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Standard Test Method for Evaluation of Surgical Hand Scrub Formulations¹

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1. Scope

1.1 This test method is designed to measure the reduction of microbial flora on the skin. It is intended for determining both immediate and persistent (continuing antimicrobial effect) microbial reductions, after single or repetitive treatments, or both. It may also be used to measure cumulative antimicrobial activity after repetitive treatments.

1.2 A knowledge of microbiological techniques is required for these procedures.

~~1.3 In this method, metric units are used for all applications, except for distance, in which case inches are used and metric units follow in parentheses.~~

~~1.4 Performance of this procedure requires the knowledge of regulations pertaining to the protection of human subjects.~~

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1.4 The values stated in SI units are to be regarded as standard. No other units of measurement are included in this standard.

1.4.1 In this test method, SI units are used for all applications, except for distance, in which case inches are used and SI units follow in parentheses.

1.5 *This standard does not purport to address all of the safety concerns, if any, associated with its use. It is the responsibility of the user of this standard to establish appropriate safety and health practices and determine the applicability of regulatory limitations prior to use.*

2. Referenced Documents

2.1 *ASTM Standards:*²

D1193 [Specification for Reagent Water](#)

E1054 ~~[Test Methods for Evaluation of Inactivators of Antimicrobial Agents](#)~~ [Test Methods for Evaluation of Inactivators of Antimicrobial Agents](#)

E2180 [Test Method for Determining the Activity of Incorporated Antimicrobial Agent\(s\) In Polymeric or Hydrophobic Materials](#)

2.2 *Other Documents:*

21 CFR Parts 50 and 56³

~~AATCC Test Method 147-1993 Antibacterial Assessment of Textile Materials: Parallel Streak Method~~ AATCC 147–2004 [Antibacterial Assessment of Textile Materials: Parallel Streak Method](#)⁴

JIS Z 2801 :2000, [Antimicrobial Products—Test for Antimicrobial Activity and Efficacy](#)⁵

¹ This test method is under the jurisdiction of ASTM Committee E35 on Pesticides and Alternative Control Agents and is the direct responsibility of Subcommittee E35.15 on Antimicrobial Agents.

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² 21 CFR Ch. 1, Parts 50 and 56.

³ For referenced ASTM standards, visit the ASTM website, www.astm.org, or contact ASTM Customer Service at service@astm.org. For *Annual Book of ASTM Standards* volume information, refer to the standard's Document Summary page on the ASTM website.

⁴ For referenced ASTM standards, visit the ASTM website, www.astm.org, or contact ASTM Customer Service at service@astm.org. For *Annual Book of ASTM Standards* volume information, refer to the standard's Document Summary page on the ASTM website.

⁵ Available from U.S. Government Printing Office, 732 N. Capitol St., Washington, DC 20401, U.S. Government Bookstore, <http://bookstore.gpo.gov/baskets/cfr-listing.jsp>.

⁶ United States Code of Federal Regulations.

⁷ Technical Manual of the American Association of Textile Chemists and Colorists (AATCC), 2009, Vol 82, P.O. Box 12215, Research Triangle Park, NC 27709, <http://www.aatcc.org>.

⁸ Technical Manual of the American Association of Textile Chemists and Colorists, P.O. Box 12215, Research Triangle Park, NC 27709.

⁹ Available from Japanese Industrial Standards Committee, Divisional Council on Consumer Life, Japanese Standards Association (JSA), 4-1-24 Akasaka Minato-Ku, Tokyo, 107-8440, Japan, <http://www.jisa.or.jp>.

Horowitz, W. (Ed.), 2000, Official Methods of Analysis of AOAC International 17th Ed., Ch 17, p. 4, Sec. 17.2.01 (m). Assoc. of Off. Anal. Chemist, Washington, D.C.

United States Pharmacopeia, 25, 2001, United States Pharmacopeial Convention, Inc., Rockville, MD. Chapter 61 “Microbial Limits Test”

USP 32 United States Pharmacopeia, Chapter 61 “Microbial Limits Test”, 2009⁶

3. Terminology

3.1 Definitions:

3.1.1 *active ingredient*—a substance added to a formulation specifically for the inhibition or inactivation of microorganisms.

3.1.2 *cleansing wash*—~~a non-antimicrobial wash intended to remove gross soil or residues from the hands of the subjects prior to collecting baseline samples.~~ —a non-antimicrobial wash intended to remove gross soil or residues from the hands.

3.1.3 *cleansing wash formulation*—a liquid castile soap or other liquid soap with neutral pH which does not contain an antimicrobial.

3.1.4 *cumulative effect*—a progressive decrease in the number of microorganisms recovered following repeated applications.

3.1.5 *internal reference formulation*—a formulation with demonstrated performance characteristics within the laboratory.

3.1.6 *neutralization*—a process that results in quenching or inactivation of the antimicrobial activity of a formulation. This may be achieved through dilution of the formulation or through the use of chemical agents called neutralizers.

3.1.7 *persistence*—prolonged or extended antimicrobial activity that prevents or inhibits the proliferation or survival of microorganisms after treatment.

3.1.8 *sampling fluid*—a buffered solution that aids in recovery of microorganisms from the skin and neutralization of the active ingredient in test and internal reference formulations.

3.1.9 *test formulation*—a formulation containing an active ingredient(s).

4. Summary of Test Method

4.1 This test method is conducted on ~~subjects~~individuals selected from a group of ~~volunteers~~subjects who have refrained from using any antimicrobials for at least ~~two weeks~~one week prior to initiation of the test. Subjects are selected from this group on the basis of high initial bacterial count, $\geq 1 \times 10^5$ CFU/per hand as determined by baseline measurements of the bacteria on their hands using the recovery techniques in this method.

4.2 The selected subjects perform a simulated surgical scrub under the supervision of an individual competent in aseptic technique. One hand of each subject is sampled immediately after the scrub (within 1 min), and the other hand, 6 h after scrubbing. Only one hand of a subject is sampled at a specified time. Optionally, another sampling time, 3 h for example, can be added between the immediate and 6 h sampling times. If this is desired, the panel size must be increased by 50 % to obtain the same number of data points at each designated sampling interval. Also, a sampling time randomization must be generated such that one-third of the hands are sampled at each sampling interval with only one hand of a subject being sampled at a sampling time interval.

~~4.3 If demonstration of cumulative activity is desired, eleven additional scrubs are performed over a 5-day period, one additional time on Day 1, three times on Days 2, 3, and 4 and once on Day 5. The hands are sampled again after the last scheduled scrub.~~

NOTE1—The researcher should be cautioned that components of chemical neutralizer systems such as lecithin and polysorbate 80 may interfere in the determination of cumulative effect on the skin.” 1—Data for submission to some regulatory bodies may require the addition of a positive and negative control in addition to the test product. For the negative control, 0.9 % saline can be used when testing alcohol products and the product vehicle can be used as the negative control when testing non-alcoholic products.

4.3 If demonstration of cumulative activity is desired, eleven additional scrubs are performed over a 5-day period, one additional time on Day 1, three times on Days 2, 3, and 4 and once on Day 5. The hands are sampled again after the last scheduled scrub.

5. Significance and Use

5.1 The procedure in this test method should be used to evaluate the activity of the test formulation in reducing the bacterial population of the hands immediately after a single use and to determine persistent activity (inhibition of growth) after 6 h. Optionally, measurements of persistent activity after a 3 h period and measurements of cumulative activity may be made after repetitive uses over a five day period.

6. Apparatus

6.1 *Colony Counter*—Any of several types may be used, for example, Quebec Colony Counter. —Use any of several types.

6.2 *Incubator*—Any incubator that can maintain a temperature of $30 \pm 2^\circ\text{C}$ may be used. —Any incubator capable of maintaining a temperature of $30 \pm 2^\circ\text{C}$.

⁶ Johnson’s Baby Wash Head-to-Toe® Johnson’s and Johnson’s Inc., Skillman, NJ 08558-9418.

⁶ Available from U.S. Pharmacopeia (USP), 12601 Twinbrook Pkwy., Rockville, MD 20852-1790, <http://www.usp.org>.

6.3 *Sterilizer*—~~Any suitable steam sterilizer that can produce the conditions of sterility is acceptable.~~—Any suitable steam sterilizer capable of producing the conditions of sterilization.

6.4 *Timer (stop-clock)*—~~That can be read for minutes and seconds.~~—one that displays minutes and seconds.

6.5 *Hand Washing Sink*—A sink of sufficient size to permit subjects to wash without touching hands to sink surface or other subjects.

6.5.1 *Water Faucet(s)*—To be located above the sink at a height that permits the hands to be held higher than the elbows during the washing procedure. (It is desirable for the height of the faucet(s) to be adjustable.)

6.6 *Tap Water Temperature Regulator and Temperature Monitor*—To monitor and regulate water temperature to $40 \pm 2^\circ\text{C}$.

7. Reagents and Materials

7.1 *Petri Dishes*—100 by 15 mm. Required for performing Standard Plate Count.

NOTE 2—Pre-sterilized/disposable plastic petri dishes are available from most local laboratory supply houses.

7.2 *Bacteriological Pipets*—10.0 and 2.2 or 1.1-mL capacity.

NOTE 3—Pre-sterilized/disposable bacteriological pipets are available from most local laboratory supply houses.

7.3 *Water-Dilution Bottles*—Any sterilizable container having a 150 to 200-mL capacity and tight closures may be used.

NOTE 4—Dilution bottles of 160-mL capacity having a screw-cap closure are available from most local laboratory supply houses.

7.4 *Cleansing Wash Formulation*—~~A formulation without an active ingredient.~~—A mild, non-antimicrobial soft soap such as the following or any other liquid soap with neutral pH which does not contain an antimicrobial:

Soft soap, 200 g/L

Linseed oil

Potassium hydroxide

Ethanol

Distilled or high purity water

50 parts by weight

9.5 parts

7 parts

as needed

7.4.1 Add linseed oil to a solution of potassium hydroxide in 15 parts water and heat up to approximately 70°C while constantly stirring. Add the ethanol and continue heating while stirring until the saponification process is completed and a sample dissolves clearly in water and almost clearly in alcohol. The weight of the soft soap is then brought up to 100 parts by addition of hot water. Take 200 g of the soft soap in 1 L of water. Dispense in to appropriate containers and sterilize in an autoclave.

7.5 *Gloves for Sampling*—Loose-fitting, unlined, powder-free latex gloves which do not demonstrate antimicrobial activity, or equivalent glove. An equivalent is a glove composed of any material that is unlined, does not leak and does not demonstrate antimicrobial activity. A zone of inhibition test such as AATCC Test Method 147 may be used to evaluate the antibacterial activity.

7.6—Loose-fitting, unlined, powder-free latex gloves which possess no antimicrobial properties,⁷ or equivalent. 15-10

NOTE 5—A zone of inhibition test such as AATCC 147–2004, Test Method E2180, or Japanese Standard JIS Z 2801 may be used to evaluate antimicrobial properties of gloves.

7.6 *Test Formulation*—Directions for use of active test formulation should be utilized if available. If not available, use directions provided in this test method (see 11.3).

7.7 *Water*—Sterile deionized water or equivalent (Specification D1193, Type III).

7.8 *Sampling Fluid*⁸—Dissolve 0.4 g KH_2PO_4 , 10.1 g Na_2HPO_4 and 1.0 g isooctylphenoxypolyethoxyethanol¹³ in 1 L of water. Adjust to obtain a final pH 7.8 ± 0.1 . Dispense to achieve a final volume of 75 ± 1 mL into water dilution bottles, or other suitable containers, and sterilize. Optionally, the sampling fluid may be sterilized before dispensing into sterile containers. The ability of the sampling fluid to neutralize or quench the antimicrobial activity of the test formulation must be validated (see Practices E1054). The validation test should be conducted *in vivo* in accordance with how the surgical hand scrub study is conducted. If the sampling fluid does not quench the antimicrobial activity of the test formulation and the internal reference formulation, an antimicrobial inactivator should be included if required., 1.0 g isooctylphenoxypolyethoxyethanol (for example, Triton X-100), and appropriately validated neutralizers in 1 L distilled water. Adjust pH to 7.8 ± 1 with 0.1 N HCl or 0.1 N NaOH. Dispense to achieve a final volume of 75 ± 1 mL and sterilize.

7.9 *Dilution Fluid*—Butterfield's buffered phosphate diluent adjusted to pH 7.2 ± 0.1 (or other suitable diluent) and containing

⁷ Bently, M.V., Kedor, E.R., Vianna, R.F., Collett, J.H., Influence of lecithin and urea on the in vitro permeation of hydrocortisone acetate through skin from hairless mouse. *International Journal of Pharmaceutics*. Vol. 146: 255–262.

⁸ The sole source of supply of the apparatus (Ansell #579500, sterile, Encore Acclaim Latex Surgical Gloves) known to the committee at this time is PSS Medical, Inc. (Cat #105613). If you are aware of alternative suppliers, please provide this information to ASTM International Headquarters. Your comments will receive careful consideration at a meeting of the responsible technical committee, which you may attend.

¹³ Kato, A., Ishibahi, Y., Effect of egg yolk lecithin on transdermal delivery of bunazosin. *Journal of Pharmacy and Pharmacology*. Vol 39: 399–400.

¹⁴ Peterson, A. F., "The Microbiology of the Hands: Evaluating the Effects of Surgical Scrubs," *Developments in Industrial Microbiology*, Vol 14, 1973, pp. 125–130.