

Designation: D 4763 - 88 (Reapproved 2001)

An American National Standard

Standard Practice for Identification of Chemicals in Water by Fluorescence Spectroscopy¹

This standard is issued under the fixed designation D 4763; the number immediately following the designation indicates the year of original adoption or, in the case of revision, the year of last revision. A number in parentheses indicates the year of last reapproval. A superscript epsilon (ϵ) indicates an editorial change since the last revision or reapproval.

1. Scope

- 1.1 This practice allows for the identification of 90 chemicals that may be found in water or in surface layers on water. This practice is based on the use of room-temperature fluorescence spectra taken from lists developed by the U.S. Environmental Protection Agency and the U.S. Coast Guard (1).² Ref (1) is the primary source for these spectra. This practice is also based on the assumption that such chemicals are either present in aqueous solution or are extracted from water into an appropriate solvent.
- 1.2 Although many organic chemicals containing aromatic rings, heterocyclic rings, or extended conjugated double-bond systems have appreciable quantum yields of fluorescence, this practice is designed only for the specific compounds listed. If present in complex mixtures, preseparation by high-performance liquid chromatography (HPLC), column chromatography, or thin-layer chromatography (TLC) would probably be required.
- 1.3 If used with HPLC, this practice could be used for the identification of fluorescence spectra generated by optical multichannel analyzers (OMA) or diode-array detectors.
- 1.4 For simple mixtures, or in the presence of other non-fluorescing chemicals, separatory techniques might not be required. The excitation and emission maximum wavelengths listed in this practice could be used with standard fluorescence techniques Refs (2-6) to quantitate these ninety chemicals once identification had been established. For such uses, generation of a calibration curve, to determine the linear range for use of fluorescence quantitation would be required for each chemical. Examination of solvent blanks to subtract or eliminate any fluorescence background would probably be required.
- 1.5 This standard does not purport to address the safety concerns, if any, associated with its use. It is the responsibility of the user of this standard to establish appropriate safety and health practices and determine the applicability of regulatory limitations prior to use.

2. Referenced Documents

- 2.1 ASTM Standards:
- D 1129 Terminology Relating to Water³
- D 1193 Specification for Reagent Water³
- E 131 Terminology Relating to Molecular Spectroscopy⁴
- E 275 Practice for Describing and Measuring Performance of Ultraviolet, Visible, and Near Infrared Spectrophotometers⁴

3. Terminology

3.1 *Definitions*—For definitions of terms used in this practice, refer to Terminology D 1129, Specification D 1193, and definitions under the jurisdiction of Committee E-13 such as Definitions E 131 and Practice E 275.

4. Summary of Practice

- 4.1 This practice uses well tested fluorescence techniques to detect and identify (or determine the absence of) 90 chemicals that have relatively high fluorescence yields. Table 1 lists for each chemical an appropriate solvent (either cyclohexane, water, methyl or ethyl alcohol, depending on solubility), a suggested excitation wavelength for maximum sensitivity, a wavelength corresponding to the emission maximum, the number of fluorescence peaks and shoulders, the width (full width at half of the maximum emission intensity) of the strongest fluorescence peak and the detection limit for the experimental conditions given. Detection limits could be lowered, following identification, by using broader slit widths. A list of corrected fluorescence spectra for the chemicals included in this practice are also available (1).
- 4.2 Identification of the sample is made by comparison of the obtained spectra with information in Table 1 and by direct visual comparison of appropriate spectra with positions of principal peaks in agreement to ± 2 nm and ratios of peak heights in agreement to ± 10 % if corrected spectrofluorometers are used.
- 4.3 Spectral distortions due to self-absorption or fluorescence quenching or dimer formation may occur at higher concentrations (for example, 100 ppm or μg/mL). If this is

¹ This practice is under the jurisdiction of ASTM Committee D19 on Water and is the direct responsibility of Subcommittee D19.06 on Methods for Analysis for Organic Substances in Water.

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² The boldface numbers in parentheses refer to the list of references at the end of this practice.

³ Annual Book of ASTM Standards, Vol 11.01.

⁴ Annual Book of ASTM Standards, Vol 03.06.

suspected, the solution should be diluted and additional fluorescence spectra generated. If a suspected chemical is not detected on excitation at the appropriate wavelength, it usually can be assumed that it is not present above the detection limit, barring interference effects due to absorption or quenching that can usually be anticipated.

5. Significance and Use

5.1 This practice is useful for detecting and identifying (or determining the absence of) 90 chemicals with relatively high fluorescence yields (see Table 1). Most commonly, this practice

will be useful for distinguishing single fluorescent chemicals in solution, simple mixtures or single fluorescing chemicals in the presence of other nonfluorescing chemicals. Chemicals with high fluorescence yields tend to have aromatic rings, some heterocyclic rings or extended conjugated double-bond systems. Typical chemicals included on this list include aromatics, substituted aromatics such as phenols, polycyclic aromatic hydrocarbons (PAH's), some pesticides such as DDT, polychlorinated biphenyls (PCB's), some heterocyclics, and some esters, organic acids, and ketones.

TABLE 1 Summary of Experimental Parameters and Results

Chemical	Code	Concentra- tions, ppm	Solvent	λ _{exc} , nm	λ_{max}^{em} , nm	Number of Peaks	WHM, nm	Shoulder Number	Detection Limit (DL), ppm	λ_{DL} , nm	Comments
Acenaphthene	ACN	1.03	CH	290	323	4		3	0.001	290	
Acetone	ACT	227	CH	290	410	1			212	290	
Acridine	ACR	96	CH	285/355	386/422	4/2		2/0			
	ACR	9.6	ETOH	290/355	357/415	2/2		1/1	0.02/0.04	290/355	
Aniline	ANL	15.5	CH	280	316	1			0.037	280	
Anthracene	ATH	1.03	CH	355	378	4		1	0.001	355	
	ATH	1.55	ETOH	355	380	4		1	0.001	355	
Aroclor 1242	PC4	131	CH	270	317	2	35	1	0.3	270	
1254	PC5	129	CH	270	317	2	36	1	2	270	
Atrazine	ATZ	369	CH	290	350	1			300	290	
Azinphosmethyl	AZP	112	CH •	350	410	2	60		10	350	
	AZP	122	ETOH	340	420	2	80	2.	4	340	
Benz(a)anthracene	BAT	1.1	CH	280	386	4		1	0.003	280	
Benzene	BNZ	79	CH	250	279	3	24	1 _	2/4	250/265	
Benzonitrile	BZN	9.9	CH	260	287	200	28	10h	0.1/0.1	260/270	
Benzo(a)pyrene	BAP	0.088	CH	370	405	6	119°I	2	0.002	370	
Benzyl alcohol	BAL	99	CH	250	284	2	27	1	0.1/0.1	250/260	
Benzyl amine	BZM	118	CH	250	283	1- D	27	2	3/2	250/260	
Benzyl triethylam- monium chloride	BMA	210	H ₂ O	250	280		28		59	250	
Bisphenol A	BPA	10.5	ETOH	270	304	1	30	1	0.04/0.02	270/285	
Brucine	BRU	13.5	ETOH	280	327	1 000	56		2/2	280/295	
O-tert-Butylphenol	BOP	21	CH	265	295	<u>(13-88(</u> ∠	30	1	0.1/0.1	265/275	
p-tert-Butylphenol	BTP	17.5	CH	260	295	1716	31	540 204	0.6/0.4	260/280	
Carbaryl Standards	CBY	9.0 210 9/81	CH	285	335	2/11a-4	36	322-39L	0.01	285	
Carnauba wax	WCA	63.5	CH	260	310	1	64		42	260	
Castor oil	OCA	390	ETOH	290	328	1	43	2	20	290	
	OCA	286	CH	280/320		1			180/300	280/320	
Catechol	CTC	8.7	H_2O	265	310	1	46		0.4/0.2	265/280	
4-Chloroaniline	CAP	17.2	CH	290	328	1	36	1	0.2	290	
1-Chloronaphthalene	CNA	11.3	CH	290	328	3	34	4	0.1	290	
p-Chlorophenol	CPN	101	CH	260	305	1	30		1/0.1	260/285	
Chlorpyrifos (Duraban)	DUR	25.3	CH	280	326	1	52		1/0.5	280/295	
p-Chlorotoluene	CTN	23.8	CH	265	288	1	29	3	1/0.8	265/275	
p-Chloro-o-toluidine	COT	25	CH	290	328	1	39	1	0.09	300	
Chrysene	CRY	1.0	CH	270	383	5			0.002	270	
Coconut oil	OCC	286	CH	290	330				100	290	
Cod liver oil	OCL	323	CH	260/280	320/320	1/1	150		260,140	260,280	
				330	500	1			65	330	
Copper naphthenate	CNN	98	CH	260	326	1	60	3	3/1	260/280	
Cottonseed oil	ocs	305	CH	280/320	320/380				165,300	280,320	
Coumaphos	COU	11.4	CH	320	377	1	74		0.3	320	
o-Cresol	CRO	12.0	CH	265	293	1	30	1	0.04	280	
o-Cresol	CRP	10.3	CH	265	299	1	30		0.03	280	
Cumene	CUM	101	CH	250	283	2	28	1	3	250	
o-Cymene	CMP	11.8	CH	260	285	1	28	2	0.4/0.2	260/270	
DDD	DDD	61.0	CH	240	294	1	30	2	4	240	
ODT	DDT	87	CH	245	291	2	28	2	7	245	
1,2,5,6-Dibenzanthracene	DBA	0.015	CH	300	396	4		2	0.001	300	
Dicamba	DIC	22.2	H ₂ O	310	420	1	70	-	0.9	310	
Dichlorobenil	DIB	108	CH	285	312	1	30		0.6	285	
2,4-Dichlorophenoxy- acetic acid	DCA	159	CH	270	310	1	46	1	30	270	
Diethylbenzene	DEB	100	CH	255	283	1	28	2	0.2/0.1	255/270	
Diethylene glycol	DEG	202	CH	265	310	2			202	265	
Diethylphthalate	DEP	145/289	CH	260/280	300/320	1/1				280	

TABLE 1 Continued

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Chemical	Code	Concentra- tions, ppm	Solvent	λ_{exc} , nm	λ_{max}^{em} , nm	Number of Peaks	WHM, nm	Shoulder Number	Detection Limit (DL), ppm	λ_{DL} , nm	Comments
2,4-Dimethylphenol	DMH	10.5	CH	265	300	1	31	1	0.2/0.04	265/280	
3,5-Dimethylphenol	DPM	10.5	CH	265	295	1	28	1	0.07/0.03	265/280	
Diphenylamine	DAM	11.2	CH	290	333	1	37	2		290	photochemical
		1.2	CH	290	333	1	37	2		290	change
Diphenyldichlorosilane	DDS	157	CH	260	285	2	30	-	3/2	260/270	
Diquat dibromide	DQD	35.5	H ₂ O	310	348	1	41	1	0.055	310	
Dodecylbenzene	DDB	116	CH	250	285	3	30		*	250	* strong impurity
Bodcoyiberizerie	DDD	116	CH	220	285	3	30		13.6	220	Strong impunty
Dowtherm A	DTH	10.8	CH	260	305	2	33	2	0.035	260	
Ethylbenzene	ETB	10.0	CH	250	283	2	26		3.1/1.5	250/260	
Fluoranthene	FLA	1.0	CH	360	465	2	91	3	0.005	360	
Gallic acid											
	GLA	103	H ₂ O	290	346	1	77		0.70	290	
Hydroquinone	HDQ	1.1	H ₂ O	290	326	1	38	1	0.025	290	
Indene	IND	175	CH	260	309	2	32	3	0.12	260	
Lard	OLD	340	CH	270	330				•••	270	
	OLD	287	CH	280	330	1				280	
Linseed oil	OLS	355	CH	300	418	1	105		32	300	
Methoxychlor	MOC	95	CH	270	299	1	30	1	1.3/0.8	270,280	
Methylaniline	MAN	10.8	CH	290	325	1	35		0.01	290	
Methyl isobutyl ketone	MIK	358	CH	290	400	1			•••	290	
Methyl styrene	MSR	105	CH	255	307	1	35	2	0.12	255	
Naphthalene	NPT	10.5	CH	280	323	2	24	3	0.02	280	
1-Naphthylamine	NAD	1.85	CH	325	377	1	55	1	0.0012	325	
Nonyl phenol	NNP	17.1	CH	265	298	1	28		0.09	265	
Olive oil	OOL	237	CH	260	320	1				360	
	OOL	290	CH	310						310	
Palm oil	OPM	300	CH	260	320	1	60		218	260	
			CH	350	500	$\mathbf{n} \cap \mathbf{n}$	140	22	300	350	
Peanut oil	OPN	249	CH	260,290	120,320	1		15			
Phenol	PHN	11.9	CH	265	288	1	30	2	0.011/0.007		
Phenyl ether	DPE	20.4	CH	265	291	dor	36	110h	0.10	265	
Phthalic acid	PHA	97	H ₂ O	280	330	yai (100		84	280	
	PHA	228	H ₂ O	270	340	1	100		114	270	
Piperazine	PPZ	235	CH	280	350	1- D.					
Polyethoxylated non- ylphenol	PEN	9.5	CH	265	297		30		0.08/0.03 17	265/280	
Pyrogallol	PGA	152	H ₂ O	270	335	1	86	1	30	270	
Quinoline	QNL	113	ETOH	275	321	5		2			photolyzes
Quillonio	Q. VL	113	ETOH	355	420	<u> 63-88(2</u>	70	0			photolyzes
		c95talog/s	CH	275 350	336	$\frac{3}{2}71 \text{fa}$	1599-a	$5\frac{2}{1}2-395$	0.37 febe	1275 117astm-d47	photolyzes
December	DCC							-	0.105/0.05	265/280	
Resorcinol	RSC	10.1	H ₂ O	265	303	1	39	1	0.135/0.05		
Salicylic acid	SLA	1.5	H ₂ O	300	409	1	64		0.005	300	
Sodium dodecylben- zenesulfonate	SDB	90	CH	290	347	1	52	2	0.90	290	
Soya bean oil	OSB	290	CH	270,320					0.300	270,320	
Styrene	STY	1.1	CH	270	306	2	32	2	0.03	270	
Tanaic acid	TNA	13	H_2O	280	340	1	100		0.63	280	
1,2,3,4-Tetrahydro- naphthalene	THN	12.3	CH	260	284	1	27	2	0.21/0.13	260/270	
p-Toluidine	TLI	14.1	CH	290	325	1	34		0.03	290	
Toluene	TOL	107	CH	250	284	2	27	1	2.1/1.6	250/215	
p-Toluene sulfonic acid	TAP	120	H ₂ O	260	285	1	28	1	2.1/1.5	260/265	
Tricresylphosphate	TCP	123	CH	260	288	1	66	1	0.55/0.35	260/270	
1,3,5-Triethylbenzene	TEB	122	CH	250	292	1	28	3	12.5/1.5	250/270	
Turpentive	TPT	301	CH	260	283	1	34	3	31/13	260/270	
Undecylbenzene	UDB	87.3	CH	250	284	2	33	2	6.0	250	
Uranyl nitrate	UAN	61.0	H ₂ O	290	520	3	56	2	6.1/10.5	290/330	
-				290 260		3 1		1	2.0/1.4	260/270	
m-Xylene	XLM	114	CH		285		28				
o-Xylene	XLO	92	CH	260	285	1	30		1.5/1.3	260/270	

5.2 With appropriate separatory techniques (HPLC, TLC, and column chromatography) and in some cases, special detection techniques (OMA's and diode arrays), this practice can be used to determine these 90 chemicals even in complex mixtures containing a number of other fluorescing chemicals. With the use of appropriate excitation and emission wave-

lengths and prior generation of calibration curves, this practice could be used for quantitation of these chemicals over a broad linear range.

5.3 Fluorescence is appropriately a trace technique and at higher concentrations (greater than 10 to 100 ppm) spectral distortions usually due to self-absorption, or inner-filter effects