



Designation: C1473 – 11

Standard Test Method for Radiochemical Determination of Uranium Isotopes in Urine by Alpha Spectrometry¹

This standard is issued under the fixed designation C1473; the number immediately following the designation indicates the year of original adoption or, in the case of revision, the year of last revision. A number in parentheses indicates the year of last reapproval. A superscript epsilon (ϵ) indicates an editorial change since the last revision or reapproval.

1. Scope

1.1 This test method is applicable to the determination of uranium in urine at levels of detection dependent on sample size, count time, detector background, and tracer yield. It is designed as a screening tool for detection of possible exposure of occupational workers.

1.2 This test method is designed for 50 mL of urine. This test method does not address the sampling protocol or sample preservation methods associated with its use.

1.3 The values stated in SI units are to be regarded as standard. No other units of measurement are included in this standard.

1.4 *This standard does not purport to address all of the safety concerns, if any, associated with its use. It is the responsibility of the user of this standard to establish appropriate safety and health practices and determine the applicability of regulatory limitations prior to use.*

2. Referenced Documents

- 2.1 *ASTM Standards*:²
- C859 Terminology Relating to Nuclear Materials
 - C1000 Test Method for Radiochemical Determination of Uranium Isotopes in Soil by Alpha Spectrometry
 - C1163 Practice for Mounting Actinides for Alpha Spectrometry Using Neodymium Fluoride
 - C1284 Practice for Electrodeposition of the Actinides for Alpha Spectrometry
 - D1193 Specification for Reagent Water
 - D3084 Practice for Alpha-Particle Spectrometry of Water
 - D3648 Practices for the Measurement of Radioactivity

¹ This test method is under the jurisdiction of ASTM Committee C26 on Nuclear Fuel Cycle and is the direct responsibility of Subcommittee C26.05 on Methods of Test.

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² For referenced ASTM standards, visit the ASTM website, www.astm.org, or contact ASTM Customer Service at service@astm.org. For *Annual Book of ASTM Standards* volume information, refer to the standard's Document Summary page on the ASTM website.

3. Summary of Test Method

3.1 A urine sample with ²³²U tracer solution added is wet-ashed with nitric acid and hydrogen peroxide to destroy organic material. The uranium-bearing solution is converted to a hydrochloric acid medium. Uranium is absorbed on an anion exchange column from a 9 M hydrochloric acid solution and eluted with 0.1 M hydrochloric acid solution. The separated uranium is prepared for alpha spectrometric measurement either by electrodeposition onto a metal disk or coprecipitation with neodymium fluoride and filtration onto a membrane filter.

4. Significance and Use

4.1 This test method is used to detect possible exposures to uranium isotopes from occupational operations.

5. Interferences

5.1 The presence of ²³²U in the urine sample will be masked by the tracer addition. The alpha energies of ²³³U and ²³⁴U cannot be fully resolved by alpha spectrometric measurement. A table of uranium isotope alpha energies is given in **Appendix X1**. If neptunium is present in the sample in the plus four oxidation state, it will co-elute with the uranium.³⁻¹¹

6. Apparatus

6.1 *Alpha Spectrometry System*—Refer to Test Method C1000, and Practices C1163, D3084, and D3648 for guidance.

6.2 *Electrodeposition Apparatus*—Refer to Practice C1284 for guidance.

6.3 *Neodymium Fluoride Precipitation*—Refer to Practice C1163 for guidance.

6.4 *Borosilicate Beakers or Flasks*, 250 mL.

6.5 *Borosilicate Beakers*, 150 and 250 mL.

6.6 *Borosilicate Graduated Glass Cylinders*, 5, 25, 100, and 1000 mL.

6.7 *Ion Exchange Columns*, disposable polypropylene, with polyethylene frit, ≥ 5 -mL capacity.

6.8 *Ion Exchange Column Reservoir*, funnel, polypropylene, 100-mL capacity.

NOTE 1—See Fig. 1 for a typical ion exchange column-reservoir setup.

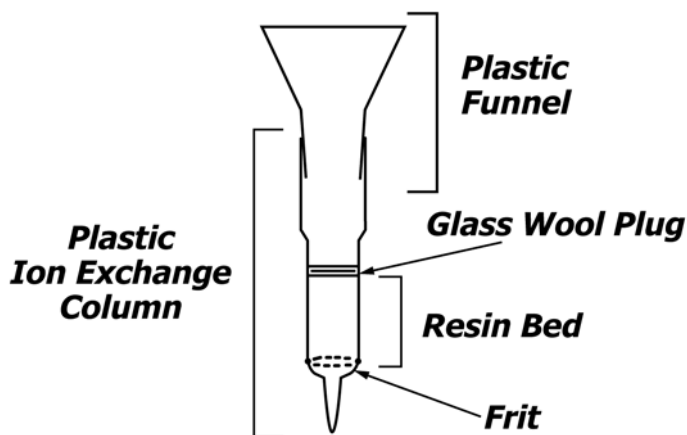


FIG. 1 Typical Ion Exchange Column Arrangement

7. Reagents

7.1 *Purity of Reagents*—Reagent grade chemicals shall be used in all tests. Unless otherwise indicated, it is intended that the reagents conform to the specifications of the Committee on Analytical Reagents of the American Chemical Society, where such specifications are available.³ Other grades may be used, provided it is first ascertained that the reagent is of sufficiently high purity to permit its use without lessening the accuracy of the determination.

7.2 *Purity of Water*—Unless otherwise indicated, references to water shall be understood to mean reagent water as defined by Type III of Specification D1193.

7.3 Reagent purity shall be such that the measured radioactivity of blank samples is minimized.

7.4 *Anion Exchange Resin*—Analytical grade, Type 1, 8 % cross linked, 100-200 mesh, chloride form.

7.5 *Hydrochloric Acid (9 M)*—Mix 750 mL of concentrated HCl with water and dilute to 1 L.

7.6 *Hydrochloric Acid (sp gr 1.19)*—Concentrated hydrochloric acid (HCl).

7.7 *Hydrochloric Acid (0.1 M)*—Mix 8.3 mL of concentrated HCl with water and dilute to 1 L.

7.8 *Hydrogen Peroxide (30 %)*.

7.9 *Nitric Acid (8 M)*—Mix 500 mL of concentrated nitric acid with water and dilute to 1 L.

7.10 *Nitric Acid (sp gr 1.4)*—Concentrated nitric acid (HNO₃).

7.11 *Uranium-232*, standard solution.⁴

³ *Reagent Chemicals, American Chemical Society Specifications*, American Chemical Society, Washington, DC. For suggestions on the testing of reagents not listed by the American Chemical Society, see *Analar Standards for Laboratory Chemicals*, BDH Ltd., Poole, Dorset, U.K., and the *United States Pharmacopeia and National Formulary*, U.S. Pharmacopeial Convention, Inc. (USPC), Rockville, MD.

⁴ Such tracer radionuclides may be available from respective national standards laboratories as well as commercial vendors that provide radionuclides traceable to their respective national standards laboratories.

8. Hazards

8.1 **Warning**—Adequate laboratory facilities, such as fume hoods and controlled ventilation, along with safe techniques must be used in this procedure. Site-specific policies for the handling of biological materials must be adhered to. Extreme care should be exercised in using hydrofluoric and other hot, concentrated acids. The use of proper gloves is recommended.

9. Sampling

9.1 Collect the urine sample in accordance with the site-specific protocol.

9.2 Preserve the urine sample in accordance with the site-specific protocol.

10. Calibration and Standardization

10.1 A ²³²U standard may be used to determine the detection efficiency of the alpha-spectrometry system which, in turn, can be used to calculate the chemical yield of each sample and the lower limit of detection (LLD) of this test method.

11. Procedure

11.1 Sample Preparation:

11.1.1 Measure 50 mL of urine in a 100-mL graduated cylinder.

11.1.2 Transfer the urine to a 250-mL beaker or flask.

11.1.3 Rinse the cylinder twice with 5 mL of 8 M HNO₃ and add the rinsings to the beaker or flask.

11.1.4 Add an appropriate amount of ²³²U tracer solution to the sample (ca. 0.02 Bq or as prescribed in the site-specific protocol) and swirl the vessel to mix.

11.1.5 Evaporate the contents of the vessel to near dryness on a medium hot plate.

11.1.6 Remove the vessel from the hot plate and cool for 1 min.

11.1.7 Add 10 mL of concentrated HNO₃ to the vessel. Swirl to mix.

11.1.8 Return the vessel to the hot plate and evaporate the solution to near dryness.

11.1.9 Remove the vessel from the hot plate and allow to cool for 1 min.

11.1.10 Slowly add 10 to 15 mL of 30 % hydrogen peroxide to the vessel to cover the residue. Swirl the vessel to mix.

11.1.11 Return the vessel to the hot plate and evaporate the solution to near dryness.

11.1.12 Repeat 11.1.9 and 11.1.10.

11.1.13 Return the vessel to the hot plate and evaporate the solution to obtain a white sample residue.

11.1.14 Remove the vessel from the hot plate and allow to cool for 1 min.

11.1.15 Add 10 to 15 mL of concentrated HCl to the vessel and swirl to mix.

11.1.16 Return the vessel to the hot plate and evaporate the solution to near dryness.

11.1.17 Remove the vessel from the hot plate and allow to cool for 1 min.

11.1.18 Repeat 11.1.15 – 11.1.17 twice more. Proceed to 11.1.19.