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**Mikrobiologija v prehranski verigi - Ugotavljanje in uporaba kardinalnih vrednosti
(ISO/DIS 23691:2024)**

Microbiology of the food chain - Determination and use of cardinal values (ISO/DIS 23691:2024)

Mikrobiologie der Lebensmittelkette - Bestimmung und Verwendung von Kardinalwerten (ISO/DIS 23691:2024)

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Microbiologie de la chaîne alimentaire - Détermination et utilisation des valeurs cardinales (ISO/DIS 23691:2024)

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Microbiology of the food chain — Determination and use of cardinal values

*Microbiologie de la chaîne alimentaire — Détermination et
utilisation des valeurs cardinales*

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Foreword

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The procedures used to develop this document and those intended for its further maintenance are described in the ISO/IEC Directives, Part 1. In particular, the different approval criteria needed for the different types of ISO documents should be noted. This document was drafted in accordance with the editorial rules of the ISO/IEC Directives, Part 2 (see www.iso.org/directives).

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Introduction

Under the general principles of the Codex Alimentarius on food hygiene, it is the responsibility of the Food Business Operators (FBO) to control microbiological hazards in foods and to manage microbial risks. Therefore, the FBO shall implement validated control measures, within the hazard analysis and critical control point (HACCP) system, and conduct studies in order to investigate compliance with the food safety criteria throughout the food chain.

In the framework of Microbial Risk Assessment (MRA), several complementary approaches are developed to estimate risks posed by pathogens or spoilage microorganisms in the food chain. MRA is adopted by regulators under the auspices of the international agency for setting food standards. Predictive Microbiology is one of the recognized scientific approaches used to validate control measures within the HACCP system, as well as to assess microbiological safety and quality of food, food production processes, food storage conditions, and food preparation recommendations dedicated to consumers.

Therefore, this document provides technical rules, procedures and calculations to estimate the cardinal values of a microorganism of concern and use them in combination with challenge tests results to simulate and predict its growth in the raw materials, intermediate or end-products under reasonably foreseeable food processes, storage and use conditions.

To do so, different sections are developed:

- to identify the environmental factor(s) in scope (e.g. Temperature, pH, a_w , organic acids),
- to define the appropriate experimental design,
- to estimate the cardinal values of a microorganism in broth medium,
- to perform a challenge test in the matrix of interest and derive the food correction factor and the maximum microbial population density,
- to use the cardinal values and the food correction factor to predict the growth of the studied microorganism in different conditions of interest (e.g. changes in time and temperature throughout the chill chain, changes in formulation with addition of organic acids or preservatives).

Regulatory authorities may have specific recommendations, and these differences have been included as much as possible in this document. It is however possible that additional requirements need to be incorporated to get a regulatory approval of the study.

The use of the ISO 23691 involves expertise in relevant fields such as food microbiology, predictive microbiology and statistics. This expertise encompasses an understanding of sampling theory and design of experiments, statistical analysis of microbiological data and overview of scientifically recognized and available mathematical concepts used in predictive microbiology.

Microbiology of the food chain — Determination and use of cardinal values

WARNING — In order to safeguard the health of laboratory personnel, it is essential that tests for detecting *target microorganism(s)* are only undertaken in properly equipped laboratories, under the control of a skilled microbiologist, and that great care is taken in the disposal of all incubated materials. Persons using this document should be familiar with normal laboratory practice. This document does not purport to address all of the safety aspects, if any, associated with its use. It is the responsibility of the user to establish appropriate safety and health practices.

1 Scope

This document establishes basic principles and specifies requirements and methods to determine the cardinal values of bacteria and yeast strains and use them to predict microbial growth.

Four main steps are required: (1) Determination of the cardinal values in culture medium, (2) Determination of the correction factor in the target food, (3) Validation of the model and (4) Simulations.

Four environmental factors are considered: temperature, pH, a_w and inhibitors (e.g. organic acids).

NOTE Microbial competition is not considered as an inhibitor in this standard and can be addressed by proper modelling approaches.

The determination of cardinal values requires a two-step approach:

- the determination of maximum specific growth rates of the studied strain grown in broth under a defined range of values of the studied environmental factor(s), and
- the use of recognized predictive microbiology secondary models to fit the obtained experimental data to obtain the cardinal values.

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The use of cardinal values in microbial growth simulation is based on predictive microbiology primary and secondary models. The cardinal values are combined with challenge test data to consider the matrix effect. Depending on the goal of the growth simulation, it is important to account for variation of cardinal values between strains within a bacterial or yeasts species.

Cardinal values are a good indicator of a strain growth ability for the studied environmental factors. They are therefore used as criteria to select strains, in addition to their origin and virulence, when performing growth challenge tests (standard ISO 20976-1) or in methods validation (ISO 16140 standards serie).

NOTE This document focuses on the determination of cardinal values for one strain. The same methodology can be used to characterize multiple strains independently to cover biological strain variability and include these results in the predictions.

2 Normative references

The following documents are referred to in the text in such a way that some or all of their content constitutes requirements of this document. For dated references, only the edition cited applies. For undated references, the latest edition of the referenced document (including any amendments) applies.

ISO 7218, *Microbiology of food and animal feeding stuffs — General requirements and guidance for microbiological examinations*

ISO 11133, *Microbiology of food, animal feed and water — Preparation, production, storage and performance testing of culture media*

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3.10

extrinsic factor

factor in the surrounding environment of the food or the broth, such as temperature or packaging gaseous composition, which affects the growth kinetics of the microorganism

3.11

gamma concept γ

the gamma concept establishes that intrinsic (e.g. pH, water activity, inhibitors) and extrinsic factors (e.g. temperature, packaging gaseous composition) affect the maximum specific growth rate independently, using linear and or non-linear functions e.g. γ (temperature), γ (pH), γ (water activity), γ (inhibitors), normalized between zero (no growth) and one (optimum condition for growth). When combined, the effect of the factors is multiplicative

3.12

gamma function $\gamma(X)$

non linear, dimensionless function, normalized between zero (no growth) and one (optimum condition for growth) describing the relative effect of a studied factor (X) on the maximum specific growth rate (e.g. γ (temperature), γ (pH), γ (water activity), γ (inhibitors))

3.13

growth curve

graphic representation of the increasing number of living cells of a microbial population in any given intrinsic and extrinsic condition over a period of time

3.14

inoculum

microbial suspension used to contaminate the studied food or broth at a desired concentration

3.15

intrinsic factor

factor related to the food matrix itself or the broth, such as nutrients, water activity, organic acids or pH, and which affects the growth kinetics of the micro-organism

3.16

lag phase

phase, directly after inoculation, during which the microbial population is adapting to the environment, before it enters the *exponential growth phase* (3.9)

3.17

lag time λ

kinetic parameter in time unit to characterize the duration of the *lag phase* (3.16)

3.18

maximum specific growth rate μ_{\max}

kinetic parameter (h^{-1}) to characterize the *exponential growth phase* (3.9), represented by the slope of the curve showing the evolution of the natural logarithm of the population as a function of time, under constant growth conditions When the maximum specific growth rate is estimated in food, this is noted $\mu_{\max, \text{food}}$

3.19

Minimal Inhibitory Concentration**MIC**

estimated parameter representing the lowest concentration of an inhibitor that gives a value of maximum specific growth rate of zero

ISO/DIS 23691:2024(en)**3.20****modified broth**

culture medium with specific composition (e.g. increased salt content) or characteristic (e.g. pH) to study intrinsic factors

3.21**Monte Carlo simulation**

iterative random sampling method that propagates variability about model parameters to approximate the distribution of input variables. Monte Carlo simulations are extensively used in quantitative risk assessment and decision making

3.22**optimum growth rate**

$\hat{\mu}$

highest value among the maximum specific growth rates, estimated at the optimum conditions for growth of the microorganism in a studied food or broth

3.23**optimum growth rate in broth**

$\hat{\mu}_{Broth}$

highest value among the maximum specific growth rates in broth, estimated at the optimum conditions for growth of the microorganism

3.24**optimum growth rate in food**

$\hat{\mu}_{Food}$

highest value among the maximum specific growth rates in the food, estimated at the optimum conditions for growth of the microorganism

3.25

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organizing laboratories

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laboratories with responsibility for determining the *cardinal values* (3.4) and performing the simulations. Data collection and data analysis (including fitting and simulation) are performed in a single or in multiple laboratories.

3.26**pH value**

measure of the concentration of acidity or alkalinity of a material in an aqueous solution

[SOURCE: ISO 5127:2017, 3.12.2.29, modified — Notes 1 and 2 to entry have been removed.]

3.27**pKa**

quantitative measure (negative base-10 logarithm) of the acid dissociation constant or Ka value, which indicates the strength of an acid in solution (the lower the pKa value the stronger the acid)

3.28**primary model**

mathematical model describing the changes of microbial concentration as a function of time under constant and known conditions of intrinsic and / or extrinsic factor(s)

3.29**relative standard error**

r

standard error (se) (3.31) divided by the parameter estimate and expressed as a percentage

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3.30**secondary model**

mathematical model describing the effects of the intrinsic and / or extrinsic factor(s) (e.g. temperature, pH, a_w) on the parameters of the *primary model* (3.28) (e.g. maximum specific growth rate)

3.31**standard error****se**

measure of the uncertainty associated with the estimated parameter or the overall model fit

3.32**stationary phase**

phase in which the microbial population no longer increases (reaches and remains at its maximum concentration)

3.33**strong acid**

is characterized by its negative pKa. It ionizes completely in an aqueous solution by losing one proton. Hydrochloric and sulfuric acids are examples of strong acids

3.34**uncertainty**

refers to variation that originates from lack of or incomplete knowledge of some characteristics of a system. It originates from parameter uncertainty and model uncertainty. Sources of parameter uncertainty include lack of data, measurement errors, sampling errors and systematic errors. Sources of model uncertainty include model structure, excluded variables, model resolution, extrapolation. The standard error represents the uncertainty associated with the parameter

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refers to variation that is inherent to a given system, typically as a result of true heterogeneity of the studied population and is irreducible by additional measurement. Three variation sources are distinguished: between strain variability (intraspecies variability), within strain variability and analytical variability. The between strain variability is not included in this standard as it is designed to study only one strain at a time. The standard deviation represents the within strain biological variability associated with the parameter

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3.36**water activity**

a_w
ratio of the water-vapor pressure in the medium or foodstuff to the vapor pressure of pure water at the same temperature. It represents the water available for the microorganisms to use

[SOURCE: ISO 18787:2017, 3.1, modified — The definition has been condensed and the formula and Notes 1 and 2 to entry have been removed. The sentence “It represents the water available for the microorganisms to use” was added.]

3.37**weak acid**

is characterized by its high pKa. It does not dissociate completely in aqueous solution. Acetic acid and citric acid are examples of weak acids

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4 Principle

4.1 General

The general equation used to describe the effect of different independent intrinsic and extrinsic factors on the maximum specific growth rate of a microorganism is based on a modular approach called the "gamma concept"^[28] and described in [Equation \(1\)](#).

$$\mu_{\max} = \dot{\mu} \cdot \gamma(T) \cdot \gamma(\text{pH}) \cdot \gamma(a_w) \cdot \gamma(\text{inhibitors}) \quad (1)$$

where

μ_{\max}	maximum specific growth rate (h^{-1}) of the studied strain in the matrix;
$\dot{\mu}$	optimum growth rate (h^{-1}) of the studied strain in the matrix;
$\gamma(T)$	dimensionless function describing the relative effect of the Temperature on microbial growth;
$\gamma(\text{pH})$	dimensionless function describing the relative effect of the pH on microbial growth;
$\gamma(a_w)$	dimensionless function describing the relative effect of the a_w on microbial growth;
$\gamma(\text{inhibitors})$	dimensionless functions describing the relative effect of different measurable inhibitors like the undissociated form of the weak (organic) acids (HA) or CO_2 .

The γ terms all vary between 0 and 1, $\gamma = 0$ when growth is fully inhibited by the studied factor, and $\gamma = 1$ when growth is not at all inhibited by the studied factor.

There are various secondary models available in the literature to describe the mathematical expression of the gamma terms. In this standard, the cardinal models are used and presented in [4.2](#).

For the adequate use of the models and interpretation of data, knowledge of and experience in using predictive microbiology models is essential.

4.2 Mathematical models

Under the gamma concept, the different intrinsic and extrinsic factors (e.g. temperature, pH, water activity, inhibitors) have separate and independent effects on the maximum specific growth rate, which implies that the cardinal values associated with a factor are also estimated separately and independently.

Various mathematical models have been developed in the literature.

- For describing the effects of temperature, one of the two following models shall be used: the CTMI (Cardinal Temperature Model with Inflection) model ([Equation 2](#)) shall be used when optimal and super-optimal temperatures are required^[25] while the restricted Ratkowsky (linear) model ([Equation 3](#))^[21] shall be used when the input temperature ranges from the minimum supporting growth up to a reference temperature that is below the optimal temperature.