

Designation: D 4779 – 93

Standard Test Method for Total, Organic, and Inorganic Carbon in High Purity Water by Ultraviolet (UV) or Persulfate Oxidation, or Both, and Infrared Detection ¹

This standard is issued under the fixed designation D 4779; the number immediately following the designation indicates the year of original adoption or, in the case of revision, the year of last revision. A number in parentheses indicates the year of last reapproval. A superscript epsilon (ε) indicates an editorial change since the last revision or reapproval.

1. Scope

1.1 This test method covers the determination of total carbon (TC), organic carbon (OC), and inorganic carbon (IC), in makeup water and high purity process water such as demineralizer effluent, condensate, and electronic grade rinse water. The tested concentration range is from 50 to 1000 µg of carbon per litre.

1.2 It is the user's responsibility to ensure the validity of this test method for waters of untested matrices.

1.3 *This standard does not purport to address all of the safety problems, if any, associated with its use. It is the responsibility of the user of this standard to establish appro-*
priate safety and health practices and determine the applica as peak heigh *priate safety and health practices and determine the applicability of regulatory limitations prior to use.*

2. Referenced Documents

2.1 *ASTM Standards:*

- D 1129 Terminology Relating to Water²
- D 1192 Specification for Equipment for Sampling Water and Steam in Closed Conduits²

D 1193 Specification for Reagent Water²

- D 2777 Practice for Determination of Precision and Bias of Applicable Methods of Committee D-19 on Water²
- D 3370 Practices for Sampling Water from Closed Con $duits²$

3. Terminology

3.1 *Definitions*—For definitions of terms used in this test method, refer to Terminology D 1129.

3.2 *Definitions of Terms Specific to This Standard:*

3.2.1 *inorganic carbon (IC)*—carbon in the form of carbon dioxide, carbonate ion, or bicarbonate ion.

3.2.2 *organic carbon (OC; frequently also TOC)*—carbon in the form of organic compounds.

² *Annual Book of ASTM Standards*, Vol 11.01.

3.2.3 *total carbon (TC)*—the sum of inorganic and organic carbon.

4. Summary of Test Method

4.1 For total carbon measurement, sample is injected into a gas-sparged reactor containing acidified potassium persulfate $(K_2S_2O_8)$ or sodium persulfate $(Na_2S_2O_8)$ solution; either elevated temperature or ultraviolet (UV) radiation is used to enhance the oxidation. Both inorganic and organic carbon compounds are converted into $CO₂$, which is swept, either directly or by trapping and thermal desorption, to a $CO₂$ specific linearized infrared detector. Output signal is measured as peak height or integrated area and results displayed as fractional milligrams of carbon per litre or equivalent. For direct organic carbon determination, the sample is acidified and
 EXECUTE: direct organic carbon determination, the sample is acidified and

sparaed to remove increasing carbon, prior to ovidation (pures sparged to remove inorganic carbon, prior to oxidation (purgeable organic compounds may be lost in this procedure). For Water² **CUIMENT** able organic compounds may be lost in this procedure). For inorganic carbon measurement, the CO₂ sparged off in the organic carbon step may be quantified, or the sample may be injected into the reactor with the UV source off so that organics $\frac{\text{ASTM D477}}{\text{are not oxidized}}$.

 $\frac{1}{2}$ D 2777 Practice for Determination of Precision and Bias of $\frac{1}{2}$ Organic carbon may also be measured as the difference between" total carbon'' and "inorganic carbon'' results.

5. Significance and Use

5.1 Accurate measurement of organic carbon in water at low and very low levels is of particular interest to the electronic, pharmaceutical, and steam power generation industries.

5.2 Elevated levels of organics in raw water tend to degrade ion exchange capacity. Elevated levels of organics in high purity water tend to encourage biological growth and, in some cases, are directly detrimental to the processes that require high purity water.

5.3 In the case of steam power generation, naturally occurring organics can degrade to $CO₂$ and low molecular weight organic acids which, in turn, are corrosive to the process equipment.

5.4 Inorganic carbon can also cause problems in a steam power system. $CO₂$ entering steam condensate that contains ammonia, reacts to form ammonium carbonate, which is not removed by the condenser air ejection system. If condensate

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¹ This test method is under the jurisdiction of ASTM Committee D-19 on Water and is the direct responsibility of Subcommittee D19.11 on Standards for Water for Power Generation and Processes.

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polishers are operated beyond the ammonia break, continued operation on an ammonium cycle can result in selective exhaustion of the anion resin to the carbonate form, eluting silica, chloride, and sulfate into the condensate. The effect is immediately felt with powdered resin systems that have a very small inventory of anion resin.

6. Interferences and Limitations

6.1 If IC level is much higher than OC, the latter should be determined directly by acidifying the sample and sparging off IC before injection. Determination of OC by difference may introduce large error in such circumstances.

6.2 The process of removing IC by sparging may also remove some organic compounds, termed purgeable organic carbon (POC). The measurement done on the sparged sample will therefore be nonpurgeable organic carbon, and will not necessarily be equal to the OC figure arrived at by subtracting the IC measurement from the TC measurement. Users of this test method are responsible for determining whether the POC fraction is significant in their samples.

6.3 High-purity water is a very active scavenger of $CO₂$ and other impurities from air, syringes, bottles, pipes, etc. Stringent precautions must be taken to prevent sample contamination during collection, transportation, storage, and analysis.

6.4 *Method Accuracy*:

6.4.1 To produce accurate OC data, both method blank and may be used to overy must be known recovery must be known.

6.4.1.1 *Method Blank*—The blank response of a method that has been found satisfact ust be determined and subtracted from the sample response. must be determined and subtracted from the sample response. This is especially true when making very low level measure-This is especially true when making very low level measure-

ments as in the case of high purity water applications. Some 8.1 Purity of Red

previewall as a preview of a particle was (1) the semple and in all tests. Un examples of contributors to method blank are: (1) the sample injection device used; (2) inlet septa; (3) chemical conversion method used; and (4) carrier gases, etc.

6.4.1.2 *Method Recovery*—To produce valid OC data, it must be assumed that all compounds are converted to a detectable species (that is, $CO₂$) with the same efficiency, independent of compound type or sample matrix. Since the conversion efficiency can be affected by many factors, it should be checked from time to time with selected compound types.

6.4.2 As an aid to checking recovery, the following compounds are listed in decreasing order of oxidation rate by UV-promoted persulfate oxidation:

6.4.2.1 Potassium acid phthalate (KHP),

- 6.4.2.2 Urea,
- 6.4.2.3 Nicotinic acid,
- 6.4.2.4 Pyridine,
- 6.4.2.5 n-Butanol,
- 6.4.2.6 Acetic acid,
- 6.4.2.7 Leucine, and
- 6.4.2.8 Acetonitrile.

6.4.3 As an expedient for most applications, method validation can be checked using KHP, acetic acid, and acetonitrile in deionized water. Ideally, all solutions should give equivalent conversion efficiencies (for example, percent recovery).

6.5 As with other methods for TC, IC, OC, and other water quality parameters such as COD, this test method inherently entails limitations. For example, the relatively low temperature oxidation will not oxidize graphite or fines from an activated carbon bed. Certain dissolved organics in water may not fully oxidize in this test method, yielding an error. One such component known to produce low recovery is carbon tetrachloride. The users of this test method are encouraged to verify performance of the method on the compounds or sample types of interest in their application.

7. Apparatus

7.1 *Carbon Analyzer*³ —A reagent and sample introduction mechanism, a gas-sparged reaction vessel, a gas demister or dryer, or both, a $CO₂$ trap (optional), a $CO₂$ -specific infrared detector, a control system, and a display.

7.2 *Sparging Apparatus*—A glass vessel and supply of $CO₂$ -free gas to be bubbled through a water sample to remove inorganic carbon as $CO₂$.

7.3 *Sample Injector*—An all-fluorocarbon sampling valve, such as used for sample introduction in liquid chromatography, may be used to introduce the sample.

7.4 Fig. 1 shows a diagrammatic presentation of an analyzer that has been found satisfactory for this purpose.

8. Reagents and Materials

8.1 *Purity of Reagents*—Reagent grade chemicals shall be used in all tests. Unless otherwise indicated, it is intended that all reagents shall conform to the specifications of the Commit-ASTM D477 tee on Analytical Reagents of the American Chemical Society,⁴ where such specifications are available. Other grades may be 0.4.1.2 Method Recovery—10 produce valid OC data, it where such specifications are available. Since grades may be
nust be assumed that all compounds are converted to a used, provided it is first ascertained that the reage sufficient purity to permit its use without decreasing the accuracy of the determination.

> 8.2 *Purity of Water*—Unless otherwise stated, references to water shall be understood to mean reagent water conforming to Specification D 1193, Type II. The OC of this water should be measured regularly and this value should be taken into consideration when preparing standards. It will typically be in the

⁴ *Reagent Chemicals, American Chemical Society Specifications,* American Chemical Society, Washington, DC. For suggestions on the testing of reagents not listed by the American Chemical Society, see *Analar Standards for Laboratory Chemicals,* BDH Ltd., Poole, Dorset, U.K., and the *United States Pharmacopeia and National Formulary,* U.S. Pharmaceutical Convention, Inc. (USPC), Rockville, MD.

FIG. 1 Diagrammatic Presentation of an Analyzer

³ Model DC-80 TOC analyzer marketed by Dohrmann and model 700 TOC analyzer marketed by OI Corp were used in the collaborative study.

range of 0.2 mg/L or less. Organic-free water is desired for establishing the method blank when measuring OC below 1 mg of carbon per litre. Absolutely carbon free water may not be realistically obtainable and measurement of its carbon level, if any, may be beyond the scope of this test method. However, a working approximation of this goal is the solution contained in the reaction vessel of certain designs of instrument. Alternatively, water that has been acidified, mixed with persulfate to a final concentration of 2 % w/v, heated, or exposed to ultraviolet radiation, or both, and thoroughly sparged (see 9.3) may be used.

8.3 Amber glass bottles should be used to store reagent water, organic-free water, and also standard solutions. See Section 9 for preparation of bottles. It is preferable to dedicate bottles for these uses.

8.4 *Gas Supply*—Use a gas free of $CO₂$ and organic matter, of a purity as specified by the equipment manufacturer. Oxygen is recommended.

8.5 *Organic Carbon, Standard Solution*—Prepare a highconcentration standard using a water-soluble, stable reagent grade compound (see 6.4.2). This stock solution can then be further diluted to a concentration suitable for the method used. For example, to prepare a 2000 mg/L carbon standard of potassium hydrogen phthalate (KHP), note that KHP contains 0.471 g of carbon per gram, so 1 L of standard can be made by 0.471 g of carbon per gram, so 1 L of standard can be made by it may be nece dissolving 2 divided by 0.471, or 4.25 g, of KHP in 1 L of the final fill-to water using a volumetric flask and reagent-grade water.

8.5.1 When preparing low-level standards, keep in mind the sample to about 2. Three diventure of the regional water used for dilutions (see is sufficient to acidify higher than and the sample of the regional water used for OC content of the reagent water used for dilutions (see Appendix X1).

Example 1.69 Confirm this on a set of the same 1.69 Confirm this on a set of the same and the same preview of the same may be used neat or diluted, as required by the manufacturer. Since it is added to the sample, the H_3PO_4 must be of the 77^{11} $^{\circ}$ P₃ highest quality and must be handled carefully to minimize contamination.

8.7 *Persulfate Solution*—Prepare by dissolving an appropriate weight of $K_2S_2O_8$ or $Na_2S_2O_8$ in 1 L of water, to produce the concentration specified by the manufacturer. If specified, add 1 mL of H_3PO_4 (sp gr 1.69) and mix well. Store in a cool, dark location.

NOTE 1—Certain instruments may require carbon to be removed from acid and reagent as completely as possible. See the manufacturer's instructions.

9. Sample Handling

9.1 *Containers and Their Treatment*—Only amber glass bottles with TFE-fluorocarbon-lined bottle closures should be used. Clear glass bottles may be used if protected from sunlight. Where possible, bottles with volumes greater than 200 mL should be used, since small volumes (for example, 10 mL) are proportionately more susceptible to accidental contamination.

9.1.1 Clean sample bottles with chromic acid, rinse several times with water, and dry overnight at 400°C in a muffle furnace. If bottles are new, cleaning with laboratory detergent and rinsing with water may be sufficient, but blank values must be checked.

9.1.2 Rinse the TFE-fluorocarbon-linked closures several times with water, then allow them to soak in water overnight. Rinse these closures again with water before use.

9.1.3 Put the closures loosely on the bottles while the bottles are still warm. When the bottles have cooled to room temperature, tighten the closure.

9.1.4 Follow this cleaning procedure before each re-use of the bottles.

9.2 *Sampling and Preservation*—Collect the sample in accordance with Specification D 1192 and Practice D 3370.

9.2.1 It is recommended that any sample conditioning condensers, coolers, and associated fittings and valves used should be of either stainless steel or TFE-fluorocarbon and be maintained leak-free. When sampling steam condensate, the sample should be at less than 50°C and preferably near ambient temperature. Sampling points should never be in dead-ended portions of the high purity water systems.

9.2.2 Prior to taking a sample, flush the sample line using a continuous flow of high purity water. Sometimes several hours may be required, depending upon the length of the lines and flow rate permitted. Do not readjust flow rate before sampling.

9.2.3 When sampling, rinse and empty the bottle and the closure three times, fill the bottle from the bottom to overflowing and cap the bottle. If OC is to be measured instead of TC, it may be necessary to add H_3PO_4 to the sample bottle before the final fill-to-overflow, depending on the instrument used. Follow the manufacturer's instruction to bring the pH of the sample to about 2. Three drops of acid per 250 mL of sample is sufficient to acidify high purity water to a pH of 2 ± 1 . Confirm this on a separate aliquot.

9.2.4 If the sample cannot be analyzed within 24 h of collection, it should be refrigerated at 4°C in an atmosphere free of organic vapors.

9.3 *Sparging to Remove Inorganic Carbon*—Since, by defiontamination, arcs.iteh.ai/catalog/standards/sist/1db22eda-7 nition, low levels of carbon are to be expected in high-purity water, it is important to remove as much IC as possible before measuring OC. Use a high-efficiency fritted-glass sparger to admit 200 mL/min of carbon-free oxygen or nitrogen to the acidified sample in the original sample bottle. Fit the sparger and a coiled gas vent line into a bottle cap with TFEfluorocarbon-backed liner. This cap should temporarily replace the original bottle cap when sparging is taking place. Continue sparging for at least 5 min. Take care to prevent crosscontamination during transfer of the sparger from one sample to another. Alternatively, such sparging may be done within the instrument.

> 9.4 *Analysis of Sample*—To minimize contamination, convey the sample to the reaction vessel through inert tubing, with volume determined by a sample loop filled and emptied by manual or automatic switching. It is important that the sample not be transferred to another container, or to a syringe.

> 9.4.1 Purge the head space in the sample bottle with pure gas as the sample is withdrawn to prevent contamination by laboratory air. This can most readily be accomplished if the sample pick-up tube, purge line, and gas vent line are all inserted into a bottle cap with TFE-fluorocarbon-backed liner that temporarily replaces the original bottle cap during sample analysis.