# INTERNATIONAL STANDARD

ISO 13885-3

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# **Gel permeation chromatography (GPC)** —

Part 3: Water as eluent

Chromatographie par perméation de gel (GPC) — Partie 3: Eluant à l'eau

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ISO 13885-3:2020

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Fore	word	<b>v</b>
1	Scope	
2	Normative references	
3	Terms and definitions	
4	Principle	
5	Apparatus	
3	5.1 Eluent supply	3
	5.2 Pump	
	5.3 Injection system 5.4 Separation columns	
	5.5 Column temperature control	
	5.6 Detector	
6	Reagents	6
7	Calibration of the apparatus	6
	7.1 General	6
	7.2 Specification for the calibration standard	
	7.3 Preparation of the calibration solutions for injection	7
	<ul><li>7.4 Conditions for calibration runs</li><li>7.5 Measurement of elution volume</li></ul>	7 7
	7.6 Plotting the calibration curve	
8	Sampling (https://standards.iteh.ai)	8
9	Preparation for the test	
,	9.1 Preparation of the injection solution	8
	9.2 Preparation of the apparatus	
10	Analytical parameters ISO 13885-3-2020	
<b>11</b> ta	Data acquisition and evaluation 37f5d6c4-ae98-4b57-a02e-24ae3e5b6fb7/iso-13885-3	-2020 9
	11.1 General	
	11.2 Calculation of the net chromatogram from the raw data	
	11.2.1 Determination of the baseline 11.2.2 Correction of the measured values and of the net chromatogram	
	11.2.3 Evaluation limits	
	11.3 Calculation of the average values	10
	11.4 Calculation of the distribution curves	11
<b>12</b>	Precision	
	12.1 General	
	12.2 Repeatability	
13	Test report	
15	13.1 General	
	13.2 General data on the equipment and settings	
	13.2.1 Data on the equipment used	
	13.2.2 Calibration	
	13.2.3 Evaluation 13.3 Special data on the sample	
Anna	ex A (informative) Conversion of experimental parameters for variant column sizes	
	ex B (informative) Example of a data sheet for a polymer standard	
Ann	ex C (informative) Explanations	18

Bibliography......23

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ISO 13885-3:2020

https://standards.iteh.ai/catalog/standards/iso/37f5d6c4-ae98-4b57-a02e-24ae3e5b6fb7/iso-13885-3-2020

#### Foreword

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The procedures used to develop this document and those intended for its further maintenance are described in the ISO/IEC Directives, Part 1. In particular, the different approval criteria needed for the different types of ISO documents should be noted. This document was drafted in accordance with the editorial rules of the ISO/IEC Directives, Part 2 (see <a href="www.iso.org/directives">www.iso.org/directives</a>).

Attention is drawn to the possibility that some of the elements of this document may be the subject of patent rights. ISO shall not be held responsible for identifying any or all such patent rights. Details of any patent rights identified during the development of the document will be in the Introduction and/or on the ISO list of patent declarations received (see <a href="https://www.iso.org/patents">www.iso.org/patents</a>).

Any trade name used in this document is information given for the convenience of users and does not constitute an endorsement.

For an explanation of the voluntary nature of standards, the meaning of ISO specific terms and expressions related to conformity assessment, as well as information about ISO's adherence to the World Trade Organization (WTO) principles in the Technical Barriers to Trade (TBT), see <a href="https://www.iso.org/iso/foreword.html">www.iso.org/iso/foreword.html</a>.

This document was prepared by Technical Committee ISO/TC 35, *Paints and varnishes*.

A list of all parts in the ISO 13885 series can be found on the ISO website.

Any feedback or questions on this document should be directed to the user's national standards body. A complete listing of these bodies can be found at <a href="https://www.iso.org/members.html">www.iso.org/members.html</a>.

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## Gel permeation chromatography (GPC) —

### Part 3:

### Water as eluent

WARNING — This document can involve hazardous materials, operations or equipment. It does not purport to address all of the safety problems, if any, associated with its use. It is the responsibility of the user to establish appropriate safety and health practices and to ensure compliance with any national regulatory conditions.

#### 1 Scope

This document specifies the determination of the molar-mass distribution and the average molar mass values  $M_{\rm n}$  (number average) and  $M_{\rm w}$  (weight average) of polymers that are soluble in water by gel permeation chromatography (GPC).

NOTE Also known as size exclusion chromatography (SEC).

This method is applicable to neutral polymers and polyanions (e.g. polycarboxylates, polysaccharides, fully hydrolyzed polyvinyl alcohols and high-molecular polyethylene oxides). It is not applicable to polycations [e.g. polyvinylpyrrolidone, polyvinylpyridine, salts of poly(diallyl-N,N-dimethyl-azacyclopentane), chitosan].

Despite good solubility in the mobile phase and even though the chromatograms obtained show good repeatability, it is possible that this method cannot be used with certain polymer types because of specific interactions (e.g. adsorption) within the sample/eluent/column system (see also <u>Clause 12</u>).

The conditions specified in this document are not applicable to the GPC analysis of polymer samples with  $M_{\rm w}$  values greater than  $10^6$  g/mol and/or polymers with elution limits outside the calibration range (see 7.6 and  $10^6$  g/mol and/or polymers with elution limits outside the calibration range (see 1.6 and  $10^6$  g/mol and/or polymers with elution limits outside the calibration range (see 1.6 and  $10^6$  g/mol and/or polymers with elution limits outside the calibration range (see 1.6 and  $10^6$  g/mol and/or polymers with elution limits outside the calibration range (see 1.6 and  $10^6$  g/mol and/or polymers with elution limits outside the calibration range (see 1.6 and  $10^6$  g/mol and/or polymers with elution limits outside the calibration range (see 1.6 and  $10^6$  g/mol and/or polymers with elution limits outside the calibration range (see 1.6 and  $10^6$  g/mol and/or polymers with elution limits outside the calibration range (see 1.6 and  $10^6$  g/mol and/or polymers with elution limits outside the calibration range (see 1.6 and  $10^6$  g/mol and/or polymers with elution limits outside the calibration range (see 1.6 and  $10^6$  g/mol and/or polymers with elution limits outside the calibration range (see 1.6 and  $10^6$  g/mol and/or polymers with elution limits outside the calibration range (see 1.6 and  $10^6$  g/mol and/or polymers with elution limits outside the calibration range (see 1.6 and  $10^6$  g/mol and/or polymers with elution limits outside the calibration range (see 1.6 and  $10^6$  g/mol and/or polymers with elution limits outside the calibration range (see 1.6 and  $10^6$  g/mol and/or polymers with elution limits outside the calibration range (see 1.6 and  $10^6$  g/mol and/or polymers with elution limits outside the calibration range (see 1.6 g/mol and/or polymers).

This document includes no correction methods (e.g. for the elimination of peak broadening). If absolute molar mass values are required, an absolute method (e.g. membrane osmometry for  $M_{\rm n}$  or light scattering for  $M_{\rm w}$ ) can be used.

#### 2 Normative references

The following documents, in whole or in part, are normatively referenced in this document and are indispensable for its application. For dated references, only the edition cited applies. For undated references, the latest edition of the referenced document (including any amendments) applies.

ISO 1513, Paints and varnishes — Examination and preparation of test samples

ISO 4618, Paints and varnishes — Terms and definitions

ISO 15528, Paints, varnishes and raw materials for paints and varnishes — Sampling

#### 3 Terms and definitions

For the purposes of this document, the terms and definitions given in ISO 4618 and the following apply.

ISO and IEC maintain terminological databases for use in standardization at the following addresses:

ISO Online browsing platform: available at <a href="https://www.iso.org/obp">https://www.iso.org/obp</a>

#### ISO 13885-3:2020(E)

IEC Electropedia: available at <a href="http://www.electropedia.org/">http://www.electropedia.org/</a>

#### 3.1

# gel permeation chromatography GPC

separation of molecules, mainly based on exclusion effects such as differences in size and/or shape of molecules (size exclusion chromatography) or in charge (ion exclusion chromatography)

[SOURCE: ISO 13885-1:2020, 3.1]

#### 3.2

#### system peak

signal peculiar to the *gel permeation chromatography* (3.1) using a refractive index detector

Note 1 to entry: These signals appear at the total penetration limit of the columns and are not part of the sample, but of the overall system.

[SOURCE: ISO 13885-1:2020, 3.2]

#### 3.3

#### polycation

water-soluble polymer that carries permanent positive charges or forms temporary cationic structures due to its  $pK_B$  value under the measurement conditions

#### 3.4

#### polyanion

water-soluble polymer that carries permanent negative charges or forms temporary anionic structures due to its  $pK_S$  value under the measurement conditions

#### 3.5

#### neutral polymer

water-soluble polymer that carries no charges and forms o-charged groups or structures under the measurement conditions

#### ISO 13885-3:2020

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The dissolved (molecularly disperse) molecules of a polymer sample are fractionated on a porous column material, with separation taking place according to the size of the molecule (or, more precisely, the polymer coil size which forms in this eluent). Small molecules diffuse into the pores of the column material more frequently and are therefore retarded more than large molecules. Thus, large molecules are eluted earlier, small molecules later. Under the test conditions given, the elution volume is solely a function of the coil size of the molecule.

The polymer content of a sample is determined, the sample is then diluted with eluent to give a concentration of less than 5 g/l and an aliquot of the diluted sample is injected into the GPC system. The concentration of the molecules eluted from the column is measured in order of decreasing coil size with a concentration-sensitive detector (typically a differential refractometer). With the aid of a calibration curve that has been determined for the particular GPC system, the relative molar-mass distribution, the relative quantities  $M_{\rm n}$  and  $M_{\rm w}$  and the heterogeneity or polydispersity  $M_{\rm w}/M_{\rm n}$  are calculated from the chromatogram obtained.

#### 5 Apparatus

The apparatus shall consist of the components shown in Figure 1, which are described below.

All components which come into contact with the eluent or the sample solution shall not exhibit any adsorption and memory effects and shall not exhibit any expansion effects at the increased temperature.

The eluent described in <u>Clause 6</u> can cause corrosion in the case of long-term use. For this reason, it is necessary that high-quality steel, titanium, polyetherketones or polytetrafluoroethylene are used for

all components and that the individual modules are connected to one another by means of capillary tubes made of high-quality steel or titanium.

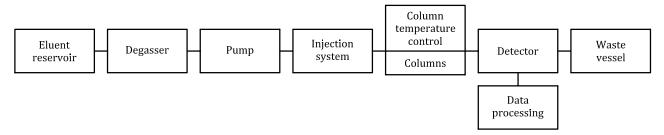


Figure 1 — Block diagram of a GPC apparatus

#### 5.1 Eluent supply

The eluent reservoir shall adequately protect the eluent against external influences such as the atmosphere, if necessary by means of a blanket of inert gas above the liquid level.

The eluent reservoir shall contain a sufficient quantity of the eluent to bring the apparatus to equilibrium and to carry out several repeat analyses.

The eluent shall be degassed, either before it is introduced into the reservoir or by use of a device fitted between the reservoir and the pump, to prevent malfunctions of the pump or the formation of bubbles in the detector. The method of degassing used (e.g. bubble trap, online purging with helium or vacuum degassing) is open to choice.

### **5.2 Pump**

The pump shall ensure that the eluent flow through the separation column is as smooth and pulse-free as possible. The flow rate shall be 1 ml/min (see Annex A). To fulfil these requirements, the pump shall operate at optimum efficiency at this flow rate.

The flow rate of the pump used shall have a variation of max. 0,1 %.

It shall be ensured that the specifications required in <u>5.6</u> for detector noise can be adhered to by means of an appropriate pump structure or a downstream pulsation damper.

#### **5.3** Injection system

The injection system serves to introduce a given amount of the sample solution into the eluent stream in a rapid and smooth fashion. This introduction can be carried out either manually or automatically.

If the introduction is carried out manually, ensure that the sample loop is filled completely with solvent before loading with the sample.

Memory effects from the previous sample solution in the injection system shall be avoided by design measures or by adequate flushing.

#### **5.4** Separation columns

The apparatus shall have one or more columns connected in series and packed with spherical porous material, the diameter of the pores corresponding to the size of the polymer molecules being analysed.

The packing material typically consists of an acrylate copolymer, produced by a special polymerization process, with OH functionality and without basic groups. The packing material shall swell only slightly in the eluent and therefore cannot deform under the pressure developed at the set flow rate.

#### ISO 13885-3:2020(E)

To keep adsorptive interactions between the packing material surface and the polymer molecules to be investigated as low as possible, silica phases and modified silica phases shall not be used. Furthermore, the sample being analysed shall not be changed, either chemically or structurally, within the chromatographic system.

Certain polymers interact with the surface of the packing material (e.g. by adsorption), and other effects can sometimes interfere with the GPC separation mechanism. Details of such effects and notes on possible remedies are discussed in  $\underline{\mathsf{Annex}\,\mathsf{C}}$ . If it is intended to compare analyses of such polymers by different laboratories, the laboratories shall agree on details of the test conditions that are not covered by this document.

For good repeatability of test results, it is necessary to adhere to the minimum requirements specified below with regards to peak broadening (expressed in terms of a number of theoretical plates) and separation efficiency.

#### a) Number of theoretical plates

The number of theoretical plates, N, shall be determined, for the apparatus used per metre of column used, from the peak width at half height (see Figure 2). Inject up to 20 µg of ethylene glycol on to each column under the same test conditions as are used for analysing polymers; the injection volume shall not exceed 20 µl (see Annex A). Evaluate the chromatogram obtained using the Formula (1):

$$N = 5.54 \times \left(\frac{V_{\rm e}}{W_{1/2}}\right)^2 \times \frac{100}{L}$$
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where

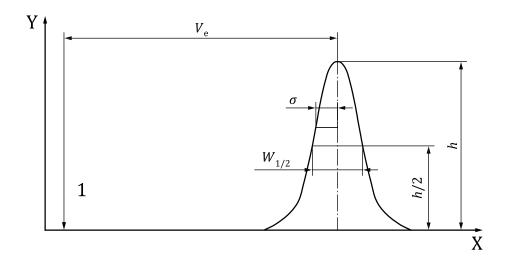
*V<sub>e</sub>* is the elution volume measured at the peak maximum;

 $W_{1/2}$  is the peak width at half height (see Figure 2); the same units shall be used for  $V_e$  and  $W_{1/2}$ ;

*L* is the length of the column (column combination), in centimetres.

Express as the result the number of theoretical plates per metre of column length. To conform to the requirements of this document, a column combination shall have at least 10 000 theoretical plates per metre.

NOTE See Annex C for tailing and fronting (asymmetry) of the peak used to calculate the plate count.



#### Key

X elution volume

Y peak intensity

1 injection

*V<sub>e</sub>* elution volume measured at the peak maximum

 $W_{1/2}$  peak width at half height of the peak

h maximum peak height

 $\sigma$  standard deviation

Figure 2 — Determination of the number of theoretical plates by the half-height method

## b) Separation efficiency **Document Preview**

To ensure adequate resolution, the  $\log_{10} M$  versus elution volume,  $V_{\rm e}$ , calibration curve for the column combination used shall not exceed a specified gradient. For the purposes of this document, the relation given in Formula (2) shall apply in the area of the peak maximum for the polymer sample under investigation:

$$\frac{V_{e,M_x} - V_{e,(10 \times M_x)}}{A_c} > 6.0$$
 (2)

where

 $V_{e,M_x}$  is the elution volume for pullulan of molar mass,  $M_x$ , in cubic centimetres;

 $V_{\rm e,(10\times M_{\rm v})}$  is the elution volume for 10 times that molar mass, in cubic centimetres;

 $A_{\mathrm{c}}$  is the cross-sectional area of the column, in square centimetres;

 $M_{\scriptscriptstyle \rm X}$  is the molar mass of pullulan; it shall be selected such that the peak maximum for the polymer sample under investigation lies approximately halfway between these two elution volumes.

#### 5.5 Column temperature control

Carry out the test at room temperature (15 °C to 35 °C) or at a higher temperature of max. 40 °C. The temperature of the column shall not change by more than 1 °C during the analysis (see Annex C).