



Designation: D3686 – 08

Standard Practice for Sampling Atmospheres to Collect Organic Compound Vapors (Activated Charcoal Tube Adsorption Method)¹

This standard is issued under the fixed designation D3686; the number immediately following the designation indicates the year of original adoption or, in the case of revision, the year of last revision. A number in parentheses indicates the year of last reapproval. A superscript epsilon (ϵ) indicates an editorial change since the last revision or reapproval.

1. Scope

1.1 This practice covers a method for the sampling of atmospheres to determine the presence of certain organic vapors by means of adsorption on activated charcoal using a charcoal tube and a small portable sampling pump worn by a worker. A list of some of the organic chemical vapors that can be sampled by this practice is provided in **Annex A1**. This list is presented as an information guide and should not be considered as absolute or complete.

1.2 This practice does not cover any method of sampling that requires special impregnation of activated charcoal or other adsorption media.

1.3 The values stated in SI units are to be regarded as the standard.

1.4 *This standard does not purport to address all of the safety concerns, if any, associated with its use. It is the responsibility of the user of this standard to establish appropriate safety and health practices and determine the applicability of regulatory limitations prior to use.* A specific safety precaution is given in **9.4**.

2. Referenced Documents

2.1 ASTM Standards:²

D1356 Terminology Relating to Sampling and Analysis of Atmospheres

D3687 Practice for Analysis of Organic Compound Vapors Collected by the Activated Charcoal Tube Adsorption Method

D5337 Practice for Flow Rate Adjustment of Personal Sampling Pumps

¹ This practice is under the jurisdiction of ASTM Committee **D22** on Air Quality and is the direct responsibility of Subcommittee **D22.04** on Workplace Air Quality. Current edition approved Dec. 15, 2008. Published January 2009. Originally approved in 1978. Last previous edition approved in 2001 as D3686 - 95(2001) ^{ϵ 1}. DOI: 10.1520/D3686-08.

² For referenced ASTM standards, visit the ASTM website, www.astm.org, or contact ASTM Customer Service at service@astm.org. For *Annual Book of ASTM Standards* volume information, refer to the standard's Document Summary page on the ASTM website.

2.2 NIOSH Standards:

CDC-99-74-45 Documentation of NIOSH Validation Tests³
HSM-99-71-31 Personal Sampler Pump for Charcoal Tubes; Final Report³

NIOSH Manual of Analytical Methods, Fourth Edition⁴

2.3 OSHA Standards:

29 CFR 1910 Code of Federal Regulations Relating to Labor, Occupational Safety and Health Administration, Department of Labor⁵

OSHA Chemical Sampling Information⁶

OSHA Sampling and Analytical Methods⁷

2.4 UK Health and Safety Executive:⁸

Methods for Determination of Hazardous Substances (MDHS)

2.5 Berufsgenossenschaftliches Institut für Arbeitsschutz (BGIA)⁹

GESTIS Analytical Methods

3. Terminology

3.1 For definitions of terms used in this method, refer to Terminology **D1356**.

3.2 Activated charcoal refers to properly conditioned charcoal.

4. Summary of Practice

4.1 Air samples are collected for organic vapor analysis by aspirating air at a known rate and for an appropriate time through sampling tubes containing activated charcoal.

³ Available from National Technical Information Service (NTIS), 5285 Port Royal Rd., Springfield, VA 22161, <http://www.ntis.gov>.

⁴ NIOSH Manual of Analytical Methods. <http://www.cdc.gov/niosh/nmam/>

⁵ Available from U.S. Government Printing Office Superintendent of Documents, 732 N. Capitol St., NW, Mail Stop: SDE, Washington, DC 20401, <http://www.access.gpo.gov>.

⁶ OSHA Chemical Sampling Information. http://osha.gov/dts/chemicalsampling/toc/toc_chemsamp.html

⁷ OSHA Sampling and Analytical Methods. <http://osha.gov/dts/sltc/methods/toc.html>

⁸ Health Safety Executive. Methods for the Determination of Hazardous Substances (MDHS) guidance. <http://www.hse.gov.uk/pubns/mdhs/index.htm#>

⁹ Berufsgenossenschaftliches Institut für Arbeitsschutz. GESTIS Analytical methods. http://www.hvbg.de/e/bia/gestis/analytical_methods/index.html

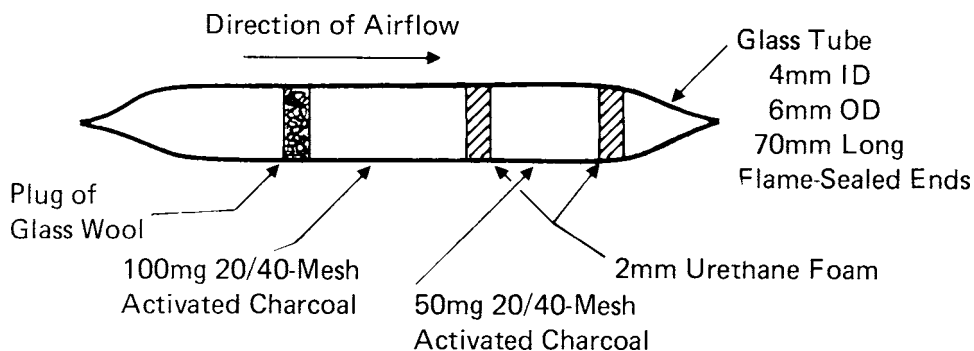


FIG. 1 Activated Charcoal Adsorption Sampling Tube

4.2 Instructions are given to enable assembly of charcoal tubes suitable for sampling purposes.

4.3 Information on the correct use of the charcoal tube sampling device is presented.

4.4 Practice D3687 describes a practice for the analysis of these samples.

5. Significance and Use

5.1 Promulgations by the U.S. Occupational Safety and Health Administration (OSHA) in 29 CFR 1910.1000 designate that certain organic compounds must not be present in workplace atmospheres at concentrations above specific values.

5.2 This practice, when used in conjunction with Practice D3687, will provide the needed accuracy and precision for the determination of airborne time-weighted average concentrations of many of the organic chemicals cited in CDC-99-74-45, HSM-99-71-31, NIOSH Manual of Analytical Methods, 29 CFR 1910.1000, OSHA Chemical Sampling Information, OSHA Sampling and Analytical Methods, HSE Methods for the Determination of Hazardous Substances, and BGIA GETIS Analytical Methods.

5.3 A partial list of chemicals for which this method is applicable is given in Annex A1, along with their OSHA permissible exposure limits.

6. Interferences

6.1 Water mist and vapor can interfere with the collection of organic compound vapors. Humidity greater than 60 % can reduce the adsorptive capacity of activated charcoal up to 50 % for some chemicals (1).¹⁰ Presence of condensed water droplets in the sample tube will indicate a suspect sample.

6.2 High levels of organic vapors can interfere by reducing the amount of time a workplace can be sampled before the charcoal sampler becomes saturated.

7. Apparatus

7.1 Charcoal Tube:

7.1.1 A sampling tube consists of a length of glass tubing usually containing two sections of activated charcoal that are held in place and separated by nonadsorbent material. The tube is sealed at each end.

7.1.1.1 Sampling tubes are commercially available. The tubes are usually divided into two sections with the front section containing 100 to 800 mg of activated charcoal and the back section containing 50 to 400 mg of activated charcoal. The 100/50-mg tube ((2-4) and Fig. 1) is the one most frequently used, it consists of a glass tube that is 70-mm long, 6-mm outside diameter, 4-mm inside diameter, and contains two sections of 20/40 mesh-activated coconut-shell charcoal separated by a 2-mm section of urethane foam. The front section of 100 mg is retained by a plug of glass wool, and the back section of 50 mg is retained by either a second 2-mm portion of urethane foam or by a plug of glass wool. Both ends of the tube are usually flame-sealed.

NOTE 1—Urethane foam is known to adsorb certain pesticides (5). Contaminated urethane foam should not be used for this practice.

7.1.1.2 When it is desirable to sample highly volatile compounds for extended periods, or at a high volume flow rate, a larger device capable of efficient collection can be used, provided the proportions of the tube and its charcoal contents are scaled similarly to the base dimensions to provide nominally the same linear flow rate and contact time with the charcoal bed.

7.1.2 The back portion of the sampler tube usually contains 50 % of the mass of activated charcoal present in the front section. The back section adsorbs vapors that penetrate the front section and serves as a warning that breakthrough may have occurred. (Annex A1 gives recommended maximum tube loading information for many chemicals.)

7.1.3 The adsorptive capacity and extraction efficiency (also called desorption efficiency) of different batches of activated charcoal can vary. Commercial tubes, if used, should be purchased from the same batch and in sufficient number to provide sampling capability for an adequate period of time. Care must be taken to have enough tubes from the same batch for a given study.

7.1.4 Pressure drop across the sampling tube should be less than 25 mm Hg (3.3 kPa) at a flow rate of 1000 mL/min and less than 4.6 mm Hg (0.61 kPa) at a flow rate of 200 mL/min.

¹⁰ The boldface numbers in parentheses refer to the list of references at the end of this standard.

7.1.5 Charcoal sampling tubes prepared in accordance with this practice and with sealed glass ends can be stored indefinitely. Accrediting bodies, such as American Industrial Hygiene Association, may require that sampling media have an expiration date. Accredited facilities must abide by this date.

7.2 Sampling Pumps:

7.2.1 Any pump with a flow rate that can be accurately determined, that can be set at the desired sampling rate, and that can maintain the desired sampling rate for a sufficient time is suitable. Primarily though, this practice is intended for use with small personal sampling pumps.

7.2.2 Pumps having stable low flow rates (10 to 200 mL/min) are preferable for long period sampling (up to 8 h) or when the concentration of organic vapors is expected to be high. Reduced sample volumes will avoid exceeding the adsorptive capacity of the charcoal tubes. (Suggested flow rates and sampling times are given in [Annex A1](#) for anticipated concentration ranges.) (Sample volumes are discussed in [9.5](#).)

7.2.3 Pumps are available that will provide stable flow rates between $\pm 5\%$ of the desired flow rate. Pumps should be calibrated before and after sampling.

7.2.4 All sampling pumps must be carefully calibrated with the charcoal tube device in the proper sampling position.

7.2.5 A sampling tube holder with flexible tubing is used to connect the sampling tube to the sampling pump. The sampling tube holder is used to protect the worker from the sharp end of the sampling tube.

8. Reagents

8.1 *Activated Charcoal*—Prior to being used to make sampling devices the charcoal should be heated in an inert gas at an appropriate temperature for a sufficient time. Commercially available coconut-shell charcoal (20/40 mesh) has been found to have adequate adsorption capacity for many volatile chemicals. Other charcoals, such as JXC charcoal, petroleum-based charcoal, and proprietary charcoals, can be used for appropriate applications.

9. Sampling with Activated Charcoal Samplers

9.1 *Calibration of the Sampling System*—Calibrate the sampling system, including pump, flow regulator, tubing to be used, and a representative charcoal tube (or an equivalent induced resistance) with a primary flow-rate standard or with a calibrated secondary standard to within $\pm 5\%$ of the desired flow rate as described in Practice [D5337](#). Calibrate the sampling pump in a clean location with similar temperature and barometric pressure as the sampling site. Normally, calibrated pump flow rates or sample air volumes are not corrected for temperature or barometric pressure.

9.2 Break open both ends of the charcoal tube to be used for sampling, ensuring that each opening is at least one half the inside diameter of the tube.

9.3 Insert the charcoal tube into the connective flexible tubing, placing the back-up section nearest to the pump. At no time should there be any tubing ahead of the sampling tubes. Use a sampling tube holder to protect the worker from the sharp end of the sampling tube.

9.4 For a breathing zone sample, fasten the sampling pump to the worker, and attach the sampling tube as close to the worker's breathing zone as possible. Position the tube in a vertical position to avoid channeling of air through the charcoal sections. (**Warning**—Assure that the presence of the sampling equipment is not a safety hazard to the worker and that the equipment will not interfere with the worker's duties.)

9.4.1 Turn on the pump.

9.4.2 Record the flow rate, the starting time, and depending on the make of pump used, the register reading.

9.5 *Sampling Volumes*—The minimum sample volume will be governed by the detection limit of the analytical method, and the maximum sample volume will be determined by either the adsorptive capacity of the charcoal or limitations of the pump battery.

9.5.1 One method of calculating required sample volumes is to determine first the concentration range, over which it is important to report an exact number, for example from 0.2 to 2 times the permissible exposure concentration, and then calculate the sample volumes as follows:

$$\text{Minimum sample volume, m}^3 = \quad (1)$$

$$\frac{\text{minimum detection limit, mg}}{0.2 \times \text{permissible exposure limit, mg/m}^3}$$

$$\text{Maximum sample volume, m}^3 = \quad (2)$$

$$\frac{\text{tube capacity for vapors, mg}}{2 \times \text{permissible exposure limit, mg/m}^3}$$

9.5.2 Select a sampling rate that, in the sampling time desired, will result in a sample volume between the minimum and maximum calculated in [9.5.1](#).

9.5.2.1 Generally a long sampling time at a low flow rate is preferable to short-term, high-volume sampling. This is consistent with the fact that most health standards are based on 8-h/day time-weighted averages of exposure concentrations. Often, two 4-h samples are preferable to a single 8-h sample so that if one is lost then the other can be used to partially document exposure. Work practices may change during the day and be better documented with two samples.

9.5.2.2 A sample flow rate of less than 10 mL/min, however, should not be used. Calculations based upon diffusion coefficients for several representative compounds indicate that sampling at less than 10 mL/min may not give accurate results.¹¹

9.5.2.3 Sampling information for a large number of organic chemicals is given in Annex A1, in the NIOSH Manual of Analytical Methods, OSHA Chemical Sampling Information file and methods, UK HSE MDHS database, and German BGIA GESTIS Analytical Methods database. Other appropriate sources of information and guidance, including Annex A1, can also be used.

9.5.3 When spot checks are being made of an environment, a sample volume of 10 L is adequate for determining vapor concentrations in accordance with exposure guidelines. Particularly volatile organic chemicals may require a lesser sample

¹¹ Heitbrink, W. A., "Diffusion Effects Under Low Flow Conditions," American Industrial Hygiene Association Journal, Vol 44, No. 6, 1983, pp. 453-462.