

Designation: E 478 – 89a (Reapproved 1996)

Standard Test Methods for Chemical Analysis of Copper Alloys¹

This standard is issued under the fixed designation E 478; the number immediately following the designation indicates the year of original adoption or, in the case of revision, the year of last revision. A number in parentheses indicates the year of last reapproval. A superscript epsilon (ϵ) indicates an editorial change since the last revision or reapproval.

1. Scope

1.1 These test methods cover the chemical analysis of copper alloys having chemical compositions within the follow-ing limits:²

Element	Concentration, %
Aluminum	12.0 max
Antimony	1.0 max
Arsenic	1.0 max
Cadmium	1.5 max
Cobalt	1.0 max
Copper	40.0 min
Iron	6.0 max
Lead	27.0 max
Manganese	6.0 max
Nickel	50.0 max
Phosphorus	1.0 max
Silicon	5.0 max
Sulfur	0.1 max
Tin	20.0 max
Zinc	50.0 max

1.2 The test methods appear in the following order:

	Sections
Aluminum by the Carbamate Extraction-(Ethyl-	
enedinitrilo) Tetraacetate Titrimetric Test	
Method	70-77
Copper by the Combined Electrodeposition	
Gravimetric and Oxalyldihydrazide Photomet-	
ric Test Method [50 %, minimum] Catalog/Stan	dards/9-17/1154
Iron by the 1,10-Phenanthroline Photometric	
Test Method [0.003 to 1.25 %]	18-27
Lead by the Atomic Absorption Test Method	89-99
Lead by the (Ethylenedinitrilo)tetraacetic Acid	
(EDTA) Titrimetric Test Method [2.0 to	
30.0 %]	28-35
Nickel by the Dimethylglyoxime Extraction Pho-	
tometric Test Method [0.03 to 5.0 %]	36-45
Nickel by the Dimethylglyoxime Gravimetric	
Test Method [4 to 50 %]	54-61
Silver in Silver-Bearing Copper by the Atomic	
Absorption Test Method	100-111
Tin by the lodatimetric Titration Test Method	
[0.5 to 20 %]	62-69
Tin by the Phenylfluorone Photometric Test	
Method %]	112-122
Zinc by Atomic Spectrometry	78-88

¹ These test methods are under the jurisdiction of ASTM Committee E-1 on Analytical Chemistry for Metals, Ores, and Related Materials and are the direct responsibility of Subcommittee E01.05 on Cu, Pb, Zn, Cd, Sn, Be, their Alloys and Related Metals.

Current edition approved Oct. 27, 1989. Published January 1990. Originally published as E 478 - 73. Last previous edition E 478 - 89.

² The actual limits of application of each test method are presented in 1.2.

Zinc by the (Ethylenedinitrilo)tetraacetic Acid (EDTA) Titrimetric Test Method [2 to 40 %]

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1.3 This standard does not purport to address all of the safety concerns, if any, associated with its use. It is the responsibility of the user of this standard to establish appropriate safety and health practices and determine the applicability of regulatory limitations prior to use. Specific hazard statements are given in Section 5, Note 4, and Section 106.

2. Referenced Documents

2.1 ASTM Standards:

E 29 Practice for Using Significant Digits in Test Data to Determine Conformance with Specifications³

E 50 Practices for Apparatus, Reagents, and Safety Precautions for Chemical Analysis of Metals⁴

E 60 Practice for Photometric and Spectrophotometric Methods for Chemical Analysis of Metals⁴

E 173 Practice for Conducting Interlaboratory Studies of Methods for Chemical Analysis of Metals⁴

E 255 Practice for Sampling Copper and Copper Alloys for Determination of Chemical Composition⁴

E 1024 Guide for Chemical Analysis of Metals and Metal Bearing Ores by Flame Atomic Absorption Spectrophotometry⁵

3. Significance and Use

3.1 These test methods for the chemical analysis of metals and alloys are primarily intended as referee methods to test such materials for compliance with compositional specifications. It is assumed that all who use these methods will be trained analysts capable of performing common laboratory procedures skillfully and safely. It is expected that work will be performed in a properly equipped laboratory.

4. Apparatus, Reagents, and Photometric Practice

4.1 Apparatus and reagents required for each determination are listed in separate sections preceding the procedure. The apparatus, standard solutions, and certain other reagents used in more than one procedure are referred to by number and shall conform to the requirements prescribed in Practices E 50,

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³ Annual Book of ASTM Standards, Vol 14.02.

⁴ Annual Book of ASTM Standards, Vol 03.05.

⁵ Annual Book of ASTM Standards, Vol 03.06.

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except that photometers shall conform to the requirements prescribed in Practice E 60.

4.2 Photometric practice prescribed in these test methods shall conform to Practice E 60.

5. Safety Precautions

5.1 For precautions to be observed in the use of certain reagents in these test methods, refer to Practices E 50.

6. Sampling

6.1 For procedures for sampling the material, refer to Practice E 255. However, this method does not supersede any sampling requirements specified in a specific ASTM material specification.

7. Rounding Calculated Values

7.1 Calculated values shall be rounded to the desired number of places as directed in 3.4 to 3.6 of Practice E 29.

8. Interlaboratory Studies

8.1 These test methods have been evaluated in accordance with Practice E 173 unless otherwise noted in the precision section.

COPPER BY THE COMBINED ELECTRODEPOSITION GRAVIMETRIC AND OXALYLDIHYDRAZIDE PHOTOMETRIC TEST METHOD

9. Scope

9.1 This test method covers the determination of copper in Concentrations greater than 50 %.

10. Summary of Test Method

10.1 After dissolution of the sample in nitric and hydrofluoric acids, the oxides of nitrogen are reduced with hydrogen peroxide, and the copper deposited electrolytically. Loss of platinum from the anode is minimized by the addition of lead. The copper oxalyldihydrazide complex is formed with the copper remaining in the electrolyte. Photometric measurement is made at approximately 540 nm.

11. Interferences

11.1 The elements ordinarily present do not interfere if their concentrations are under the maximum limits shown in 1.1.

12. Apparatus

12.1 Apparatus No. 9—Gauze cathodes are recommended where rapid electrolysis is used.

12.2 Polytetrafluoroethylene or Polypropylene Beakers, 250-mL capacity.

12.3 Polytetrafluoroethylene or Polypropylene Split Covers.

13. Reagents

13.1 Ammonium Chloride Solution (0.02 g/L)—Dissolve 0.02 g of ammonium chloride (NH₄Cl) in water and dilute to 1 L.

13.2 *Hydrogen Peroxide* (3 %)—Dilute 100 mL of 30 % hydrogen peroxide to 1 L.

13.3 *Lead Nitrate Solution* (10 g/L)—Dissolve 10.0 g of lead nitrate (Pb(NO₃)₂) in water and dilute to 1 L.

14. Procedure

14.1 Transfer a 2.000-g sample, weighed to the nearest 0.1 mg, to a 250-mL poly(tetrafluoroethylene) or polypropylene beaker, add 2 mL of HF, and 30 mL of HNO₃ (1 + 1). Cover with a cover glass and allow to stand for a few minutes until the reaction has nearly ceased. Warm but do not heat over 80°C. When dissolution is complete, add 25 mL of 3 % H_2O_2 and 3 mL of Pb(NO₃)₂ solution. Rinse the cover glass and dilute to approximately 150 mL with NH₄Cl solution.

14.2 With the electrolyzing current off, position the anode and the accurately weighed cathode in the solution so that the gauze is completely immersed. Cover the beaker with a split plastic cover.

14.3 Start the electrolysis and increase the voltage until the ammeter indicates a current which is equivalent to about 1.0 A/dm^2 and electrolyze overnight. Alternatively electrolyze at a current density of $4 A/dm^2$ for 1.5 h. (The more rapid procedure requires the use of gauze electrodes).

14.4 Slowly withdraw the electrodes (or lower the beaker) with the current still flowing, and rinse with a stream of water from a wash bottle. Quickly remove the cathode, rinse it in water, and then dip into two successive baths of ethanol or methanol. Dry in an oven at 110°C for 3 to 5 min.

14.5 Return the voltage to zero, and turn off the switch. Reserve the electrolyte.

14.6 Allow the electrode to cool to room temperature, and weigh.

15. Calculation

STM E47 15.1) Calculate the percentage of copper as follows:

fcd7-27d3-4e58-| Copper, % = $[(A + B/C] \times 100 \text{ m} - e478 - 89a (1))6$

where:

A = deposited copper, g,

- B = copper in the electrolyte as calculated in 16.10, g, and
- C = sample used, g.

16. Photometric Determination of the Residual Copper in the Electrolyte

16.1 *Interferences*—The elements ordinarily present do not interfere if their concentrations are under the maximum limits shown in 1.1.

16.2 *Concentration Range*—The recommended concentration range is from 0.0025 to 0.07 mg of copper per 50 mL of solution, using a 2-cm cell.

NOTE 1—This procedure has been written for cells having a 2-cm light path. Cells having other dimensions may be used, provided suitable adjustments can be made in the amounts of sample and reagents used.

16.3 *Stability of Color*—The color fully develops in 20 min, and is stable for 1 h.

16.4 Reagents:

16.4.1 Acetaldehyde Solution (40%)—Dilute 400 mL of acetaldehyde to 1 L with water.

16.4.2 Boric Acid Solution-Reagent No. 136.

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16.4.3 *Citric Acid Solution* (200 g/L)—Dissolve 200 g of citric acid in water and dilute to 1 L.

16.4.4 *Copper, Standard Solution A* (1 mL = 1.0 mg Cu)— Transfer a 1.000-g sample of electrolytic copper (purity: 99.9 % minimum) to a 250-mL beaker and add 10 mL of HNO_3 (1 + 1). Evaporate till nearly to dryness. Add 5 mL of water to dissolve the residue. Transfer to a 1-L volumetric flask, dilute to volume, and mix.

16.4.5 *Copper, Standard Solution B* (1 mL = 0.010 mg Cu)—Using a pipet, transfer 10 mL of copper solution A (1 mL = 1.0 mg Cu) to a 1-L volumetric flask, dilute to volume and mix.

16.4.6 *Oxalyldihydrazide Solution* (2.5 g/L)—Dissolve 2.5 g of oxalyldihydrazide in warm water and dilute to 1 L.

16.5 Preparation of Calibration Curve:

16.5.1 Calibration Solutions:

16.5.1.1 Transfer 25 mL of boric acid solution to a 250-mL volumetric flask and then add a solution containing 150 mL of water, 2 mL of HF, and 30 mL of HNO_3 (1 + 1). Dilute to volume, and mix.

16.5.1.2 Transfer 10 mL of this solution to each of four 50-mL volumetric flasks. Using pipets, transfer 1, 3, 5, and 7 mL of copper solution B (1 mL = 0.010 mg Cu) to the flasks. Proceed as directed in 16.5.3.

16.5.2 *Reference Solution*—Add 10 mL of boric acid solution prepared as directed in 16.5.1.1 to a 50-mL volumetric flask and proceed as directed in 16.5.3.

16.5.3 *Color Development*—Add in order, and with mixing after each addition, 5 mL of citric acid solution, 6 mL of NH₄OH, 10 mL of acetaldehyde solution, and 10 mL of oxalyldihydrazide solution. Cool, dilute to volume, and mix. Allow to stand for 30 min and proceed as directed in 16.5.4.

16.5.4 Photometry:

16.5.4.1 *Multiple-Cell Photometer*—Measure the cell correction using absorption cells with a 2-cm light path and a light band centered at approximately 540 nm. Using the test cell, take the photometric readings of the calibration solutions.

16.5.4.2 *Single-Cell Photometer*—Transfer a suitable portion of the reference solution to an absorption cell with a 2-cm light path and adjust the photometer to the initial setting using a light band centered at approximately 540 nm. While maintaining this adjustment, take the photometric readings of the calibration solutions.

16.5.5 *Calibration Curve*—Plot the net photometric-readings of the calibration solutions against milligrams of copper per 50 mL of solution.

16.6 *Test Solution*—Transfer the reserved electrolyte to a 250-mL volumetric flask containing 25 mL of boric acid solution, dilute to volume, and mix. Using a pipet, transfer 10 mL to a 50-mL volumetric flask (Note 2). Proceed as directed in 16.8.

NOTE 2—If the solution shows a permanganate color, add sodium nitrite solution (20 g/L) dropwise until the color is discharged, and then proceed as directed in 16.8.

16.7 Reference Solution—Proceed as directed in 16.5.2.

16.8 Color Development—Proceed as directed in 16.5.3.

16.9 *Photometry*—Take the photometric reading of the test solution as directed in 16.5.4.

16.10 *Calculation*—Convert the net photometric reading of the test solution to milligrams of copper by means of the calibration curve. Calculate the grams of copper in the total electrolyte as follows:

Copper,
$$g = (A \times 25)/1000$$
 (2)

where:

A = copper found in 50 mL of the final test solution, mg.

17. Precision

17.1 Eight laboratories cooperated in testing this test method and obtained the data summarized in Table 1.

TABLE 1 Statistical Information

Test Specimen	Copper Found, %	Repeatability (R ₁ , E 173)	Reproducibility (R ₂ , E 173)
1. Bronze ounce metal (NBS	83.56	0.09	0.13
124d, 83.60 Cu) 2. AAB 521	91.98	0.03	0.08
3. AAB 655	95.38	0.09	0.14
4. AAB 681	57.60	0.10	0.09
5. AAB 715	68.95	0.08	0.21

IRON BY THE 1,10-PHENANTHROLINE PHOTOMETRIC TEST METHOD

18. Scope

18.1 This test method covers the determination of iron in concentrations from 0.003 to 1.25 %.

19. Summary of Test Method

19.1 The sample is dissolved in hydrochloric acid and hydrogen peroxide, and the excess oxidant removed by evaporation. The iron is extracted with methyl isobutyl ketonebenzene mixture. The iron is extracted from the organic phase into a hydroxylamine hydrochloride solution and the redcolored 1,10-phenanthroline complex is formed. Photometric measurement is made at approximately 510 nm.

20. Concentration Range

20.1 The recommended concentration range is from 0.005 to 0.125 mg of iron per 50 mL of solution, using a 2-cm cell.

NOTE 3—This test method has been written for cells having a 2-cm light path. Cells having other dimensions may be used, provided suitable adjustments can be made in the amounts of sample and reagents used.

21. Stability of Color

21.1 The color develops within 5 min and is stable for at least 4 h.

22. Interferences

22.1 Elements ordinarily present do not interfere if their concentrations are under the maximum limits shown in 1.1.

23. Reagents

23.1 *Hydroxylamine Hydrochloride Solution* (10 g/L)— Prepare a solution as directed for Reagent No. 131, but dilute to 500 mL.

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23.2 Iron, Standard Solution A (1 mL = 0.125 mg Fe)— Prepare a solution as directed for Reagent No. 4, but use 0.1250 g instead of the specified weight.

23.3 *Iron, Standard Solution B* (1 mL = 0.00625 mg Fe)—Using a pipet, transfer 50 mL of iron solution A to a 1-L volumetric flask, dilute to volume with HCl (1 + 49), and mix.

23.4 *Methyl Isobutyl Ketone-Benzene Mixture*—Mix 200 mL of methyl isobutyl ketone (MIBK) and 100 mL of benzene.

23.5 1,10-Phenanthroline-Ammonium Acetate Buffer Solution—Dissolve 1.0 g of 1,10-phenanthroline monohydrate in 5 mL of HCl in a 600-mL beaker. Add 215 mL of acetic acid, and, while cooling, carefully add 265 mL of NH₄OH. Cool to room temperature. Using a pH meter, check the pH; if it is not between 6.0 and 6.5, adjust it to that range by adding acetic acid or NH₄OH as required. Dilute to 500 mL.

24. Preparation of Calibration Curve

24.1 Calibration Solutions:

24.1.1 Using pipets, transfer 1, 2, 5, 10, 15, and 20 mL of iron solution B (1 mL = 0.00625 mg Fe) to 50-mL volumetric flasks. Dilute to 20 mL.

24.1.2 Add 20 mL of $NH_2OH \cdot HCl$ solution, mix, and allow to stand 1 min. Proceed as directed in 24.3.

24.2 *Reference Solution*—Transfer 20 mL of water to a 50-mL volumetric flask and proceed as directed in 24.1.2.

24.3 *Color Development*—Add 5 mL of 1,10phenanthroline-ammonium acetate buffer solution, dilute to volume, and mix. Allow to stand at least 5 min but not more than 4 h.

24.4 *Photometry*:

24.4.1 *Multiple-Cell Photometer*—Measure the cell correction using absorption cells with a 2-cm light path and a light band centered at approximately 510 nm. Using the test cell, take the photometric readings of the calibration solutions.

24.4.2 *Single-Cell Photometer*—Transfer a suitable portion of the reference solution to an absorption cell with a 2-cm light path and adjust the photometer to the initial setting, using a light band centered at approximately 510 nm. While maintaining this adjustment, take the photometric readings of the calibration solutions.

24.5 *Calibration Curve*—Plot the net photometric readings of the calibration solutions against milligrams of iron per 50 mL of solution.

25. Procedure

25.1 Test Solution:

25.1.1 Select and weigh a sample in accordance with the following:

Iron, %	Sample Weight, g	Tolerance in Sample Weight, mg	Aliquot Volume, mL
0.003 to 0.02	2.0	2.0	25
0.02 to 0.10	1.0	1.0	10
0.05 to 0.20	0.5	0.5	10
0.10 to 0.40	0.5	0.5	5
0.25 to 1.25	0.2	0.5	5

Transfer it to a 400-mL beaker, or to a poly(tetrafluoroethylene) beaker if HF is to be used. 25.1.2 Carry a reagent blank through the entire procedure, using the same amounts of all reagents but with the sample omitted.

25.1.3 Add 12 mL of HCl (7 + 3) per gram of sample, and then H₂O₂ as needed to completely dissolve the alloy. Add HF as needed to decompose high-silicon alloys. When dissolution is complete, add 10 mL of concentrated HCl per gram of sample and heat carefully to decompose excess peroxide. Cool to room temperature, transfer to a 100-mL volumetric flask, dilute to volume with HCl (1 + 1), and mix.

25.1.4 Using a pipet, transfer an aliquot in accordance with 25.1.1 to a 125-mL conical separatory funnel. Add HCl (1 + 1), as required, to adjust the volume to 25 mL.

25.1.5 Add 20 mL of MIBK-benzene mixture to the separatory funnel and shake 1 min. Allow the phases to separate, discard the aqueous phase, wash the organic phase 3 times with 3 to 5-mL portions of HCl (1 + 1) to remove copper, and discard the washings. Extract the iron from the organic phase by shaking vigorously 30 s with 10 mL of NH₂OH·HCl solution. Transfer the aqueous phase to a 50-mL volumetric flask. Repeat the extraction with a second 10-mL portion of NH₂OH·HCl solution, and transfer the extract to the 50-mL flask.

25.2 *Reference Solution*—Use the reagent blank solution prepared as directed in 25.1.2.

25.3 Color Development—Proceed as directed in 24.3.

25.4 Photometry-Proceed as directed in 24.4.

26. Calculation

26.1 Convert the net photometric reading of the test solution to milligrams of iron by means of the calibration curve. Calculate the percentage of iron as follows:

$$Iron, \% = A/(B \times 10)$$

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(3)

where:

A = iron found in 50 mL of the final test solution, mg, and B = sample represented in 50 mL of the final test solution,

27. Precision

g.

27.1 Seven laboratories cooperated in testing this method, submitting nine pairs of values, and obtained the data summarized in Table 2.

TABLE 2 Statistical Information

Test Specimen	Iron Found, %	Repeatability (R ₁ , E 173)	Reproducibility (<i>R</i> ₂ , E 173)
1. Cast bronze (NBS 52c, 0.004 Fe)	0.0034	0.0005	0.0010
2. Ounce metal (NBS 124d	, 0.187	0.012	0.017
0.18 Fe)			
3. Cupro Nickel, 30 Ni	0.60	0.015	0.044
4. Silicon bronze (NBS 158a, 1.23 Fe)	1.24	0.019	0.037

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LEAD BY THE (ETHYLENEDINITRILO)TETRAACETIC ACID (EDTA) TITRIMETRIC TEST METHOD

28. Scope

28.1 This test method covers the determination of lead in concentrations from 2.0 to 30.0 %.

29. Summary of Test Method

29.1 Lead diethyldithiocarbamate is extracted with chloroform from an alkaline tartrate-cyanide solution. After the removal of organic material, lead is titrated with disodium (ethylenedinitrilo) tetraacetic acid (EDTA) solution.

30. Interferences

30.1 Elements ordinarily present do not interfere if their concentrations are under the maximum limits shown in 1.1.

31. Apparatus

31.1 Separatory Funnels, 250-mL capacity.

31.2 Magnetic Stirrer and Poly(tetrafluoroethylene)-Covered Magnetic Stirring Bar.

32. Reagents

32.1 Ascorbic Acid.

32.2 Chloroform (CHCl₃).

32.3 Disodium (Ethylenedinitrilo) tetraacetic Acid (EDTA), Standard Solution (0.025 M)—Prepare a solution as directed for Reagent No. 22, using 9.3 g instead of the specified weight. Standardize as follows: Using a pipet, transfer 25 mL of lead solution (1 mL = 6.0 mg Pb) to a 250-mL beaker and dilute to 100 mL. Proceed as directed in 33.7. Calculate the lead equivalent of the solution as follows:

Lead equivalent,
$$g/mL = A/B$$

where:

A = weight of lead, g, and

B = EDTA solution required for titration of the lead solution, mL.

32.4 *Fluoboric Acid* (37 to 40%).

32.5 Hexamethylenetetramine.

32.6 *Lead, Standard Solution* (1 mL = 6.0 mg Pb)— Transfer 1.500 g of lead (purity 99.9 % minimum) to a 150-mL beaker. Add 10 mL of HNO_3 (1 + 1) and heat until dissolution is complete. Boil to remove oxides of nitrogen, cool, transfer to a 250-mL volumetric flask, dilute to volume, and mix.

32.7 *Sodium Cyanide Solution* (200 g/L)—Dissolve 200 g of sodium cyanide (NaCN) in water and dilute to 1 L. Store in a plastic bottle.

NOTE 4—**Caution:** The preparation, storage, and use of NaCN solutions require care and attention. Avoid inhalation of fumes and exposure of skin to the chemical and its solutions. Work in a well-ventilated hood. Refer to Section 6 of Practices E 50.

32.8 Sodium Diethyldithiocarbamate Solution (100 g/L)— Dissolve 10 g of sodium diethyldithiocarbamate in water and dilute to 100 mL. Do not use a solution that has stood more than 24 h. 32.9 *Sodium Hydroxide Solution* (250 g/L)—Dissolve 250 g of sodium hydroxide (NaOH) in water and dilute to 1 L. Store in a plastic bottle.

32.10 *Sodium Tartrate Solution* (250 g/L)—Dissolve 250 g of sodium tartrate dihydrate in water and dilute to 1 L.

32.11 *Xylenol Orange Indicator Solution* (1 g/L)—Dissolve 0.050 g of xylenol orange powder in a mixture of 25 mL of water and 25 mL of ethanol.

33. Procedure

33.1 Select a sample in accordance with the following:

Lead, %	Sample Weight, g	
2.0 to 20.0	1.00	
20.0 to 30.0	0.60	

Weigh the sample to the nearest 0.5 mg, and transfer it to a 250-mL beaker.

33.2 Add 5 mL of HBF_4 and then 10 mL of HNO_3 (1 + 1). Cover the beaker and heat until dissolution is complete. Boil until oxides of nitrogen have been expelled, and cool.

33.3 Wash the cover and walls of the beaker. Add 25 mL of sodium tartrate solution, 25 mL of NaOH solution, and 25 mL of NaCN solution (**Caution**, Note 4), mixing after each addition. Cool to room temperature.

33.4 Transfer to a 250-mL separatory funnel. Add 15 mL of sodium diethyldithiocarbamate solution and 15 mL of $CHCl_3$, and shake for 30 s. Allow the layers to separate; draw off the lower organic layer into a 250-mL beaker, retaining the aqueous layer. Add 5 mL more of diethyldithiocarbamate solution to the separatory funnel and mix. If no precipitate forms, proceed as directed in 33.5. If a precipitate does form, add 5 mL of diethyldithiocarbamate solution and 10 mL of $CHCl_3$, shake for 30 s, and draw off the organic layer into the 250-mL beaker containing the extract.

-33.5 Extract twice with additional 10-mL portions of CHCl₃, adding the extracts to the extracts in the 250-mL beaker.

33.6 Add 10 mL of HCl (1 + 1) to the combined extracts, and place on a hot plate. Cover the beaker with a raised cover glass, and evaporate the solution to a volume of 2 to 3 mL. Wash the cover and walls of the beaker, dilute to 100 mL, and heat to dissolve salts.

33.7 Place the beaker on a magnetic stirrer and stir (Note 5). Add 10 to 20 mg of ascorbic acid and 3 or 4 drops of xylenol orange solution. Add enough hexamethylenetetramine to color the solution purple. Add 4 or 5 drops of NaCN solution (**Caution**, Note 4) and titrate with the EDTA solution. When a yellow color begins to appear, stop the titration and add 2 to 3 g of hexamethylenetetramine and a drop of xylenol orange solution. Titrate dropwise until the color changes from purplish-red to yellow.

Note 5—The titration may be performed in either a hot or cold solution.

34. Calculation

34.1 Calculate the percentage of lead as follows:

Lead, % =
$$[(C \times D)/E] \times 100$$
 (5)

(4)

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where:

C = standard EDTA solution used, mL,

D = equivalent of EDTA solution, g/mL, and

E = sample used, g.

35. Precision

35.1 Due to limited data, a precision statement conforming to the requirements of Practices E 173 cannot be furnished. However, in a cooperative program conducted by six laboratories, the between-laboratory range was 3.13 to 3.20 % lead on a sample averaging 3.16 %, and 14.05 to 14.23 % on a sample averaging 14.15 %.

NICKEL BY THE DIMETHYLGLYOXIME-EXTRACTION PHOTOMETRIC TEST METHOD

36. Scope

36.1 This test method covers the determination of nickel in concentrations from 0.03 to 5.0 %.

37. Summary of Test Method

37.1 A dimethylglyoxime complex of nickel is formed in the presence of copper, and extracted with chloroform. Photometric measurement is made at approximately 405 nm.

38. Concentration Range

38.1 The recommended concentration range is 0.015 to 0.3 mg of nickel per 20 mL of solution, using a 2-cm cell.

NOTE 6—This procedure has been written for a cell having a 2-cm light path. Cells having other dimensions may be used, provided suitable adjustments can be made in the amounts of sample and reagents used.

39. Stability of Color

39.1 The color is stable for at least 2 h.

40. Interferences

40.1 The elements ordinarily present do not interfere if their concentrations are under the maximum limits shown in 1.1.

41. Reagents

41.1 *Chloroform* (CHCl₃).

41.2 Complexing Solution—Mix 240 mL of sodium tartrate solution, 90 mL of NaOH solution, 480 mL of sodium acetate solution, and 200 mL of $Na_2S_2O_3$ solution.

41.3 Dimethylglyoxime Solution (10 g/L in alcohol)— Reagent No. 104.

41.4 Hydroxylamine Hydrochloride Solution (10 g/L)— Dissolve 10 g of hydroxylamine hydrochloride (NH₂OH·HCl) in water, and dilute to 1 L. Adjust the pH to 7.0 with NH₄OH.

41.5 *Nickel, Standard Solution A* (1 mL = 1.0 mg Ni)— Dissolve 1.000 g of nickel metal (purity, 99.8 % min) in 10 mL of HNO₃. When dissolution is complete, boil gently to expel oxides of nitrogen, cool, transfer to a 1-L volumetric flask, dilute to volume, and mix.

41.6 Nickel, Standard Solution B (1 mL = 0.2 mg Ni)— Using a pipet, transfer 100 mL of nickel solution A (1 mL = 1.0 mg Ni) to a 500-mL volumetric flask, dilute to volume, and mix. 41.7 *Sodium Acetate Solution* (200 g/L)—Reagent No. 127, but use 200 g instead of the specified weight.

41.8 Sodium Hydroxide Solution (1 N)—Dissolve 40 g of sodium hydroxide (NaOH) in water, cool, transfer to a 1-L volumetric flask, dilute to volume, and mix. Store in a plastic bottle.

41.9 Sodium Sulfate, anhydrous (Na₂SO₄).

41.10 *Sodium Tartrate Solution* (100 g/L)—Dissolve 100 g of sodium tartrate dihydrate in water, and dilute to 1 L.

41.11 Sodium Thiosulfate Solution (200 g/L)—Dissolve 200 g of sodium thiosulfate pentahydrate ($Na_2S_2O_3$ · 5H₂O) in water, and dilute to 1 L.

42. Preparation of Calibration Curve

42.1 Calibration Solutions:

42.1.1 Transfer 1.000 g of copper (purity, 99.99 % min) to each of five 250-mL beakers, add 20 mL of HCl (1 + 1), and add 10 mL of H₂O₂ solution in small portions. When dissolution is complete, boil for 1 min to destroy excess peroxide, and cool.

42.1.2 Using pipets, transfer 2, 5, 10, 20, and 30 mL of nickel solution B (1 mL = 0.2 mg Ni) to the beakers. Transfer the solutions to 500-mL volumetric flasks, dilute to volume, and mix.

42.1.3 Using a pipet, transfer 25 mL to a 250-mL conical separatory funnel. Add 5 mL of NH_2OH ·HCl solution and 50 mL of complexing solution, shaking after each addition. Using indicator paper, check the pH, which should be between 6.5 and 7.2. If necessary, adjust the pH with HCl (1 + 1) or dilute NaOH solution.

42.2 *Reference Solution*—Transfer 1.000 g of copper (purity, 99.99 % min) to a 250-mL beaker and proceed as directed in 41.1, omitting the addition of nickel solution.

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42.3.1 Add 3 mL of dimethylglyoxime solution, and shake for 1 min. Using a pipet, transfer 20 mL of $CHCl_3$ to the solution, and shake again for 40 s. Allow the phases to separate.

42.3.2 Transfer the yellow-colored chloroform phase to a 25-mL Erlenmeyer flask fitted with a ground-glass stopper and containing about 1 g of Na_2SO_4 . Shake to stir the Na_2SO_4 into the CHCl₃. Decant the clear CHCl₃ solution into an absorption cell, and cover immediately to prevent loss of solvent.

42.4 *Photometry*:

42.4.1 *Multiple-Cell Photometer*—Measure the cell correction using absorption cells with a 2-cm light path and a light band centered at approximately 405 nm. Using the test cell, take the photometric readings of the calibration solutions.

42.4.2 *Single-Cell Photometer*—Transfer a suitable portion of the reference solution to an absorption cell with a 2-cm light path and adjust the photometer to the initial setting, using a light band centered at approximately 405 nm. While maintaining this adjustment, take the photometric readings of the calibration solutions.

42.5 *Calibration Curve*—Plot the net photometric readings of the calibration solutions against milligrams of nickel per 20 mL of solution.