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Živila - Smernice za kalibracijo ter kvantitativno določanje ostankov pesticidov in organskih onesnaževal z uporabo kromatografskih metod

Foodstuffs - Guidelines for the calibration and quantitative determination of pesticide residues and organic contaminants using chromatographic methods

Lebensmittel - Anleitung zur Kalibrierung und quantitativer Bestimmung von Pflanzenschutzmittelrückständen und organischen Kontaminanten mit chromatographischen Verfahren **Standards.iten.ai**)

Produits alimentaires - Lignes directrices pour l'étalonnage et le dosage des résidus de

pesticides et contaminants organiques par des méthodes chromatographiques

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67.050 Splošne preskusne in analizne metode za živilske proizvode

General methods of tests and analysis for food products

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Foodstuffs - Guidelines for the calibration and quantitative determination of pesticide residues and organic contaminants using chromatographic methods

Produits alimentaires - Lignes directrices pour l'étalonnage et le dosage des résidus de pesticides et contaminants organiques par des méthodes chromatographiques Lebensmittel - Anleitung zur Kalibrierung und quantitativer Bestimmung von Pflanzenschutzmittelrückständen und organischen Kontaminanten mit chromatographischen Verfahren

This Technical Specification (CEN/TS) was approved by CEN on 14 July 2019 for provisional application.

The period of validity of this CEN/TS is limited initially to three years. After two years the members of CEN will be requested to submit their comments, particularly on the question whether the CEN/TS can be converted into a European Standard.

CEN members are required to announce the existence of this CEN/TS in the same way as for an EN and to make the CEN/TS available promptly at national level in an appropriate form. It is permissible to keep conflicting national standards in force (in parallel to the CEN/TS) until the final decision about the possible conversion of the CEN/TS into an EN is reached.

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European foreword

This document (CEN/TS 17061:2019) has been prepared by Technical Committee CEN/TC 275 "Food analysis - Horizontal methods", the secretariat of which is held by DIN.

Attention is drawn to the possibility that some of the elements of this document may be the subject of patent rights. CEN shall not be held responsible for identifying any or all such patent rights.

This document will supersede CEN/TS 17061:2017.

Compared to CEN/TS 17061:2017, the following changes have been made:

- Annex A (informative) containing a list of abbreviations was added;
- The document has been editorially revised.
- Annex A (informative) contains a list of abbreviations.

According to the CEN/CENELEC Internal Regulations, the national standards organisations of the following countries are bound to announce this Technical Specification: Austria, Belgium, Bulgaria, Croatia, Cyprus, Czech Republic, Denmark, Estonia, Finland, France, Germany, Greece, Hungary, Iceland, Ireland, Italy, Latvia, Lithuania, Luxembourg, Malta, Netherlands, Norway, Poland, Portugal, Republic of North Macedonia, Romania, Serbia, Slovakia, Slovenia, Spain, Sweden, Switzerland, Turkey and the United Kingdom.

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1 Scope

This Technical Specification gives guidelines for the execution of calibration and quantitative evaluation of chromatographic procedures for the determination of pesticides and organic contaminants in residue analysis. In addition, the essential requirements for calibration are outlined.

The calibration of analytical procedures and the evaluation of analytical results need to be conducted according to uniform principles in order to allow for a comparison of analytical results (even from different analytical procedures). They constitute the basis of any method validation and of the quality assurance within laboratories [1], [2], [3].

This Technical Specification does not consider issues of identification/qualification and extraction efficiency.

2 Normative references

There are no normative references in this document.

3 Terms and definitions

No terms and definitions are listed in this document.

ISO and IEC maintain terminological databases for use in standardization at the following addresses:

- IEC Electropedia: available at http://www.electropedia.org/
- ISO Online browsing platform: available at <u>https://www.iso.org/obp</u>
 - (standards.iteh.ai)

4 Principle

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This document describes the stapproach aforathe tacalibration 1069 chromatographic procedures. The following types of calibration are discussed in more detail:n-ts-17061-2019

- external calibration with linear calibration function;
- external calibration with quadratic calibration function;
- calibration with internal standard and linear calibration function;
- calibration with internal standard and quadratic calibration function;
- calibration with standards labelled with stable isotopes (isotopic dilution analysis);
- standard addition to final extract;
- standard addition to sample.

For this purpose, the calibration function and the selection criteria are illustrated on the basis of examples. The calculation formulae refer to the final extract ready for analysis ("test solution").

The description is rounded off by essential items of quality assurance, e.g. the qualification of chromatographic systems or the quality control chart.

5 General

Calibration of a system is understood as the determination of a functional relationship between a measurable quantity and a concentration to be determined. The chosen type of calibration depends on

the various analytical problems/tasks. It is performed in connection with the respective series of measurements.

Basic calibration is regarded as the determination of the functional relationship when an analyte is to be determined for the first time by means of a particular measurement system.

Depending on the problem and on the type of reference solution used, it is distinguished between:

- calibration with external standard;
- calibration with internal standard;
- calibration with standard addition;
- *calibration of the entire procedure.*

In case of the calibration with external standard, the calibration solutions can be prepared either with a pure solvent (standards in solvent) or with sample extracts which evidently do not contain significant amounts of the analyte(s) (matrix-matched standards).

The application of a simple linear-regression calculation requires a linear relationship between the content of substance and the measured value. The linearity test can be performed visually and/or mathematically. A mathematical check is performed, e.g. by means of the goodness-of-fit test according to Mandel or by means of residual analysis. The residuals are the deviations of the measurement values from the values predicted by the regression line (see 9.1, Example 1).

6 Execution and calculation of calibrations

6 Execution and calculation of calibrations (standards.iteh.ai)

6.1 General/specifications

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6.1.1 Working ranges://standards.iteh.ai/catalog/standards/sist/85107909-18eb-450c-850a-

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The range of measurement represented by the lowest and the highest calibration point constitutes the range of concentrations for which the determined calibration function applies (working range). Only within this range, the measured values are reliable and, therefore, can be used for the calculation of analyte contents. At the upper and lower end, the prediction interval becomes wider i.e. the measurement error increases progressively. The highest precision is found in the middle of the working range [4].

The detector response from the analytes in the sample extract has to lie within the working range. Extracts containing residues above the calibrated range shall be diluted. If the calibration solutions are matrix-matched the matrix concentration in the calibration standards should also be diluted, see [5].

The calibration range shall be adjusted to the respective residue concentrations in the test solution (real-sample concentrations which often occur in practice) and should cover a maximum of two orders of magnitudes. Where appropriate, several calibration functions shall be established by means of calibration solutions.

The lower limit of the practical working range usually represents the lowest calibration level, see [5]. It shall be equal to or lower than the Reporting Limit (RL). The RL may not be lower than the Limit of Quantification (LOQ).

6.1.2 Number of calibration points

6.1.2.1 General

For the working range of calibration functions, calibration solutions with different concentrations of pesticides or contaminants are prepared (depending on the requirement, three to five calibration

points), the concentrations of which are as equidistantly distributed over the working range as possible. The concentrations shall start at the lower limit of the practical working range. If the working range has to cover one order of magnitude, three calibration points are necessary, while five calibration points are necessary for two orders of magnitude (depending on the covered concentration range, e.g. 1, 3, 10, 30 and 100 times the lowest calibrated concentration).

6.1.2.2 Acceptability of single-point calibrations

A single-point calibration is sufficient if the linearity of the calibration function has been checked over a longer period of time and has been evaluated as stable and if the blank values as well as the intercept are negligibly small. The concentration level should be in the upper fraction of the working range. The analyte concentrations in the calibration and test solutions should be within the range proposed DG-SANTE, if the test solution is compared to one calibration solution only (see [5] for more details). However, a check of the basic calibration shall be made every working day, and the measurement of a minimum number of representative analytes is indispensable, see [5].

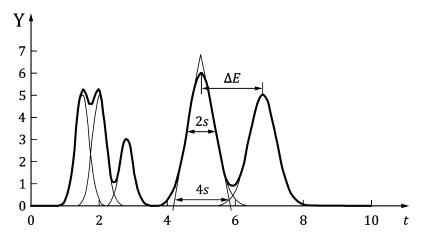
6.1.3 Permissible quantities for determination of response (peak areas or peak heights and peak ratio, respectively)

The analyte signal, thus the peak, produced by the detector can be quantitatively evaluated through determination of the peak height or peak area. By principle, the height as well as the area of the peak depends on the analyte concentration and mass, respectively. The peak height indicates the distance from the baseline to the maximum of the peak. In case of well resolved peaks, the peak height is proportional to the analyte concentration. The evaluation by means of peak height should only be performed in case of reproducible peak shape and constant width at half-height (half-width). It leads to difficulties if there are two peaks with a poorer resolution than approximately 1,25.

A resolution *R* can be defined by means of the distance ΔE and the width $4 \times s$ of the peaks, see Figure 1 and Formula 1. The width $4 \times s$ is determined through the intercept of both inflexional tangents on the baseline or calculated from the standard deviation *s*.

$$R = \frac{\Delta E}{4 \times s}$$
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407b2e184dea/sist-ts-cen-ts-17061-2019 (1)

In case of well resolved peaks, the peak area is proportional to the analyte concentration. In contrast to the peak height, the peak area usually provides accurate results, even for asymmetric peaks. The prerequisite for peak-area determinations is always the precise definition of the baseline.



Key

- *t* retention time
- Y intensity

Figure 1 — Definition of the resolution of peaks

The calculation by means of the intensity ratios of peaks (peak ratio) is used for calibrations with internal standard (ISTD), e.g. for EN 15662 (QuEChERS) and for calibrations with internal standard labelled with stable isotopes (stable-isotope labelled standards). This procedure of calibration and calculation requires to know at least the ratio of amounts of internal standard added to test samples (or to extract aliquots) and to calibration standards.

6.1.4 Stability of calibration functions

The calibration standards should be injected at least at the start and end of a sample sequence (bracketing calibration). Bracketed samples containing pesticide residues or organic contaminants should be re-analysed if the drift between the two bracketing injections exceeds the limit given by DG-SANTE. In general such bracketed samples have to be re-analysed if the calibration level corresponding to the RL was not measurable throughout the batch, see [5] for more details.

6.2 Calibration functions

6.2.1 Selection of appropriate calibration function

Calibration functions can be linear, logarithmic, exponential as well as polynomial of 2nd order.

Whenever possible, the simplest acceptable calibration function should be used. The use of linear weighted regression (e.g. 1/x or $1/x^2$ weighting) is recommended.

A calibration function is a unique plotting of the set of all *x*-values (concentration or mass) against the set of all *y*-values (peak area or height), i.e. exactly one *y*-value is assigned to each *x*-value.

Before determining the established type of calibration function and testing the linearity of a calibration function, the homogeneity of variances should be checked first as it represents a basic prerequisite for the applicability of statistical methods.

For this purpose, calibration solutions are prepared where between three and ten concentration levels with two to six measurements periconcentration are recommended of ten standard samples of the lowest and the highest working range concentration are separately analysed at a time. The variation (scatter) of the measured values at the limits of the working range is tested for significant differences by means of a simple F-test for variance inhomogeneity.

In case that the F-test indicates a significant difference of variances, there are three opportunities to proceed:

- selection of a narrower working range;
- application of weighted regression;
- application of multiple curve fitting.

The non-consideration of a present inhomogeneity of variances results in a wider prediction interval so that an analytical result which has been determined accordingly shows a higher uncertainty of measurement.

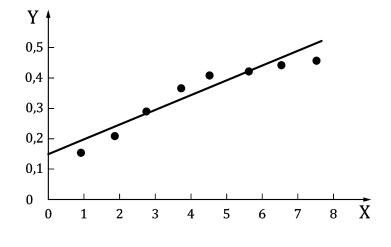
The linearity of the calibration function can be mathematically checked by means of a linearity test. For this purpose, the goodness-of-fit test according to Mandel or the residual analysis is appropriate. In every case, a graphical representation (plot) of the calibration data are recommended (see 9.2, Example 2).

6.2.2 Visual linearity test

In the simplest case, the determination of the type of calibration function is performed by means of graphical representation of the calibration data including the calibration line and a subjective assessment. If this indicates an obvious nonlinearity of the measured values (see Figure 2), a separate

statistic linearity test can be omitted. In cases of doubt, however, the linearity should be checked mathematically [5].

Calibration curves (graphs) shall not be forced through the zero point (origin). Further examples can be found in Clause 9 on Examples.



Кеу

X concentration

Y measured value

Figure 2 - Graphical representation of calibration data (standards.iteh.ai)

6.2.3 Mathematical check of linearity

Within the laboratory, the mathematical check of linearity is of secondary importance. Regarding the execution of linearity tests refer to the Examples 1 and 2 and to [6], 2019

As an important criterion of linearity, the coefficient of determination, R^2 , is well accepted. However, the coefficient of determination does not allow a sufficient statement regarding the statistical significance of the linear relationship. Calculation of the residuals is recommended to avoid overreliance on coefficients of determination. If individual residuals exceed an acceptable level as defined by DG-SANTE, an alternative calibration function shall be used according to [5].

6.2.4 Calibration with interpolation functions

Several methods of measurement show a basically nonlinear relationship between the measured signals and the concentration and the amount of analyte, respectively. In cases where the linear regression is inappropriate, a polynomial of 2nd order (quadratic calibration function) is usually fitted to the calibration data.

Like the linear calibration function, the polynomial of 2nd order has a prediction interval. Nevertheless, the functional equation has a higher degree of complexity which is why this procedure needs computer support for evaluation.

In case of the nonlinear calibration function, the number of necessary calibration points depends on the desired accuracy and on the reasonable effort. For a global interpolation function, a single mathematical equation describes the entire calibration. However, this is only possible in exceptional cases. In most cases, such a function does not show the expected properties between the calibration points (e.g. polynomial "oscillation"). Therefore, local interpolation functions are employed where each interval between two interpolation nodes is characterized by a separate interpolation function. The application of the calibration function requires the selection of the corresponding local interpolation function for each signal value.

In case of interpolation with polynomials calculated from calibration points in proximity, the curvature of the calibration graph between the interpolation nodes can also be taken into account. The computational effort increases especially when determining the inverse function. Therefore, the calibration function is often calculated once the *x*-values and *y*-values have been interchanged.

The calibration function piecewise defined by polynomials will show break (knee) points between the interpolation nodes. With further increased computational effort, this can be prevented by means of spline functions which are also piecewise defined. If derivations are needed, these, starting from a certain differentiation level, will become discontinuous.

6.3 Test for matrix effects

The test for matrix effects is performed by means of a comparison of the calibration lines from the calibration with standards in a pure solvent and with standards in a matrix (matrix-matched standards) (see Figure 11). Significantly higher or lower signals in matrix calibration indicate matrix effects (see 9.3, Example 3).

Matrix-matched standards can be prepared in the same way as standards in solvents. Typically, the solvent used to fill up is replaced by an extract of blank samples (residue-free material), that is prepared according to the method used for the routine sample preparation (generally without use of an internal standard). The matrix-matched standards prepared in this way shall contain $\ge 80 \%$ (V/V) of the test solution of control samples in order to obtain comparable matrix effects of samples and standards during GC- or HPLC-MS(/MS) analysis. Blank samples of the same commodities as the test sample (apple for apple samples etc.) are used to optimally compensate for matrix effects.

The stability of matrix-matched standards can be lower than that of standards in solvents and, therefore, shall be controlled periodically. ards.iteh.ai)

If the quantitative evaluation of a peak is impaired by peaks of co-extracted accompanying substances (e.g. peak overlaps), an extract solution should be injected for comparison which contains the corresponding accompanying substance in the test solution of the same but residue-free test material.

6.4 Basic calibration and calibration by means of external standard

6.4.1 Basic calibration

The basic calibration serves as a qualification test for the measurement system used, i.e. no steps of sample preparation (treatment) such as extraction or digestion are undertaken in the course of it, but only standard solutions are analysed.

To perform the basic calibration, equal volumes of calibration solutions with appropriate concentrations are injected and the peak areas/heights obtained are plotted against the concentrations of the calibration solutions (see 9.4, Example 4). Evaluation is carried out with the most suitable calibration function. It is important that a continuous and reproducible relationship between signal intensity and analyte concentration is obtained.

A validated calibration function of the basic calibration can be also used for quantification with external standards if it has been proven before that the matrix does not cause a significant increase or decrease of the analyte signal.

6.4.2 Calculation by means of external standard in case of linear calibration function without significant ordinate intercept

The quantitative evaluation is done by determination of the peak areas (or peak heights) and comparison with the peak areas (or peak heights) of analyte solutions with known concentrations. In the course of this, equal volumes of the test solutions and calibration solutions are injected into the chromatographic column. If the signal of the analyte is within the linear range and if the straight line

intercepts the ordinate in the range of the origin (of coordinates), the analyte concentration in the test solution can be calculated according to the following very simple Formula (2):

$$\rho_{\rm A} = \frac{y_{\rm A}}{b} \tag{2}$$

where

- ρ_{A} is the mass concentration of the analyte in the test solution [e.g. μ g/ml];
- *y*_A is the peak area (or peak height) of the analyte measured in the test solution [e.g. counts];
- *b* is the slope of the calibration function [e.g. counts \cdot ml/µg].

6.4.3 Calculation by means of external standard using linear calibration function

If a significant ordinate intercept has been calculated for the calibration function but the analyte signal is within the linear range, the analyte concentration in the test solution is calculated by rearranging the established calibration function according to Formula (3):

$$\rho_{\rm A} = \frac{\left(y_{\rm A} - c\right)}{b} \tag{3}$$

where

- ρ_{A} is the mass concentration of the analyte in the test solution [e.g. µg/m];
- $y_{\rm A}$ is the peak area (or peak height) of the analyte in the test solution [e.g. counts];
- *b* is the slope of the calibration function [e.g. counts \cdot m]/µg];
- *c* is the ordinate intercept of the calibration function/[eig.7counts]-450c-850a-

6.4.4 Calculation by means of external standard using quadratic calibration function

A quadratic calibration function can be used if the distribution of residuals or other mathematical tests indicate that a linear calibration function is inappropriate. For a calculation of the analyte concentration in the test solution in case of quadratic calibration function, the rearrangement of the equation theoretically yields two solutions of which only one makes analytical sense, see Formula (4):

$$\rho_{\rm A} = \frac{-b \pm \sqrt{b^2 - 4a(c - y_{\rm A})}}{2a}$$
(4)

where

- ρ_A is the concentration of the analyte in the test solution [e.g. μ g/ml];
- *y*_A is the peak area (or peak height) of the analyte in the test solution [e.g. counts];
- *a* is the slope of the calibration function in the quadratic term of the equation [e.g. $counts \cdot ml^2/\mu g^2$];
- *b* is the slope of the calibration function in the linear term of the equation [e.g. counts \cdot ml/µg];
- *c* is the ordinate intercept of the calibration function [e.g. counts].

6.4.5 Calculation by means of external standard using nonlinear calibration function or weighted regression

If the execution of calibrations with nonlinear (e.g. exponential or logarithmic) calibration function becomes necessary, one will usually rely on appropriate instrument software for calculating the analyte concentration in the final extract. Before using such software, it shall be checked with suitable self-calculated examples.

The same applies to the calculation of analyte content using a calibration function which has been obtained by means of weighted regression.

6.5 Calculation by means of internal standard

6.5.1 General

The internal standard shall neither belong to the substance group to be analysed nor be present in the sample. It shall have sufficient signal intensities and needs to be separately detectable.

For one analysis, several ISTDs can be used in order to detect and, if appropriate, compensate various errors which occur during preparation (treatment) and measurement. The ISTD solution can be added at different points of the analytical course and for various reasons [7]:

- before extraction and sample digestion, respectively, to compensate for all errors in the entire procedure (correction standard for the calculation of analyte concentration, i.e. classic ISTD for quantification);
- before extraction and sample digestion, respectively, only to control the recovery (single steps of preparation) without making any correction (quality-assurance standard or surrogate standard);
- after the preparation has been completed and before the instrumental measurement, for the compensation of sensitivity fluctuations of the instrument and variations of injection volumes during the measurement sequence (injection standard) and for the calculation of the recovery (rates) of correction standards, respectively.

Hereafter, only the application as correction standard for the entire procedure is considered. This application is only reasonable if the ISTD, in comparison to the analyte, shows nearly the same behaviour during preparation and detection. The use of stable-isotope labelled compounds as ISTD is particularly advantageous.

The identification of the residues contained in the test solution is first carried out by means of their relative retention times related to the retention time of the respective ISTD. But before that, the calibration line needs to be determined using ISTD addition as well. For calibration and for testing the linearity of the measurement signal, the ratio of the peak areas of analyte and ISTD is, separately for each agent, plotted against the concentration ratio of analyte and ISTD in the respective calibration solution (see 9.5, Example 5).

Possible errors:

If an internal standard is used for the calculation of residues, its signal intensity influences all quantified contents and, consequently, the accuracy of analyses. Unwanted factors can change the intensity of the internal standard and, thereby, lead to errors in the calculation of residue contents. Thus, losses of internal standard during sample purification or the selective signal suppression (matrix effect) give rise to overestimation of residue contents. To the contrary, the contents of all detected analytes are underestimated as a consequence of signal enhancement of ISTD due to matrix effects. Such matrix effects are caused by coeluting sample components and usually occur to an increasing degree only for certain matrices.