearest 0,01 ml;

c is the concentration of the HCl, in mol/l;

m_0 is the mass of the test portion, in g;

4,202 is a constant combining the equivalent mass of NCO (42,02 mg/mequiv), conversion of g to mg, and conversion to 100 %.

11.2.2 The amine equivalent may be calculated as follows:

$$\text{Amine equivalent} = \frac{1\,000 \times m_0}{(V_1 - V_2) \times c}$$

11.2.3 For isocyanates based on a single isomer or isomer mixture (for example, "pure" TDI or MDI), the assay may be calculated, in %, as follows:

$$\text{Assay} = \frac{(V_1 - V_2) \times c \times E}{1\,000 \times m_0} \times 100$$

where

V_1 is the volume of HCl required for titration of the blank, in ml, to the nearest 0,01 ml;

V_2 is the volume of HCl required for titration of the test portion, in ml, to the nearest 0,01 ml;

c is the concentration of the HCl, in mol/l;

m_0 is the mass of the test portion, in g;