
**Horizontal methods for molecular
biomarker analysis — Methods
of analysis for the detection of
genetically modified organisms and
derived products —**

**Part 5:
Real-time PCR based screening
method for the detection of the FMV
promoter (P-FMV) DNA sequence**

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*Méthodes horizontales d'analyse moléculaire de biomarqueurs —
Méthodes d'analyse pour la détection des organismes génétiquement
modifiés et des produits dérivés —*

*Partie 5: Méthode de dépistage PCR en temps réel pour la détection de
la séquence ADN du promoteur FMV (P-FMV)*



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Foreword

ISO (the International Organization for Standardization) is a worldwide federation of national standards bodies (ISO member bodies). The work of preparing International Standards is normally carried out through ISO technical committees. Each member body interested in a subject for which a technical committee has been established has the right to be represented on that committee. International organizations, governmental and non-governmental, in liaison with ISO, also take part in the work. ISO collaborates closely with the International Electrotechnical Commission (IEC) on all matters of electrotechnical standardization.

The procedures used to develop this document and those intended for its further maintenance are described in the ISO/IEC Directives, Part 1. In particular the different approval criteria needed for the different types of ISO documents should be noted. This document was drafted in accordance with the editorial rules of the ISO/IEC Directives, Part 2 (see www.iso.org/directives).

Attention is drawn to the possibility that some of the elements of this document may be the subject of patent rights. ISO shall not be held responsible for identifying any or all such patent rights. Details of any patent rights identified during the development of the document will be in the Introduction and/or on the ISO list of patent declarations received (see www.iso.org/patents).

Any trade name used in this document is information given for the convenience of users and does not constitute an endorsement.

For an explanation on the meaning of ISO specific terms and expressions related to conformity assessment, as well as information about ISO's adherence to the World Trade Organization (WTO) principles in the Technical Barriers to Trade (TBT) see the following URL: www.iso.org/iso/foreword.html.

The committee responsible for this document is ISO/TC 34, *Food products*, Subcommittee SC 16, *Horizontal methods for molecular biomarker analysis*.

A list of all the parts in the ISO/TS 21569 series can be found on the ISO website.

Horizontal methods for molecular biomarker analysis — Methods of analysis for the detection of genetically modified organisms and derived products —

Part 5:

Real-time PCR based screening method for the detection of the FMV promoter (P-FMV) DNA sequence

1 Scope

This document specifies a procedure for the detection of a DNA sequence used in genetically modified (GM) plants by means of a real-time PCR (polymerase chain reaction). The method detects a 78 base pairs long segment of the *Figwort mosaic virus* 34S promoter DNA sequence. This segment in some GM plants is indicated as FMV promoter (P-FMV) and in other GM plants as FMV enhancer (E-FMV).

The method was developed and validated for the analysis of DNA extracted from foodstuffs. It may be suitable also for analysis of other products such as feedstuffs and seeds. The procedure requires the extraction of an adequate quantity and quality of amplifiable DNA from the test sample.

The DNA sequence amplified by the P-FMV element-specific method can be detected in samples which contain DNA of the naturally occurring *Figwort mosaic virus*. For this reason, it is necessary to confirm a positive screening result. Further analyses are required using construct-specific or event specific methods.

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2 Normative references

The following documents are referred to in the text in such a way that some or all of their content constitutes requirements of this document. For dated references, only the edition cited applies. For undated references, the latest edition of the referenced document (including any amendments) applies.

ISO 21569, *Foodstuffs — Methods of analysis for the detection of genetically modified organisms and derived products — Qualitative nucleic acid based methods*

ISO 21570, *Foodstuffs — Methods of analysis for the detection of genetically modified organisms and derived products — Quantitative nucleic acid based methods*

ISO 21571:2005, *Foodstuffs — Methods of analysis for the detection of genetically modified organisms and derived products — Nucleic acid extraction*

ISO 24276, *Foodstuffs — Methods of analysis for the detection of genetically modified organisms and derived products — General requirements and definitions*

3 Terms and definitions

For the purposes of this document, the terms and definitions given in ISO 16577 apply.

ISO and IEC maintain terminological databases for use in standardization at the following addresses:

- IEC Electropedia: available at <http://www.electropedia.org/>
- ISO Online browsing platform: available at <http://www.iso.org/obp>

4 Principle

DNA is extracted from the test portion applying a suitable method (see ISO 21571). The DNA analysis consists of two parts:

- verification of the amount, quality and amplifiability of the extracted DNA, e.g. by a taxon-specific PCR assay (according to ISO 21569 and ISO 21570), see also Reference [1];
- detection of the P-FMV DNA sequence in a real-time PCR, see Reference [2].

5 Reagents and materials

5.1 General

For the purpose of this document, only chemicals and water of recognized analytical grade, appropriate for molecular biology shall be used. Unless stated otherwise, solutions should be prepared by dissolving the corresponding reagents in water and be autoclaved. For all operations for which gloves are used it should be ensured that these are powder-free. The use of aerosol protected pipette tips (protection against cross contamination) is recommended.

5.2 PCR reagents

5.2.1 Thermostable DNA polymerase (for hot-start PCR).

5.2.2 PCR buffer solution (containing magnesium chloride and deoxyribonucleoside triphosphates dNTPs).

Ready-to-use reagent mixtures or mixtures of individual components can be used. Reagents and polymerases which lead to equal or better results may also be used.

5.2.3 Oligonucleotides (see Table 1)¹⁾.

Table 1 — Oligonucleotides

Name	DNA sequence of the oligonucleotide	Final concentration in PCR
P-FMV as the target sequence (GeneBank accession number X06166[2],[3]):		
pFMV-F	5'-CAA AAT AAC GTG GAA AAG AGC T-3'	340 nmol/l
pFMV-R	5'-TCT TTT GTG GTC GTC ACT GC-3'	340 nmol/l
Probe pFMV	5'-(FAM)-CTG ACA GCC CAC TCA CTA ATG C-(BHQ1)-3' ^a	120 nmol/l
^a FAM: 6-Carboxyfluorescein, BHQ-1: Black Hole Quencher® 1 (non-fluorescent chromophore). This information is given for convenience of users of this document and does not constitute an endorsement by ISO of the product named. Equivalent products from other manufacturers may be used if they can be shown to give equivalent or better results.		

6 Apparatus

Requirements concerning apparatus and materials shall be according to ISO 21569. In addition to the usual laboratory equipment, the following equipment is required.

6.1 Real-time PCR device, suitable for the excitation of fluorescent molecules and the detection of fluorescence signals generated during PCR.

1) In the interlaboratory trial performed for P-FMV, participants were provided with dried aliquots (per 50 reactions) of primer/probe -mixes (to be stored in dark until the start of the interlaboratory trial). Per aliquot 375 µl PCR grade water was added and allowed to settle.

7 Procedure

7.1 Preparation of test sample

It should be ensured that the test sample used for DNA extraction is representative of the laboratory sample, e.g. by grinding or homogenizing of the laboratory sample. Measures and operational steps to be taken into consideration should be according to ISO 21571 and ISO 24276.

7.2 Preparation of DNA extracts

Concerning the preparation of DNA from the test portion the general instructions and measures described in ISO 21571 shall be followed. It is recommended to choose one of the DNA extraction methods described in ISO 21571:2005, Annex A.

7.3 PCR setup

The method described applies for a total volume of 25 µl per PCR. The reaction setup is given in [Table 2](#).

Reagents are completely thawed at room temperature. Each reagent should be carefully mixed and briefly centrifuged immediately before pipetting. A PCR reagent mixture is prepared which contains all components except for the sample DNA. The required amount of the PCR reagent mixture depends on the number of reactions to be performed, including at least one additional reaction as a pipetting reserve. Add 5 µl of sample DNA to each reaction.

Table 2 — Reaction setup for the amplification
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Total reaction volume	25 µl
Sample DNA (up to 200 ng) or controls	5 µl
PCR buffer solution ^a (including MgCl ₂ , dNTPs and hot-start DNA polymerase)	12,5 µl
Primer pFMV-F and pFMV-R	see Table 1
Probe pFMV	see Table 1
Water	to 25 µl
^a In the collaborative trial the QuantiTect Multiplex PCR NoROX Kit (Qiagen GmbH, Hilden/Germany) was used. This information is given for convenience of users of this document and does not constitute an endorsement by ISO of the product named. Equivalent products from other manufacturers may be used if they yield similar or better results. If necessary, adapt the amounts of the reagents and the temperature-time programme.	

Mix the PCR reagent mixture, centrifuge briefly and pipette 20 µl into each reaction vial. For the amplification reagent control, add 5 µl of water into the respective reaction setup. Pipette either 5 µl of sample DNA or 5 µl of the respective control solution (extraction blank control, positive DNA target control). If necessary, prepare a PCR inhibition control as described in ISO 24276.

Transfer the reaction setups into the thermal cycler and start the temperature-time programme.

7.4 Temperature-time programme

The temperature-time programme as outlined in [Table 3](#) has been used in the validation study. The use of different reaction conditions and real-time PCR cyclers may require specific optimization. The time for initial denaturation depends on the master mix used.

Table 3 — Temperature-time programme

Step	Parameter	Temperature	Time	Fluorescence measurement	Cycles
1	UNG activation (optional)	50 °C	2 min	no	1
2	Initial denaturation	95 °C	15 min	no	1
3	Amplification	Denaturation	95 °C	no	45
		Annealing and elongation	60 °C	yes	

8 Accept/reject criteria

8.1 General

A corresponding real-time PCR device-specific data analysis programme is used for the identification of PCR products. The amplification results may be expressed in a different manner, depending on the device used. In the absence of detectable PCR products (e.g. negative controls) the result can be expressed as “undetermined”, “no amp”, or the maximum number of reaction cycles performed. If the amplification of the DNA target sequence in a sample (e.g. positive controls) occurred, a sigmoid shaped amplification curve should be observed. The cycle number at the crossing point of the amplification curve and the fluorescence threshold is calculated (C_t value or C_p value).

If, due to atypical fluorescence measurement data, the automatic interpretation does not provide a meaningful result, it may be required to set the baseline and the threshold manually prior to interpreting the data. In this case, the device-specific instructions given in the manual regarding the use of the interpretation software should be applied.

A positive result can be also obtained if the sample contains DNA derived from the *Figwort mosaic virus*, which can naturally infect plants. To proof the presence of a GM plant derived product additional tests using construct-specific or event-specific methods are required for confirmation.

8.2 Identification

The target sequence is considered as detected, if

- by using the P-FMV specific primers pFMV-F and pFMV-R and the probe pFMV, a sigmoid shaped amplification curve is observed and a C_t value or C_p value is calculated,
- in the PCR control reactions with no added DNA (PCR reagent control, extraction blank control), no amplification has occurred, and
- in the reactions for the amplification controls (positive DNA target control, PCR inhibition control) the expected C_t values (or C_p values) are achieved.

9 Validation status and performance criteria

9.1 General

Validation followed a two part process:

- a) in-house validation followed by a pilot interlaboratory study;
- b) a collaborative trial validation.

9.2 Robustness

The robustness of the method was tested with respect to small changes in the conditions of the following factors: real-time PCR-instruments²⁾ (ABI7500, Rotor-Gene Q, Stratagene MX 3005P); PCR-master mix kits (QuantiTect Multiplex PCR NoROX Kit, SureMaster Probe Kit); volumes, 19 µl or 21 µl master mix plus 5 µl sample DNA); annealing temperature (59 °C and 61 °C); and primer or probe concentration (both lowered by 30 % respectively). For each tested factor three PCR replicates were analysed, each with 20 copies of the target sequence. In these tests the method showed positive results under the changed conditions in all PCR reactions as expected and thus the method can be considered robust.

9.3 Collaborative trial

The reliability of the method was verified in a collaborative trial with 16 participants, organized by the CVUA Freiburg in accordance with the IUPAC protocol.^[4] The participants received 18 DNA samples for the analysis. The samples contained the P-FMV target sequence at various concentrations, or did not contain the target sequence. All samples were labelled with random coding numbers. Each participant received 3 vials (triplicate-blind) containing the following DNA solutions:

- non GM rapeseed;
- non GM soy;
- GT73 rapeseed, adjusted to a calculated concentration of 4 copies/µl;
- GT73 rapeseed, adjusted to a calculated concentration of 10 copies/µl;
- MON89788 soy, adjusted to a calculated concentration of 4 copies/µl;
- MON89788 soy, adjusted to a calculated concentration of 10 copies/µl.

For conducting the PCR experiments participants received the PCR master mix and the oligonucleotides (primers and probes).

To prepare the samples, genomic DNA was extracted from ground seeds of GT73 canola (AOCS 0304-B from AOAC Boulder/USA) and of MON89788 soy (AOCS 0906-B).³⁾ The DNA was adjusted with 0,2x TE buffer to a nominal concentration of 10 copies/µl and 4 copies/µl of the target sequence in a background of 20 ng/µl of rapeseed or soy DNA.

The DNA concentrations were determined by fluorescence detection using the PicoGreen assay.^[5] The copy numbers were established on the basis of the genome equivalents. For soy a haploid genome mass of 1,15 pg and for rapeseed of 1,23 pg was taken.^[6] The integration of one copy of the P-FMV target sequence in rapeseed GT73 and soy MON89788 genome was assumed,^{[7],[8]} and the GM material was considered homozygous.

To prepare samples not containing the target sequence, DNA was extracted from non GM rapeseed or soy (market samples) and adjusted to a concentration of approximately 20 ng/µl.

The DNA samples and a plasmid DNA standard, were tested for the presence of FMV DNA using the method described in [Annex A](#).^[9] Negative results were obtained in each case.

The collaborative trial was designed for calculating the false positive and false negative rates. Each DNA sample was tested by the participants in a single PCR test with 5 µl of the respective DNA solution, using the procedure and the conditions given in [Tables 2](#) and [3](#). The results of the collaborative trial are listed in [Table 4](#), more details on precision are provided in [Table 5](#).

2) This information is given for the convenience of users of this document and does not constitute an endorsement by ISO of the products named. Equivalent products may be used if they can be shown to lead to the same results.

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