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Fortified milk powders, infant formula and adult nutritionals — Determination of total biotin by liquid chromatography coupled with immunoaffinity column clean-up extraction

Poudres de lait fortifié, formules infantiles et produits nutritionnels pour adultes — Détermination de la teneur en biotine totale par chromatographie liquide après une purification sur colonne d'immunoaffinité

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Foreword

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The procedures used to develop this document and those intended for its further maintenance are described in the ISO/IEC Directives, Part 1. In particular, the different approval criteria needed for the different types of ISO documents should be noted. This document was drafted in accordance with the editorial rules of the ISO/IEC Directives, Part 2 (see www.iso.org/directives).

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For an explanation of the voluntary nature of standards, the meaning of ISO specific terms and expressions related to conformity assessment, as well as information about ISO's adherence to the World Trade Organization (WTO) principles in the Technical Barriers to Trade (TBT) see www.iso.org/iso/foreword.html.

This document was prepared by Technical Committee ISO/TC 34, *Food products*, in collaboration with AOAC INTERNATIONAL. It is being published by ISO and separately by AOAC INTERNATIONAL. The method described in this document is equivalent to the AOAC Official Method 2016.02: *Determination of Total Biotin by Liquid Chromatography Coupled with Immunoaffinity Column Cleanup Extraction: Multilaboratory Testing, Final Action 2016.02.*

Any feedback or questions on this document should be directed to the user's national standards body. A complete listing of these bodies can be found at www.iso.org/members.html.

Fortified milk powders, infant formula and adult nutritionals — Determination of total biotin by liquid chromatography coupled with immunoaffinity column clean-up extraction

WARNING — The use of this method can involve hazardous materials, operations and equipment. This method does not purport to address all the safety problems associated with its use. It is the responsibility of the user of this method to establish appropriate safety and health practices and determine the applicability of regulatory limitations prior to use.

1 Scope

This document specifies a method for the quantitative determination of biotin and/or biocytin in fortified milk powders, infant formula and adult nutritionals in solid (i.e. powders) or liquid (i.e. ready-to-feed liquids and liquid concentrates) forms using liquid chromatography coupled with immunoaffinity column clean-up extraction.

Precision data from an interlaboratory study is given in Annex B. A comparison between data obtained with the method in this document and EN 15607 is given in Annex C.

2 Normative references

There are no normative references in this document.

3 Terms and definitions

For the purposes of this document, the following terms and definitions apply.

ISO and IEC maintain terminological databases for use in standardization at the following addresses:

— ISO Online browsing platform: available at https://www.iso.org/obp

— IEC Electropedia: available at http://www.electropedia.org/

3.1
adult nutritional
nutritionally complete, specially formulated food, consumed in liquid form, which may constitute the sole source of nourishment, made from any combination of milk, soy, rice, whey, hydrolysed protein, starch and amino acids, with and without intact protein

3.2
infant formula
breast-milk substitute specially manufactured to satisfy, by itself, the nutritional requirements of infants during the first months of life up to the introduction of appropriate complementary feeding

[SOURCE: Codex Standard 72-1981]

4 Principle

The sample is dispersed in sodium phosphate buffer and autoclaved at 121 °C ± 2 °C for 25 min. The sample is cooled to room temperature and then diluted to 100 ml in a volumetric flask. The extract is centrifuged and filtered using a glass microfibre filter. Clear filtrate is collected for clean-up and

extraction. A biotin immunoaffinity column is mounted onto a solid phase extraction (SPE) manifold. A disposable syringe barrel is connected to the immunoaffinity column as a reservoir. The buffer in the affinity column is drained and the sample filtrate is loaded through the reservoir and allowed to flow through by gravity. The column is washed with phosphate-buffered saline (PBS) followed by water. Air is passed through the column to remove residual liquid.

Biotin and/or biocytin from the column is eluted with methanol and collected in a vial. The eluate is evaporated to dryness using a heating block set at 85 °C ± 5 °C under a gentle stream of nitrogen and the sample is re-constituted in 1 ml of water. The biotin and biocytin in the reconstituted sample are analysed simultaneously by HPLC using a photodiode array (PDA) set at 200 nm. Identification of peaks is based on absolute retention time. Quantification is by multipoint external calibration using peak area responses of the analytes. A spectrum scan (200 nm to 350 nm) can be used for the purity and identity confirmation as required.

5 Reagents and materials

5.1 General

During the analysis, unless otherwise stated, only use reagents of recognized analytical grade and distilled or demineralized water or water of equivalent purity.

5.1.1 Laboratory reagent grade water.

5.1.2 Sodium dihydrogen phosphate dihydrate (CAS # 13472-35-0).

5.1.3 Disodium hydrogen phosphate dihydrate (CAS # 10028-24-7).

5.1.4 Sodium hydroxide (CAS #1310-73-2).

5.1.5 Methanol, HPLC grade (CAS # 67-56-1).

5.1.6 Acetonitrile, HPLC grade (CAS # 75-05-8).

5.1.7 Ortho-phosphoric acid, mass fraction w (H_3PO_4) = 85 % (CAS # 7664-38-2).

5.1.8 PBS, pH = 7,4 (Thermo Scientific[1)] or equivalent).

5.1.9 Biotin, purity ≥ 99 % (Cat. No. B4501 Sigma Chemical[1)] or equivalent).

5.1.10 Biocytin, purity ≥ 98 % (Cat. No. B4261 Sigma Chemical[1)] or equivalent).

5.2 Reagent preparation

5.2.1 Sodium hydroxide, substance concentration c = 2 mol/l.

Weigh 80 g of sodium hydroxide into a 1 l volumetric flask, dissolve and make up to the mark with water.

1) This is an example of a suitable product available commercially. This information is given for the convenience of users of this document and does not constitute an endorsement by ISO of this product. Equivalent products may be used if they can be shown to lead to the same results.

5.2.2 Sodium phosphate buffer, c = 0,15 mol/l.

Weigh 9,15 g of sodium dihydrogen phosphate dihydrate and 16,31 g of disodium hydrogen phosphate dihydrate into a 1 l volumetric flask, dissolve and make up to the mark with water. Adjust the pH to 7 with 2 mol/l sodium hydroxide.

5.2.3 Phosphoric acid, w = 0,1 %.

Into a 1 l volumetric flask, transfer 500 ml water. Add 1,2 ml of ortho-phosphoric acid. Mix and make up to the mark with water.

5.2.4 Mobile phase A, 0,1 % phosphoric acid in water.

5.2.5 Mobile phase B, 100 % acetonitrile.

5.2.6 Mobile phase C, 80 % acetonitrile.

5.3 Standard preparation

5.3.1 Stock standard biotin, mass concentration ρ = 100 µg/ml.

Weigh 25 mg biotin standard in a 250 ml amber volumetric flask. Add 150 ml water and sonicate at room temperature for 90 min with occasional shaking. Make up to volume with water.

5.3.2 Stock standard biocytin, ρ = 100 µg/ml.

Weigh 10 mg biocytin standard in a 100 ml amber volumetric flask. Add 60 ml water and sonicate at room temperature for 90 min with occasional shaking. Make up to volume with water.

5.3.3 Mixed intermediate standard, ρ = 100 µg/100 ml.

Dilute 1 ml each of stock standards to 100 ml with water.

5.3.4 Calibration standard 1, ρ = 1,0 µg/100 ml.

Dilute 100 µl mixed intermediate standard to 10 ml with water.

5.3.5 Calibration standard 2, ρ = 2,5 µg/100 ml.

Dilute 250 µl mixed intermediate standard to 10 ml with water.

5.3.6 Calibration standard 3, ρ = 5,0 µg/100 ml.

Dilute 500 µl mixed intermediate standard to 10 ml with water.

5.3.7 Calibration standard 4, ρ = 7,5 µg/100 ml.

Dilute 750 µl mixed intermediate standard to 10 ml with water.

5.3.8 Calibration standard 5, ρ = 10 µg/100 ml.

Dilute 1 ml mixed intermediate standard to 10 ml with water.

5.3.9 Calibration standard 6, ρ = 20 µg/100 ml.

Dilute 2 ml mixed intermediate standard to 10 ml with water.

5.4 Calculation of concentration

The concentrations given in 5.3 are indicative only. Calculate the actual concentrations of biotin and biocytin in each calibration standard, in µg/100 ml, using Formula (1). Calibration standards should be bracketed at the beginning and at the end of an analytical run.

$$\rho_{(\text{biotin/biocytin})} = \frac{(m_1 \times P \times 10 \times V_{\text{is}})}{(V \times 10)} \quad (1)$$

where

m_1 is the mass of biotin or biocytin, in mg;

P is the percentage purity from the certificate of analysis or verified by USP/BP/Ph Eur monographs;

V_{is} is the volume of mixed intermediate standard used for the calibration standard, in ml;

V is the volume of stock standard, V = 250 ml for biotin and V = 100 ml for biocytin.

6 Apparatus

Usual laboratory glassware and equipment and, in particular, the following.

6.1 HPLC system, consisting of a PDA detector, low pressure gradient pump system, a sample injector unit, a degasser unit, and a column oven.

6.2 Column, Kinetex Phenyl-Hexyl (Cat. No. 00F-4495-E0, Phenomenex[2)]), 150 mm × 4,6 mm × 2,6 µm × 10 nm.

6.3 Glass microfibre filters (Cat. No. 1820-125, Whatman®[2)]).

6.4 Immunoaffinity column pack (R-Biopharm Rhone EASI-EXTRACT® BIOTIN P82/P82B[2)] or equivalent).

6.5 SPE manifold, with accessories.

6.6 Autoclave, set at 121 °C.

6.7 Centrifuge, variable speed.

6.8 Analytical balance, four decimal places.

6.9 Amber glass screw-cap bottle, 100 ml.

6.10 Horizontal shaker.

6.11 Volumetric flasks, 1 l, 250 ml, 100 ml and 10 ml.

6.12 Pipettors, calibrated, 10,0 ml, 5,0 ml, 1,0 ml, 200 µl, 100 µl and 50 µl.

2) This is an example of a suitable product available commercially. This information is given for the convenience of users of this document and does not constitute an endorsement by ISO of this product. Equivalent products may be used if they can be shown to lead to the same results.

6.13 Measuring cylinder, 1000 ml, 100 ml and 50 ml.

6.14 Reaction vial, Reacti-Vials (Cat. No. 13223, Thermo Scientific[2)]).

6.15 Heating block, Reacti-therm, with nitrogen blow down (Thermo Scientific[2)]).

6.16 Ultrasonic bath, set at 50 °C and room temperature.

6.17 Centrifuge tubes, 50 ml.

6.18 Vortex mixer.

6.19 Syringe filter, polytetrafluoroethylene (PTFE) 0,45 µm.

6.20 Disposable syringes, 10 ml and 1 ml.

6.21 HPLC vials, 2 ml with 200 µl glass inserts.

7 Procedure

7.1 Sample preparation

7.1.1 For mass and loading volumes for the different ranges of product, see Table 1. A slurry may be used wherever product homogeneity is suspected or unknown.

For the slurry, reconstitute 25 g of powder (m_1) with warm water (~50 °C) to a total mass of 200 g (m_2). Mix thoroughly on a horizontal shaker for 20 min and then sonicate at 50 °C for 10 min. Cool to room temperature. For liquid samples, mix well to ensure homogeneity of the sample portion and weigh the specified quantity.

7.1.2 Weigh the sample/slurry (m_3) into a 100 ml amber glass screw-cap bottle. See Table 1.

7.1.3 Add 0,15 mol/l sodium phosphate buffer to an approximate volume of 50 ml.

7.1.4 Swirl gently to mix.

7.1.5 Autoclave the sample preparation at 121 °C for 25 min.

7.1.6 Cool the sample to room temperature. Quantitatively transfer the extract into a 100 ml volumetric flask and make up to the mark with 0,15 mol/l sodium phosphate buffer, mix well.

7.1.7 Transfer the extract into a centrifuge tube and centrifuge the sample at 4 000 rpm for 15 min.

7.1.8 Filter the sample using the glass microfibre filter paper (6.3) and collect the filtrate.

7.1.9 Set up the SPE manifold (6.5). Attach the immunoaffinity column (IAC) connected to a 10 ml reservoir. Drain off buffer just above the gel.

7.1.10 Load the sample filtrate onto the column in accordance with Table 1 and initialize the flow with the help of a vacuum pump.

7.1.11 Turn off the vacuum and let the solution pass through the column by gravity at a rate of one drop per second.

7.1.12 Wash the column by passing 10 ml of PBS (5.1.8) through the column, followed by 10 ml of water. Initialize the flow with the help of vacuum at every step and then leave it to flow by gravity.

7.1.13 Remove any residual liquid from the column by introducing a gentle vacuum.

7.1.14 Introduce a reaction vial (6.14) and elute the analyte under gravity with 2 ml methanol. Elute further with an additional 1 ml of methanol. Backflush at least three times when eluting; this can be achieved by a gentle up and down motion of the syringe plunger to maximize the elution.

7.1.15 Evaporate the eluate to dryness using a heating block (6.15) set at 85 °C ± 5 °C under a gentle nitrogen blow down.

7.1.16 Remove from the heating block and cool down to room temperature (about 15 min).

7.1.17 Re-dissolve with 1 ml water and then cap the reaction vial (6.14) and vortex for 30 s. Filter by using a syringe filter into a clean glass insert in a HPLC vial for the HPLC analysis.

Table 1 — Sample mass, dilution and loading volume

Biotin µg/100g		**Sample preparation**				**Conc** µg/100 ml	
Min.	**Max.**	**Mass** (g)	**Volume** (ml)	**Load** (ml)	**Final**	**Min.**	**Max.**
0,1	0,5	20	100	50	1 ml	1	5
0,5	1,0	10	100	20	1 ml	1	2
1,0	5,0	10	100	10	1 ml	1	5
5,0	50,0	2,0 (Slurry 16 g)	100	10	1 ml	1	10
50,0	100,0	1,0 (Slurry 8 g)	100	10	1 ml	5	10
100,0	400,0	0,5 (Slurry 4 g)	100	5	1 ml	2,5	10

7.2 Chromatography

7.2.1 Set-up the HPLC system with the following configuration. Examples of chromatograms of a calibration standard and an infant formula sample used in the interlaboratory study are given in Annex A.

7.2.2 **Mobile phase A,** 0,1 % phosphoric acid.

7.2.3 **Mobile phase B,** 100 % acetonitrile.

7.2.4 **Mobile phase C,** 80 % acetonitrile.

7.2.5 **Column,** see 6.2.

7.2.6 **Column temperature,** 25 °C ± 2 °C.

7.2.7 **Retention times,** biocytin is 4,5 min to 5,5 min and biotin is 16 min to 17 min.

7.2.8 **Run time,** 27 min.

7.2.9 **Detector,** a PDA detector operating at 200 nm (spectrum scan 200 nm to 350 nm).