Designation: E2562 – 07

Standard Test Method for Quantification of *Pseudomonas aeruginosa* Biofilm Grown with High Shear and Continuous Flow using CDC Biofilm Reactor[1]

This standard is issued under the fixed designation E2562; the number immediately following the designation indicates the year of original adoption or, in the case of revision, the year of last revision. A number in parentheses indicates the year of last reapproval. A superscript epsilon (ε) indicates an editorial change since the last revision or reapproval.

1. Scope

1.1 This test method specifies the operational parameters required to grow a repeatable *Pseudomonas aeruginosa* biofilm under high shear (**1**).[2] The resulting biofilm is representative of generalized situations where biofilm exists under high shear rather than representative of one particular environment.

1.2 This test method uses the Centers for Disease Control and Prevention (CDC) biofilm reactor. The CDC biofilm reactor is a continuously stirred flow reactor with high wall shear. Although it was originally designed to model a potable water system for the evaluation of *Legionella pneumophila* (**2**), the reactor is versatile and may also be used for growing and/or characterizing biofilm of varying species (**3 and 4**).

1.3 This test method describes how to sample and analyze biofilm for viable cells. Biofilm population density is recorded as log colony forming units per surface area.

1.4 Basic microbiology training is required to perform this test method.

1.5 The values stated in SI units are to be regarded as the standard. The values given in parentheses are for information only.

1.6 *This standard does not purport to address all of the safety concerns, if any, associated with its use. It is the responsibility of the user of this standard to establish appropriate safety and health practices and determine the applicability of regulatory limitations prior to use.*

2. Referenced Documents

2.1 *ASTM Standards:*[3]

D5465 Practice for Determining Microbial Colony Counts from Waters Analyzed by Plating Methods

2.2 *Other Standards:*

Method 9050 C.1a Buffered Dilution Water Preparation[4]

3. Terminology

3.1 *Definitions:*

3.1.1 *biofilm*, *n*—microorganisms living in a self-organized, cooperative community attached to surfaces, interfaces, or each other, embedded in a matrix of extracellular polymeric substances of microbial origin, while exhibiting an altered phenotype with respect to growth rate and gene transcription.

3.1.1.1 *Discussion*—Biofilms may be comprised of bacteria, fungi, algae, protozoa, viruses, or infinite combinations of these microorganisms. The qualitative characteristics of a biofilm (including, but not limited to, population density, taxonomic diversity, thickness, chemical gradients, chemical composition, consistency, and other materials in the matrix that are not produced by the biofilm microorganisms) are controlled by the physicochemical environment in which it exists.

3.1.2 *coupon*, *n*—biofilm sample surface.

4. Summary of Test Method

4.1 This test method is used for growing a repeatable *Pseudomonas aeruginosa* biofilm in a CDC biofilm reactor. The biofilm is established by operating the reactor in batch mode (no flow of the nutrients) for 24 h. A steady state population is reached while the reactor operates for an additional 24 h with continuous flow of the nutrients. The residence time of the nutrients in the reactor is set to select for biofilm growth, and is species and reactor parameter specific. During the entire 48 h, the biofilm is exposed to continuous fluid shear from the rotation of a baffled stir bar. Controlling the rate at which the baffle turns determines the intensity of the shear stress to which the coupons are exposed. At the end of the 48 h, biofilm accumulation is quantified by removing coupons from suspended rods, scraping the biofilm from the coupon surface, disaggregating the clumps, and diluting and plating for viable cell enumeration.

[1] This test method is under the jurisdiction of ASTM Committee E35 on Pesticides and Alternative Control Agents and is the direct responsibility of Subcommittee E35.15 on Antimicrobial Agents.

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[2] The boldface numbers in parentheses refer to a list of references at the end of this standard.

[3] For referenced ASTM standards, visit the ASTM website, www.astm.org, or contact ASTM Customer Service at service@astm.org. For *Annual Book of ASTM Standards* volume information, refer to the standard's Document Summary page on the ASTM website.

[4] Eaton, A.D., Clesceri, L.S., Rice, E.W., Greenberg, A.E., (Eds.) *Standard Methods for the Examination of Water and Waste Water* , 21st Edition, American Public Health Association, American Water Works Association, Water Environment Federation, Washington D.C., 2005.

5. Significance and Use

5.1 Bacteria that exist in biofilm are phenotypically different from suspended cells of the same genotype. Research has shown that biofilm bacteria are more difficult to kill than suspended bacteria (**5**). Laboratory biofilms are engineered in growth reactors designed to produce a specific biofilm type. Altering system parameters will correspondingly result in a change in the biofilm. For example, research has shown that biofilm grown under high shear is more difficult to kill than biofilm grown under low shear (**6**). The purpose of this test method is to direct a user in the laboratory study of a *Pseudomonas aeruginosa* biofilm by clearly defining each system parameter. This test method will enable an investigator to grow, sample, and analyze a *Pseudomonas aeruginosa* biofilm grown under high shear. The biofilm generated in the CDC biofilm reactor is also suitable for efficacy testing. After the 48 h growth phase is complete, the user may add the treatment in situ or harvest the coupons and treat them individually.

6. Apparatus

6.1 *Wooden Applicator Sticks*, sterile.

6.2 *Inoculating Loop.*

6.3 *Petri Dish*, 100 by 15 mm, plastic, sterile and empty to put beneath rod while sampling.

6.4 *Culture Tubes and Culture Tube Closures*, any with a volume capacity of 10 mL and a minimum diameter of 16 mm. Recommended size is 16 by 125 mm borosilicate glass with threaded opening.

6.5 *Pipetter*—Continuously adjustable pipetter with volume capability of 1 mL.

6.6 *Vortex*—Any vortex that will ensure proper agitation and mixing of culture tubes.

6.7 *Homogenizer*—Any that can mix at 20 500 ± 5000 r/min in a 5 to 10 mL volume.

6.8 *Homogenizer Probe*—Any that can mix at 20 500 ± 5000 r/min in a 5 to 10 mL volume and can withstand autoclaving or other means of sterilization.

6.9 *Sonicator*—Any noncavitating sonicating bath that operates at 50 to 60 Hz.

6.10 *Bunsen Burner*, used to flame inoculating loop and other instruments.

6.11 *Stainless Steel Hemostat Clamp*, with curved tip.

6.12 *Environmental Shaker*, that can maintain a temperature of 35 ± 2°C.

6.13 *Analytical Balance*, sensitive to 0.01 g.

6.14 *Sterilizers*—Any steam sterilizer that can produce the conditions of sterilization is acceptable.

6.15 *Colony Counter*—Any one of several types may be used, such as the Quebec, Buck, and Wolfhuegel. A hand tally for the recording of the bacterial count is recommended if manual counting is done.

6.16 *Peristaltic Pump*—Pump head that can hold tubing with ID 3.1 mm and OD 3.2 mm.

6.17 *Magnetic Stir Plate*—Top plate 10.16 × 10.16 cm, that can rotate at 125 ± 60 r/min.

NOTE 1—A digital stir plate is recommended.

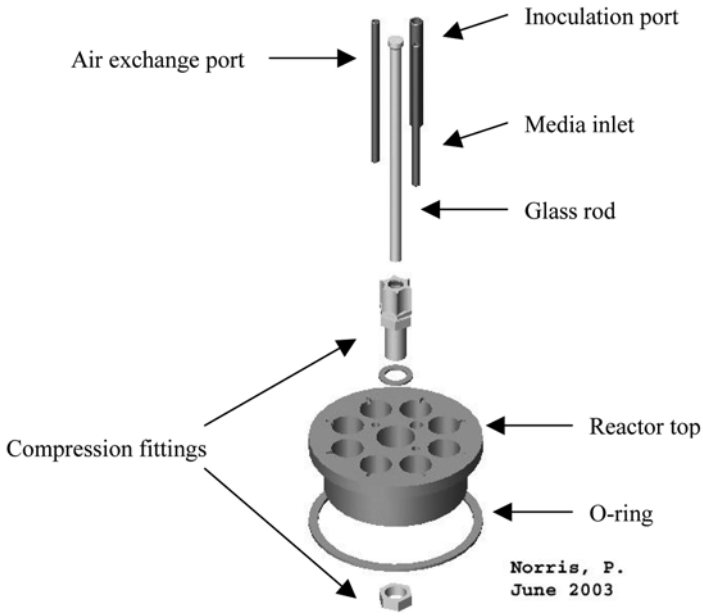

FIG. 1 Expanded Schematic of Reactor Top

6.18 *Silicone Tubing*—Two sizes of tubing: one with ID 3.1 mm and OD 3.2 mm and the other with ID 7.9 mm and OD 9.5 mm. Both sizes must withstand sterilization.

6.19 *Glass Flow Break*—Any that will connect with tubing of ID 3.1 mm and withstands sterilization.

6.19.1 *Clamp*—Used to hold flow break, extension clamp with 0.5 cm minimum grip size.

6.19.2 *Clamp Stand*—Height no less than 76.2 cm, used with clamp to suspend glass flow break vertically and stabilize tubing above reactor.

6.20 *Reactor Components.*[5]

6.20.1 *Berzelius Pyrex or Kimax Tall Beaker*, 1000 mL without pour spout, 9.5 ± 0.5 cm diameter. Pyrex/Kimax barbed outlet spout added at 400 ± 20 mL mark. Angle the spout 30 to 45° to ensure drainage. Spout should accommodate flexible tubing with an ID of 8 to 11 mm.

NOTE 2—The rods, described in 6.20.3 and baffle (6.20.5) will displace approximately 50 mL of liquid when system is completely assembled. Therefore, an outlet spout at the 400 mL mark will result in approximately a 350 mL operating volume. The user is encouraged to confirm the actual liquid volume in the reactor, when the rods and baffle are in place, before use. The measured volume is used to calculate an exact pump flow rate.

6.20.2 *Reactor Top*—See Fig. 1. Ultra-high molecular weight (UHMW) polyethylene top (10.1 cm diameter tapering to 8.33 cm) equipped with 3 holes accommodating 10 cm pieces of stainless steel or other rigid autoclavable tubing with OD of 5 to 8 mm for media inlet, air exchange, and inoculation port. Center hole, 1.27 cm diameter, to accommodate the glass rod used to support the baffle assembly. Eight rod holes, 1.905

[5] The sole source of supply of the apparatus (CDC Biofilm reactor) known to the committee at this time is BioSurface Technologies, Corp. www.biofilms.biz. If you are aware of alternative suppliers, please provide this information to ASTM International Headquarters. Your comments will receive careful consideration at a meeting of the responsible technical committee,[1] which you may attend. The user may also build the reactor.

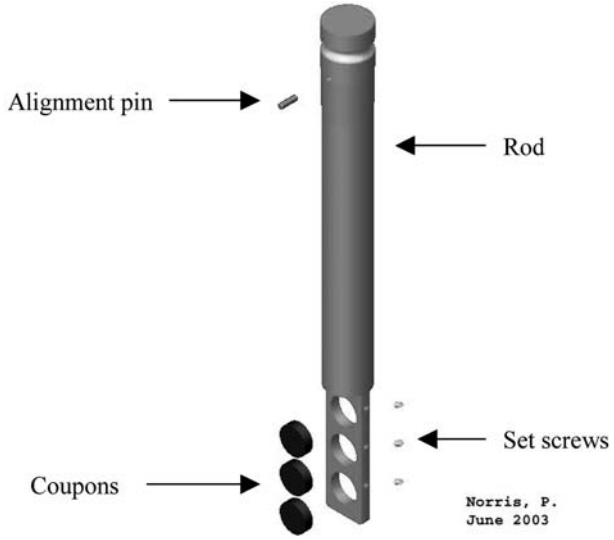

FIG. 2 Expanded Schematic of Rod and Coupons

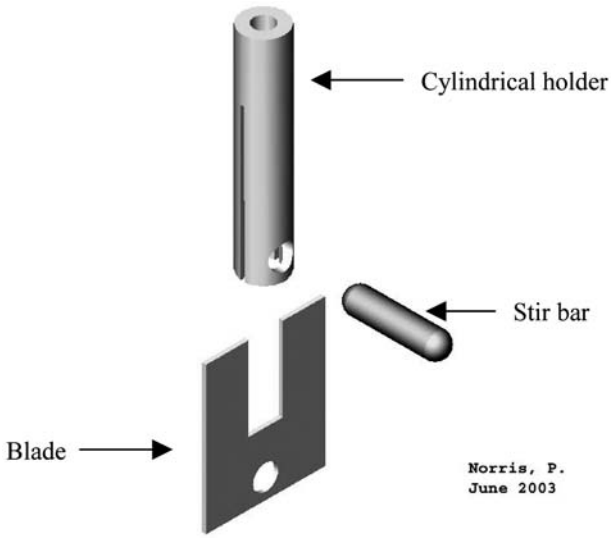

FIG. 3 Expanded Schematic of Baffled Stir Bar

cm diameter, notched to accommodate stainless steel rod alignment pin (0.236 cm OD).

6.20.3 *Polypropylene Rods*—See Fig. 2. Eight polypropylene rods, 21.08 cm long, machined to hold three coupons (see 6.20.4) at the immersed end. 316 stainless steel set screws imbedded in side to hold coupons in place. Rods fit into holes in reactor top and lock into preformed notches.

6.20.4 *Twenty-four Cylindrical Polycarbonate Coupons*—with a diameter of 1.27 ± 0.013 cm, thickness of approximately 3.0 mm.

6.20.5 *Small Allen Wrench*, for loosening set screws.

6.20.6 *Stir Blade Assembly (Baffled Stir Bar)*—See Fig. 3. PTFE blade (5.61 cm) fitted into cylindrical PTFE holder (8.13 cm) and held in place with a magnetic stir bar (2.54 cm). PTFE holder fits onto a glass rod (15.8 cm), fitted into the reactor top. The glass rod is held in place with a compression fitting and acts as a support for the moving blade assembly.

6.21 *Carboys*—Two 20-L autoclavable carboys, to be used for waste and nutrients.

6.21.1 *Two Carboy Lids*—One carboy lid with at least two barbed fittings to accommodate tubing ID 3.1 mm (one for nutrient line and one for bacterial air vent). One carboy lid with at least two 1-cm holes bored in the same fashion (one for effluent waste and one for bacterial air vent).

NOTE 3—Carboy tops can be purchased with fittings.

6.21.2 *Bacterial Air Vent (Filter)*—Autoclavable 0.2 µm pore size, to be spliced into tubing on waste carboy, nutrient carboy and reactor top; recommended diameter 37 mm.

6.22 Fig. 4 illustrates a schematic of the assembled system.

7. Reagents and Materials

7.1 *Purity of Water*—All reference to water as diluent or reagent shall mean distilled water or water of equal purity.

7.2 *Culture Media*:

7.2.1 *Bacterial Liquid Growth Broth*—Tryptic Soy Broth (TSB) is recommended.

7.2.2 *Bacterial Plating Medium*—R2A Agar is recommended.

NOTE 4—Two different TSB concentrations are used in the test method, 300 mg/L for the inoculum and batch reactor operation and 100 mg/L for the continuous flow reactor operation.

7.3 *Buffered Water*—0.0425 g/l KH_2PO_4 distilled water, filter sterilized and 0.405 g/l $MgCl{\cdot}6H_2O$ distilled water, filter sterilized (prepared according to Method 9050 C.1a).

8. Culture Preparation

8.1 *Pseudomonas aeruginosa* ATCC 700888 is the organism used in this test. Aseptically remove 3 to 5 isolated colonies with the same morphology from an R2A plate and place into 100 mL of sterile TSB (300 mg/L). Incubate bacterial suspension in an environmental shaker at 35 ± 2°C for 20 to 24 h. Viable bacterial density should equal 10^8 CFU/mL, and may be checked by serial dilution and plating.

9. Reactor Preparation

9.1 *Preparation of Polycarbonate Coupons*:

NOTE 5—Coupons can be used once and discarded or used repeatedly with proper cleaning and sterilization between each use. Check each coupon for scratching, chipping, other damage or accumulated debris before each use by screening under a dissecting microscope at a magnification of at least 20×. Discard those with visible damage to surface topography.

9.1.1 Sonicate coupons for 30 s in a 1:100 dilution of laboratory soap and tap water. The soapy water must completely cover the coupons.

9.1.2 Rinse coupons with reagent water and sonicate for 30 s in reagent water.

9.1.3 Repeat rinsing and sonication with reagent water until no soap is left on the coupon. Once the coupons are clean, care must be taken to prevent oils and other residue from contaminating the surface.

9.1.4 Place a coupon into each hole in the reactor rods, leaving the top of the coupon flush with the inside rod surface. Tighten set screw.

9.1.5 Place rods into reactor top loosely (not yet fitted into notches).