# DRAFT INTERNATIONAL STANDARD ISO/DIS 7358

ISO/TC 54

Voting begins on: **2020-04-01** 

Secretariat: UNE

Voting terminates on: 2020-06-24

## Essential oils of bergamot, lemon, bitter orange and lime, fully or partially reduced in bergapten — Determination of bergapten content by high-performance liquid chromatography (HPLC)

Huiles essentielles de bergamote, de citron, de bigarade et de limette complètement ou partiellement privées de bergaptène — Détermination de la teneur en bergaptène par chromatographie liquide à haute performance(CLHP)

ICS: 71.100.60

## iTeh STANDARD PREVIEW (standards.iteh.ai)

ISO/DIS 7358 https://standards.iteh.ai/catalog/standards/sist/8cd98e3c-7419-4e59-ab1cfb622eae8fcc/iso-dis-7358

THIS DOCUMENT IS A DRAFT CIRCULATED FOR COMMENT AND APPROVAL. IT IS THEREFORE SUBJECT TO CHANGE AND MAY NOT BE REFERRED TO AS AN INTERNATIONAL STANDARD UNTIL PUBLISHED AS SUCH.

IN ADDITION TO THEIR EVALUATION AS BEING ACCEPTABLE FOR INDUSTRIAL, TECHNOLOGICAL, COMMERCIAL AND USER PURPOSES, DRAFT INTERNATIONAL STANDARDS MAY ON OCCASION HAVE TO BE CONSIDERED IN THE LIGHT OF THEIR POTENTIAL TO BECOME STANDARDS TO WHICH REFERENCE MAY BE MADE IN NATIONAL REGULATIONS.

RECIPIENTS OF THIS DRAFT ARE INVITED TO SUBMIT, WITH THEIR COMMENTS, NOTIFICATION OF ANY RELEVANT PATENT RIGHTS OF WHICH THEY ARE AWARE AND TO PROVIDE SUPPORTING DOCUMENTATION. This document is circulated as received from the committee secretariat.



Reference number ISO/DIS 7358:2020(E)

## iTeh STANDARD PREVIEW (standards.iteh.ai)

ISO/DIS 7358 https://standards.iteh.ai/catalog/standards/sist/8cd98e3c-7419-4e59-ab1cfb622eae8fcc/iso-dis-7358



#### **COPYRIGHT PROTECTED DOCUMENT**

#### © ISO 2020

All rights reserved. Unless otherwise specified, or required in the context of its implementation, no part of this publication may be reproduced or utilized otherwise in any form or by any means, electronic or mechanical, including photocopying, or posting on the internet or an intranet, without prior written permission. Permission can be requested from either ISO at the address below or ISO's member body in the country of the requester.

ISO copyright office CP 401 • Ch. de Blandonnet 8 CH-1214 Vernier, Geneva Phone: +41 22 749 01 11 Fax: +41 22 749 09 47 Email: copyright@iso.org Website: www.iso.org

Published in Switzerland

Page

## Contents

Forew	vord	iv
1	Scope	1
2	Normative references	1
3	Principle	1
4	Reagents	1
5	Equipment and apparatus5.1Common laboratory equipment5.2HPLC system, consisting of the following	2 2 2
6	Sample preparation	3
7	Procedure   7.1 Operating conditions   7.2 Determination   7.2.1 HPLC in normal phase   7.2.2 HPLC in reversed phase	<b>3</b> 3 3 5
8	Calculation   8.1 Normal phase or reversed phase HPLC   8.1.1 Internal standard method   8.1.2 External standard method	7 7 7 8
9	Precision 9.1 Repeatability (standards.iteh.ai)	<b>8</b> 8
10	Test report	8
Annex	A (informative) Typical chromatogram of the reference substances using high- performance liquid chromatography of reference compounds in reversed phase	.9

## Foreword

ISO (the International Organization for Standardization) is a worldwide federation of national standards bodies (ISO member bodies). The work of preparing International Standards is normally carried out through ISO technical committees. Each member body interested in a subject for which a technical committee has been established has the right to be represented on that committee. International organizations, governmental and non-governmental, in liaison with ISO, also take part in the work. ISO collaborates closely with the International Electrotechnical Commission (IEC) on all matters of electrotechnical standardization.

The procedures used to develop this document and those intended for its further maintenance are described in the ISO/IEC Directives, Part 1. In particular, the different approval criteria needed for the different types of ISO documents should be noted. This document was drafted in accordance with the editorial rules of the ISO/IEC Directives, Part 2 (see <a href="https://www.iso.org/directives">www.iso.org/directives</a>).

Attention is drawn to the possibility that some of the elements of this document may be the subject of patent rights. ISO shall not be held responsible for identifying any or all such patent rights. Details of any patent rights identified during the development of the document will be in the Introduction and/or on the ISO list of patent declarations received (see <a href="https://www.iso.org/patents">www.iso.org/patents</a>).

Any trade name used in this document is information given for the convenience of users and does not constitute an endorsement.

For an explanation of the voluntary nature of standards, the meaning of ISO specific terms and expressions related to conformity assessment, as well as information about ISO's adherence to the World Trade Organization (WTO) principles in the Technical Barriers to Trade (TBT), see <a href="https://www.iso.org/iso/foreword.html">www.iso.org/iso/foreword.html</a>.

This document was prepared by Technical Committee ISO/TC 54, Essential oils.

This second edition cancels and replaces the first/edition/(ISO 7358:2002)) which has been technically revised with the addition of an alternative method using a reversed phase column.

Any feedback or questions on this document should be directed to the user's national standards body. A complete listing of these bodies can be found at <u>www.iso.org/members.html</u>.

## Essential oils of bergamot, lemon, bitter orange and lime, fully or partially reduced in bergapten — Determination of bergapten content by high-performance liquid chromatography (HPLC)

#### 1 Scope

This International Standard specifies a high-performance liquid chromatographic (HPLC) method, using either an internal standard or external standard, for the determination of the bergapten content in essential oil of bergamot [*Citrus aurantium* ssp. bergamia (Risso et Poit.) Wight et Arn. ex Engl.], in essential oil of lemon [*Citrus limon* (L.) Burm. f.], in essential oil of bitter orange (*Citrus bigaradia* Risso), and in essential oil of lime [*Citrus aurantifolia* (Christm.) Swingle and *Citrus latifolia* Tanaka], all of them fully or partially reduced in bergapten.

#### 2 Normative references

The following documents are referred to in the text in such a way that some or all of their content constitutes requirements of this document. For dated references, only the edition cited applies. For undated references, the latest edition of the referenced document (including any amendments) applies.

ISO 356, Essential oils — Preparation of test 3 diples iteh.ai)

ISO 8432:1987, Essential oils — Analysis by high performance liquid chromatography — General method

https://standards.iteh.ai/catalog/standards/sist/8cd98e3c-7419-4e59-ab1cfb622eae8fcc/iso-dis-7358

#### **3** Principle

The bergapten contents of test samples to be measured are determined after dilution by reversed phase high performance liquid chromatography or by normal phase high performance liquid chromatography with gradient elution, using an internal standardization method or an external standardization method and diode-array UV spectrometric detection.

#### 4 Reagents

Use only reagents of recognized analytical grade.

**4.1** Reference substance: bergapten (5-Methoxypsoralen),  $C_{12}H_8O_4$ , MW = 216,19 g/mol of known purity  $\ge 95$  %.

**4.2 Internal standard: coumarin** (1-Benzopyran-2-one),  $C_9H_6O_2$ , MW = 146,14 g/mol of known purity  $\ge 98$  %.

#### 4.3 Solvents

#### 4.3.1 Solvents for normal phase high performance liquid chromatography

**4.3.1.1 Chloroform**, of analytical purity, containing less than 2 % (volume fraction) of ethanol, for use in preparing the sample of essential oil containing the bergapten and internal standard as well as the mobile phase.

#### ISO/DIS 7358:2020(E)

**4.3.1.2** Hexane, HPLC grade, for use in the mobile phase (<u>4.3.1.4</u>).

**4.3.1.3 Ethyl acetate**, HPLC grade, for use in the mobile phase (<u>4.3.1.4</u>).

#### 4.3.1.4 Mobile phase

For the mobile phase, use solvents of a quality compatible with the detection system, and prepare sufficient quantities for the complete analysis. Mix, for example, one of the following:

**4.3.1.4.1** Hexane (<u>4.3.1.2</u>) and ethyl acetate (<u>4.3.1.3</u>), mixed in proportions of 80:20 by volume.

**4.3.1.4.2** Hexane (<u>4.3.1.2</u>) and chloroform (<u>4.3.1.1</u>), mixed in proportions of 85:15 by volume.

#### 4.3.2 Solvents for reversed phase high performance liquid chromatography

The solvent used for the mobile phase is a ternary mobile phase prepared with these three solvents as described in the <u>table 1</u> (see <u>7.2.2.3.1</u>).

**4.3.2.1** HPLC grade **distilled water**, to be used in mobile phase (7.2.2.3.1).

**4.3.2.2** HLPC grade **acetonitrile**, to be used in the mobile phase (7.2.2.3.1).

**4.3.2.3** HLPC grade **methanol**, to be used in the mobile phase (72.2.3.1).

**4.3.2.4** Dilution solvent, mixture of **methanol** (<u>4.3.2.3</u>) and **acetonitrile** (<u>4.3.2.2</u>) in proportions of 80:20 by volume.

5 Equipment and apparatus 5 Equipment and apparatus

#### 5.1 Common laboratory equipment

- 5.1.1 Volumetric flasks: 100 ml, 50 ml, 10 ml
- 5.1.2 Pasteur pipettes; volumetric pipettes: 1 ml, 5 ml, 10 ml and 20 ml; micropipette: 500 µl
- 5.1.3 Single-use syringes: 5 ml
- 5.1.4 PTFE filter: 0,45 μm
- 5.1.5 Vials for HPLC injection
- 5.1.6 Precision balance
- 5.2 HPLC system, consisting of the following
- 5.2.1 Liquid chromatograph

**5.2.2** Column for high performance liquid chromatography in normal phase, made of stainless steel, of length between 150 mm and 250 mm, having an internal diameter between 4 mm and 5 mm and packed with a stationary phase consisting of granulated silica of HPLC quality, with a grain size of approximately 5  $\mu$ m.

**5.2.3 Column for high performance liquid chromatography in reversed phase**, made of stainless steel, of length 150 mm (or 250 mm), having an internal diameter 4, 6 mm and packed with a stationary phase C18 type with a grain size of approximately 3,5  $\mu$ m (or 5  $\mu$ m) for example Zorbax Eclipse Plus C:18: 3,5  $\mu$ m (4,6 × 150) mm agilent.

#### 5.2.4 A ternary pump system enabling programmed solvent gradients

**5.2.5** Solvent degassing system (optional), for example ultrasonic tank.

**5.2.6 Detection system**, adjustable to wavelengths of 254 nm or 312 nm, diode-array UV spectrometric detection system.

**5.2.7 Recorder** and (optional) **integrator**, suitable for this HPLC system.

#### 6 Sample preparation

Prepare the test sample as specified in ISO 356.

Dissolve any solid deposit by moderate heating.

#### 7 Procedure

## 7.1 Operating conditions **STANDARD PREVIEW**

Adjust the flow rate of the mobile phase (4.3.1.4) so as to obtain good separation of the peaks

Adjust the flow rate of the mobile phase (4.3.1.4) so as to obtain good separation of the peaks corresponding to bergapten and coumarin from other essential oil components detectable by the UV detector (5.2.6). The flow rate is typically between 1 ml/min and 1,5 ml/mn.

Follow the procedure specified in ISO<sup>18432</sup>ae8fcc/iso-dis-7358

#### 7.2 Determination

7.2.1 HPLC in normal phase

#### 7.2.1.1 Internal standard method

#### 7.2.1.1.1 Optimization of HPLC chromatographic conditions in normal phase

#### 7.2.1.1.1.1 Separation

Verify that the bergapten is well separated from the other constituents of the essential oil in the chromatograms obtained. Secondly verify that the internal standard, coumarin (4.2), does not mask or coincide with any constituent of the essential oil. Determine the retention times of the bergapten and coumarin.

#### 7.2.1.1.1.2 Quantity of internal standard

The amount of coumarin (internal standard) added to the sample is considered suitable when the peak areas for the bergapten (in the essential oil) and the coumarin are approximately equal in the chromatograms. To determine this amount, inject (e.g. 10  $\mu$ l) a solution containing a given amount (e.g. 10 mg) of coumarin (4.2) dissolved in chloroform (4.3.1.1 e.g. 10 ml) into the HPLC. Then inject the same volume of a solution of the essential oil analyte diluted in chloroform (4.3.1.1). Adjust the mass concentrations of both of these solutions so as to obtain comparable peak areas.

#### 7.2.1.1.2 Response factor *K*

Prepare a calibration solution as follows. In a volumetric flask (5.1.1) of suitable volume, weigh, to the nearest 0,1 mg, about 20 mg of coumarin (4.2). In the same volumetric flask, weigh, to the nearest 0,1 mg, about 10 mg of bergapten (4.1) and dissolve both compounds in approximately 20 ml of chloroform (4.3.1.1).

Inject a suitable amount (see <u>7.2.1.1</u>) of the calibration solution so as to remain within the detector sensitivity range.

Measure the peak areas of the chromatogram. A typical chromatogram of the reference substances is given in <u>annex A</u>.

Calculate the response factor *K* using <u>equation (1)</u>:

$$K = \frac{m_{\rm R} \cdot A_{\rm IS}}{m_{\rm IS} \cdot A_{\rm R}} \tag{1}$$

where

- $m_{\rm R}$  is the mass, expressed in milligrams, of bergapten (reference substance) (4.1) added to the solution;
- $m_{\rm IS}$  is the mass, expressed in milligrams, of coumarin (internal standard) (4.2) added to the solution;
- $A_{\rm R}$  is the peak area, expressed in integrator units, corresponding to bergapten (reference substance) (4.1);
- A<sub>IS</sub> is the peak area, expressed in integrator units, corresponding to coumarin (internal standard) (4.2). ISO/DIS 7358

NOTE In ISO 8432 this equation is equivalent to the following: b622eae8fcc/iso-dis-7358

$$K = \frac{m_{\rm R} A_{\rm E}}{m_{\rm E} A_{\rm R}}$$

where

where

- $m_{\rm R}$  is the mass, expressed in milligrams, of bergapten (reference substance) (4.1) added to the solution;
- $m_{\rm E}$  is the mass, expressed in milligrams, of coumarin (internal standard) (4.2) added to the solution;
- $A_{\rm R}$  is the peak area, expressed in integrator units, corresponding to bergapten (reference substance) (4.1);
- $A_{\rm E}$  is the peak area, expressed in integrator units, corresponding to coumarin (internal standard) (4.2).

#### 7.2.1.1.3 Determination of bergapten

In a volumetric flask (5.1.1) of suitable volume (e.g. 15 ml) prepare the test solution. Weigh, to the nearest 0,1 mg, a suitable amount of coumarin ( $m_{\rm IS}$ ) (approximately 10 mg), as determined in 7.2.2.1, and a portion of the essential oil ( $m_{\rm s}$ ) so as to obtain a chromatogram with equal peak areas for bergapten and coumarin.

Add chloroform (<u>4.3.1.1</u>, approximately 8 ml) and shake carefully to dissolve the coumarin.

It may be necessary to prepare several dilutions of the test solution to obtain a chromatogram with comparable peak areas for bergapten and coumarin because the bergapten content in the essential oils is unknown (fully or partially reduced in bergapten). Choose the volume of the volumetric flask, the quantities of coumarin and the volume of chloroform so as to meet this requirement.

Using the same HPLC operating conditions established in <u>7.2.1.1.1</u>, inject a suitable amount of the test solution so as to remain within the detector sensitivity range.

Measure the peak areas of the chromatogram.

Measure and record the peak areas of the chromatogram corresponding to bergapten ( $A_x$ ) and coumarin ( $A_{IS}$ ).

#### 7.2.1.2 External standard method

Follow the procedure for the external standard method as specified in ISO 8432.

#### 7.2.2 HPLC in reversed phase

#### 7.2.2.1 Preparation of the reference solutions for calibration:

For each calibration, the standard solutions are to be prepared using stock solutions of bergapten and coumarin, to be stored in a cool place at 4 °C.

### 7.2.2.1.1 Stock solutionsh STANDARD PREVIEW

As accurately as possible weigh **50 mg of bergapten into a** 100 ml flask and make up to volume with the dilution solvent. Homogenize the solution and leave the solution for 3 min in the ultrasonic bath. This stock solution contains almost exactly 500 mg/l of bergapten.

As accurately as possible weigh 50 mg of counterin into a 50 ml flask and make up to volume with the elution solvent. Homogenize the solution and leave the solution for 3 min in the ultrasonic bath. This solution contains almost exactly 1 000 mg/l of counterin.

#### 7.2.2.1.2 Working solutions

Using the stock solutions, prepare a standard range of 8 calibration points for bergapten concentration: 1 mg/l, 2,5 mg/l, 5 mg/l, 10 mg/l, 25 mg/l, 50 mg/l, 100 mg/l and 200 mg/l.

- 1 mg/l standard solution: into a 50 ml flask, transfer 100 μl of bergapten stock solution and 5 ml of coumarin stock solution then make up to volume with the dilution solvent.
- 2,5 mg/l standard solution: into a 50 ml flask, transfer 250 μl of bergapten stock solution and 5 ml of coumarin stock solution then make up to volume with the dilution solvent.
- 5 mg/l standard solution: into a 50 ml flask, transfer 500 μl of bergapten stock solution and 5 ml of coumarin stock solution then make up to volume with the dilution solvent.
- 10 mg/l standard solution: into a 50 ml flask, transfer 1 ml of bergapten stock solution and 5 ml of coumarin stock solution then make up to volume with the dilution solvent.
- 25 mg/l standard solution: into a 50 ml flask, transfer 2,5 ml of bergapten stock solution and 5 ml of coumarin stock solution then make up to volume with the dilution solvent.
- 50 mg/l standard solution: into a 50 ml flask, transfer 5 ml of bergapten stock solution and 5 ml of coumarin stock solution then make up to volume with the dilution solvent.
- 100 mg/l standard solution: into a 50 ml flask, transfer 10 ml of bergapten stock solution and 5 ml of coumarin stock solution then make up to volume with the dilution solvent.