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Designation: D 6042 – 96

Standard Test Method for Determination of Phenolic Antioxidants and Erucamide Slip Additives in Polypropylene Homopolymer Formulations Using Liquid Chromatography (LC)¹

This standard is issued under the fixed designation D 6042; the number immediately following the designation indicates the year of original adoption or, in the case of revision, the year of last revision. A number in parentheses indicates the year of last reapproval. A superscript epsilon (ε) indicates an editorial change since the last revision or reapproval.

1. Scope

1.1 This test method covers a liquid-chromatographic procedure for the separation of some additives currently used in polypropylene. These additives are extracted with a cyclohexane:methylene chloride mixture using either reflux or ultrasonic bath prior to liquid-chromatographic separation. The ultraviolet absorbence (200 nm) of the compound(s) is measured, and quantitation is performed using the internal standard method.

1.2 This standard does not purport to address all of the safety concerns, if any, associated with its use. It is the responsibility of the user of this standard to establish appropriate safety and health practices and determine the applicability of regulatory limitations prior to use. Specific precautionary statements are given in Section 9.

Note 1-There is no similar or equivalent ISO standard.

2. Referenced Documents

2.1 ASTM Standards:

D 883 Terminology Relating to Plastics²

- D 1600 Terminology for Abbreviated Terms Relating to $Plastics^2$
- E 131 Terminology Relating to Molecular Spectroscopy³
- E 380 Practice for Use of the International System of Units (SI) (the Modernized Metric System)⁴
- E 691 Practice for Conducting an Interlaboratory Study to
- Determine the Precision of a Test Method⁴

3. Terminology

3.1 For definitions of plastic terms used in this test method, see Terminologies D 883 and D 1600.

3.2 For the units, symbols, and abbreviations used in this test method, refer to Terminology E 131 or Practice E 380.

3.3 Abbreviations: Abbreviations:

3.3.1 LC—liquid chromatography.

3.3.2 *PP*—polypropylene.

3.4 Trade Names:

3.5 *Vitamin E*— α -Tocopherol, or 3,4-dihydro-2,5,7,8-tetramethyl-2-(4,8,12-trimethyltridecyl)-2H-1-benzopyran-6-ol.

3.6 Irgafos 168-Tris(2,4 di-tert-butylphenyl) phosphite.

3.7 *Irganox 3114*—Tris(3,5-di-t-butyl-4-hydroxybenzyl) isocyanurate.

3.8 *Kemamide-E*—cis-13-docosenamide or erucamide.

3.9 *Irganox* 1010—tetrakis[methylene(3,5-di-t-butyl-4-hydroxy hydrocinnamate)]methane.

3.10 *Irganox 1076*—octadecyl-3,5-di-t-butyl-4-hydroxy hydrocinnamate.

3.11 *Tinuvin P*—2(2'-hydroxy-5'-methyl phenyl)benzotriazole.

4. Summary of Test Method

4.1 The PP sample is ground to a 20-mesh particle size (850 microns) and extracted by refluxing with a mixture of 75:25 methylene chloride:cyclohexane or placing in an ultrasonic bath with the same mixture.

4.2 The solvent extract is examined by liquid chromatography.

4.3 Additive concentrations are determined relative to an internal standard (contained in the solvent) using reverse-phase chromatography (C-18 column) with ultraviolet (UV) detection at 200 nm.

5. Significance and Use

5.1 Separation and identification of stabilizers used in the manufacture of polypropylene is necessary in order to correlate performance properties with polymer composition. This test method provides a means to determine erucamide slip, Vitamin E, Irgafos 168, Irganox 3114, Irganox 1010, and Irganox 1076 levels in polypropylene samples. This test method should be applicable for the determination of other antioxidants, such as Ultranox 626, Ethanox 330, Santanox R, and BHT, but the applicability of this test method has not been investigated for these antioxidants.

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¹ This test method is under the jurisdiction of ASTM Committee D-20 on Plastics and is the direct responsibility of Subcommittee D20.70 on Analytical Methods. Current edition approved Nov. 10, 1996. Published April 1997.

² Annual Book of ASTM Standards, Vol 08.01.

³ Annual Book of ASTM Standards, Vol 03.06.

⁴ Annual Book of ASTM Standards, Vol 14.02.

5.2 The additive-extraction procedure is made effective by the insolubility of the polymer sample in solvents generally used for liquid chromatographic analysis.

5.3 Under optimum conditions, the lowest level of detection for a phenolic antioxidant is approximately 2 ppm.

NOTE 2—Other methods that have been used successfully to remove additives from the plastics matrix include thin film, microwave, ultrasonic, and supercritical fluid extractions. Other methods have been used successfully to separate additives including SFC and capillary GC.

5.4 Irgafos 168 is a phosphite antioxidant. Phosphites are known to undergo both oxidation and hydrolysis reactions. Less Irgafos 168 may be seen in the polymer than originally added to the polymer due to oxidation during processing. The HPLC separation is capable of separating the phosphite, phosphate (oxidation product), and hydrolysis product. If standards are obtained, then the reaction products also can be quantified. No significant breakdown of the phosphite antioxidant has been seen due to either extraction technique or the separation presented in this standard.

6. Interferences

6.1 Any material eluting at or near the same retention time as the additive can cause erroneous results. A polymer-solventextract solution containing no internal standard should be examined to minimize the possibility of interferences.

6.2 A major source of interferences can be from solvent impurities. For this reason, the solvents should be examined prior to use by injecting a sample of solvent on the HPLC system and analyzing as in Section 10.

6.3 The oxidation product of Irgafos 168 can overlap with other additives with retention times of components that elute between Irganox 1010 and Irgafos 168. Standards should be run to ensure that components of interest do not coelute with the oxidation product of Irgafos 168.

7. Apparatus

7.1 *Liquid Chromatograph*, equipped with a variablewavelength ultraviolet detector, heated column, and gradientelution capabilities. The liquid chromatograph should be equipped with a means for a $10-\mu$ L-sample-solution injection, such as a sample loop.

7.2 *Chromatographic Column*, RP-18, 5-µm particle size, 15 cm by 4.6 mm.

NOTE 3—A Zorbax RX C-18 Column, MAC MOD Analytical, Inc., was used for this test method. The gradient described in 10.1 provides complete separation of antioxidants using this C-18 column.

7.3 *Computer System or Integrator*, coupled with the chromatograph is recommended to measure peak area.

7.4 *Wiley Mill*, equipped with a 20-mesh screen and watercooled jacket to prevent thermodegradation of antioxidants.

7.5 *Recorder*, mV scale dependent upon the output of the detector.

7.6 *Reflux Extraction Apparatus*, consisting of a condenser (24/40 ground-glass joint), a flat bottom 125 mL flask having a 24/40 ground-glass joint, and a hot plate with magnetic stirrer (see Fig. 1).

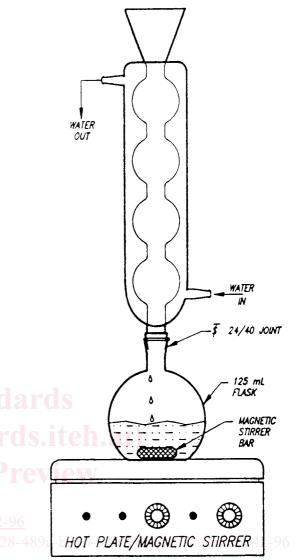


FIG. 1 Sample Extraction

7.7 *Ultrasonic Bath Apparatus*, consisting of the ultrasonic bath, a flat bottom 125-mL flask having a 24/40 ground-glass joint, and a 24/40 ground-glass stopper.

7.8 *Filter System* (Teflon)⁵ for nonaqueous solutions (pore size of 0.22 μ m) equipped with a glass 5-cm³ syringe.

7.9 Analytical Balance, capable of weighing to ± 0.0001 g.

8. Reagents and Materials

8.1 *Tinuvin-P*, 2(2' hydroxy-5'-methyl phenyl) benzotriaz-ole.

8.2 *Methylene Chloride*, HPLC grade, spectral quality or chromatography quality reagent.

8.3 *Cyclohexane*, HPLC grade, spectral quality or chromatography quality reagent.

8.4 75:25 Methylene Chloride:Cyclohexane T-P, mixture with 51.8 mg/L Tinuvin-P added as an internal standard.

8.5 *Water*, HPLC or UV quality reagent, degassed by sparging with high purity helium or by filtration under vacuum.

⁵ Registered trademark of DuPont.