



Designation: D7284 – 08^{ε1}

Standard Test Method for Total Cyanide in Water by Micro Distillation followed by Flow Injection Analysis with Gas Diffusion Separation and Amperometric Detection¹

This standard is issued under the fixed designation D7284; the number immediately following the designation indicates the year of original adoption or, in the case of revision, the year of last revision. A number in parentheses indicates the year of last reapproval. A superscript epsilon (ϵ) indicates an editorial change since the last revision or reapproval.

^{ε1} NOTE—This Test Method was editorially corrected in September 2012.

1. Scope

1.1 This test method is used to determine the concentration of total cyanide in an aqueous wastewater or effluent. The method detects the cyanides that are free (HCN and CN⁻) and strong-metal-cyanide complexes that dissociate and release free cyanide when refluxed under strongly acidic conditions.

1.2 This method may not be applicable to process solutions from precious metals mining operations.

1.3 This procedure is applicable over a range of approximately 2 to 400 µg/L (parts per billion) total cyanide. Higher concentrations can be measured with sample dilution or lower injection volume.

1.4 The determinative step of this method utilizes flow injection with amperometric detection based on Test Method D6888. Prior to analysis, samples must be distilled with a micro-distillation apparatus described in this test method or with a suitable cyanide distillation apparatus specified in Test Methods D2036.

1.5 *This standard does not purport to address all of the safety concerns, if any, associated with its use. It is the responsibility of the user of this standard to establish appropriate safety and health practices and determine the applicability of regulatory limitations prior to use.* Specific hazard statements are given in Note 2 and Section 9.

2. Referenced Documents

2.1 *ASTM Standards:*²

D1129 Terminology Relating to Water

D1193 Specification for Reagent Water

¹ This test method is under the jurisdiction of ASTM Committee D19 on Water and is the direct responsibility of Subcommittee D19.06 on Methods for Analysis for Organic Substances in Water.

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² For referenced ASTM standards, visit the ASTM website, www.astm.org, or contact ASTM Customer Service at service@astm.org. For *Annual Book of ASTM Standards* volume information, refer to the standard's Document Summary page on the ASTM website.

D2036 Test Methods for Cyanides in Water

D2777 Practice for Determination of Precision and Bias of Applicable Test Methods of Committee D19 on Water

D3856 Guide for Management Systems in Laboratories Engaged in Analysis of Water

D5847 Practice for Writing Quality Control Specifications for Standard Test Methods for Water Analysis

D6696 Guide for Understanding Cyanide Species

D6888 Test Method for Available Cyanide with Ligand Displacement and Flow Injection Analysis (FIA) Utilizing Gas Diffusion Separation and Amperometric Detection

D7365 Practice for Sampling, Preservation and Mitigating Interferences in Water Samples for Analysis of Cyanide

3. Terminology

3.1 *Definitions:*

3.1.1 For definitions of terms used in this test method, refer to Terminology D1129 and Guide D6696.

3.1.2 *total cyanide*—Total cyanide is an analytically defined term that refers to the sum total of all of the inorganic chemical forms of cyanide that dissociate and release free cyanide when refluxed under strongly acidic conditions. Total cyanide is determined analytically through strong acid distillation or UV radiation followed by analysis of liberated free cyanide on aqueous samples preserved with NaOH (pH~12). In water, total cyanide includes the following dissolved species: free cyanide, weak acid dissociable metal cyanide complexes and strong metal cyanide complexes. Also, some of the strong metal cyanide complexes, such as those of gold, cobalt and platinum, might not be fully recovered during the total cyanide analytical procedure. Additionally, total cyanide may also include some organic forms of cyanide such as nitriles that release free cyanide under the conditions of the analysis.

4. Summary of Test Method

4.1 The samples are distilled with a strong acid in the presence of magnesium chloride catalyst and captured in sodium hydroxide absorber solution.

4.2 The absorber solution is introduced into a flow injection analysis (FIA) system where it is acidified to form hydrogen cyanide (HCN). The hydrogen cyanide gas diffuses through a hydrophobic gas diffusion membrane, from the acidic donor stream into an alkaline acceptor stream.

4.3 The captured cyanide is sent to an amperometric flow-cell detector with a silver-working electrode. In the presence of cyanide, silver in the working electrode is oxidized at the applied potential. The anodic current measured is proportional to the concentration of cyanide.

4.4 Calibrations and data are processed with the instrument's data acquisition software.

5. Significance and Use

5.1 Cyanide and hydrogen cyanide are highly toxic. Regulations have been established to require the monitoring of cyanide in industrial and domestic wastes and surface waters.³

5.2 This test method is applicable for natural waters, industrial wastewaters and effluents.

6. Interferences

6.1 Improper sample collection or pretreatment can result in significant positive or negative bias, therefore it is imperative that samples be collected and mitigated for interferences as described in Practice D7365.

6.1.1 Sulfide captured in the absorber solution above 50-mg/L S^{2-} will diffuse through the gas diffusion membrane during flow injection analysis and can be detected in the amperometric flowcell as a positive response. Refer to Section 11.2 for sulfide abatement.

6.1.2 Thiocyanate in the presence of oxidants (for example, nitrates, hydrogen peroxide, chlorine or chloramine, Caro's acid), can decompose to form cyanide during the distillation

resulting in positive interference regardless of the determinative step (amperometry, colorimetry, etc.). During acidic distillation, decomposition of thiocyanate in the absence of oxidants produces elemental sulfur, sulfur(IV) oxide, as well as carbonyl sulfide which eventually leads to the formation of sulfite ion (SO_3^{2-}) in the NaOH absorbing solution. The sulfite ion slowly oxidizes cyanide to cyanate resulting in a negative interference. Therefore, samples that are known to contain significant amounts of thiocyanate may need to be analyzed with a method that does not require distillation, for example, available cyanide by Test Method D6888.

6.1.2.1 During the validation study, synthetic samples containing up to 15 mg/L SCN^- and 25 mg/L NO_3^- as N yielded less than 0.5 % of the SCN^- to be measurable CN^- . For example, a solution that did not contain any known amount of cyanide, but did contain 15-mg/L SCN^- and 25 mg/L NO_3^- as N, was measured as 53.1 $\mu\text{g/L } CN^-$.

7. Apparatus and Instrumentation

7.1 The instrument should be equipped with a precise sample introduction system, a gas diffusion manifold with hydrophobic membrane, and an amperometric detection system to include a silver working electrode, a Ag/AgCl reference electrode, and a Pt or stainless steel counter electrode. The apparatus schematic is shown in Fig. 1, and example instrument settings are shown in Table 1.⁴

NOTE 1—The instrument settings in Table 1 are only examples. The analyst may modify the settings as long as performance of the method has not been degraded. Contact the instrument manufacturer for recommended instrument parameters.

7.1.1 An autosampler is recommended but not required to automate sample injections and increase throughput. Autosamplers are usually available as an option from the instrument's manufacturer.

⁴ Both the OI Analytical CN Solution and Lachat Instruments QuikChem Automated Ion Analyzer have been found to be suitable for this analysis.

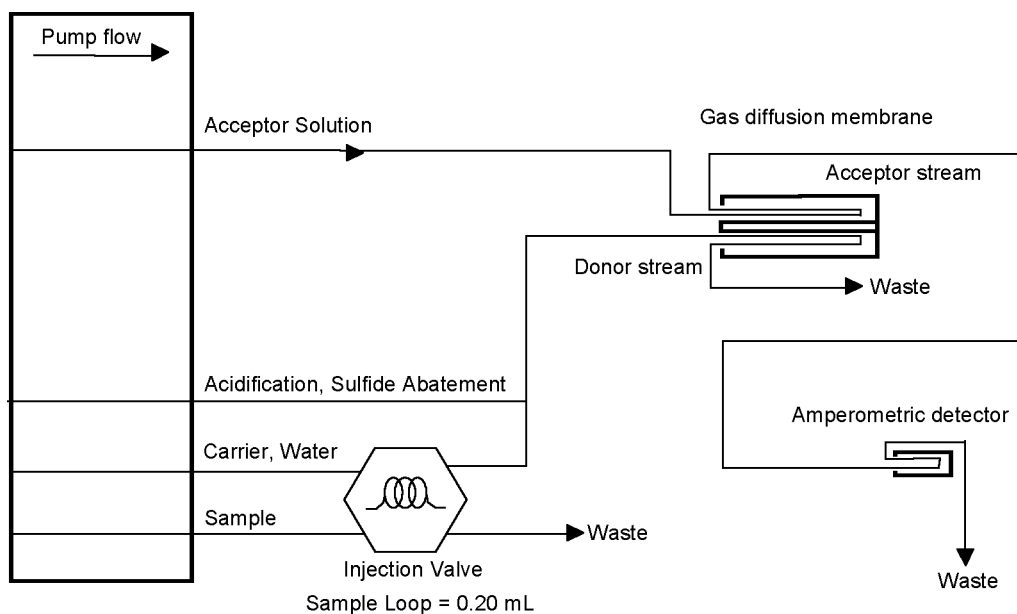


FIG. 1 Flow Injection Analysis Apparatus

³ 40 CFR Part 136.

TABLE 1 Flow Injection Analysis Parameters

FIA Instrument Parameter	Recommended Method Setting
Pump Flow Rates	0.5 to 2 mL/min
Cycle period (total)	Approximately 120 seconds
Sample load period	At least enough time to completely fill the sample loop prior to injection
Injection valve rinse time between samples	At least enough time to rinse the sample loop
Peak Evaluation Working Potential	Peak height or area 0.0 V vs Ag/AgCl

7.1.2 *Data Acquisition System*—Use the computer hardware and software recommended by the instrument manufacturer to control the apparatus and to collect data from the detector.

7.1.3 *Pump Tubing*—Use tubing recommended by instrument manufacturer. Replace pump tubing when worn, or when precision is no longer acceptable.

7.1.4 *Gas Diffusion Membranes*—A hydrophobic membrane which allows gaseous hydrogen cyanide to diffuse from the donor to the acceptor stream at a sufficient rate to allow detection. The gas diffusion membrane should be replaced when the baseline becomes noisy or every 1 to 2 weeks.⁵

7.1.5 Use parts and accessories as directed by instrument manufacturer.

7.2 *Distillation Apparatus*—The MicroDist System described below was utilized during the laboratory study to demonstrate precision and bias for this test method. A larger distillation apparatus such as the MIDI distillation described in section 7 of Test Methods D2036 can also be used to prepare

⁵ The sole source of supply of the apparatus known to the committee at this time is PALL Life Sciences Part Number M5PU025, OI Analytical Part Number A0015200, and Lachat Instruments Part Number 50398. If you are aware of alternative suppliers, please provide this information to ASTM International Headquarters. Your comments will receive careful consideration at a meeting of the responsible technical committee,¹ which you may attend.

samples prior to flow injection analysis, but the user is responsible to determine the precision and bias.

7.2.1 *Micro-Distillation Apparatus* consisting of a distillation sample tube, stop ring, membrane, and collection vessel containing 1.0 M sodium hydroxide and a breakaway collection tube as shown in Fig. 2.⁶

7.2.2 *Heater block assembly*, temperature controlled, capable of heating the MicroDist tubes to 120°C.

8. Reagents and Materials

8.1 *Purity of Reagents*—Reagent grade chemicals shall be used in all tests. Unless otherwise indicated, it is intended that all reagents shall conform to the specifications of the American Chemical Society, where such specifications are available.⁷ Other grades may be used, provided it is first ascertained that the reagent is of sufficiently high purity to permit its use without lessening the accuracy of the determination.

8.2 *Purity of Water*—Unless otherwise indicated, references to water shall be understood to mean reagent water that meets the purity specifications of Type I or Type II water, presented in Specification D1193.

8.3 *Sodium Hydroxide Solution (1.00 M)*—Dissolve 40 g NaOH in laboratory water and dilute to 1 L.

8.4 *Absorber Solution for MIDI Distillations (0.25 M NaOH)*—Dissolve 10 g NaOH in laboratory water and dilute to 1 L.

8.5 *Acceptor Solution (0.10 M NaOH)* —Dissolve 4.0 g NaOH in laboratory water and dilute to 1 L.

8.6 *Stock Cyanide Solution (1000 µg/mL CN⁻)*—Dissolve 2.51 g of KCN and 2.0 g of NaOH in 1 L of water. Standardize

⁶ The sole source of supply of the apparatus known to the committee at this time is Lachat Instruments, PN A17001 (subject to US Reg. Patent No. 5,022,967). If you are aware of alternative suppliers, please provide this information to ASTM International Headquarters. Your comments will receive careful consideration at a meeting of the responsible technical committee,¹ which you may attend.

⁷ *Reagent Chemicals, American Chemical Society Specifications*, American Chemical Society, Washington, DC. For suggestions on the testing of reagents not listed by the American Chemical Society, see *Analar Standards for Laboratory Chemicals*, BDH Ltd., Poole, Dorset, U.K., and the *United States Pharmacopeia and National Formulary*, U.S. Pharmacopeial Convention, Inc. (USPC), Rockville, MD.

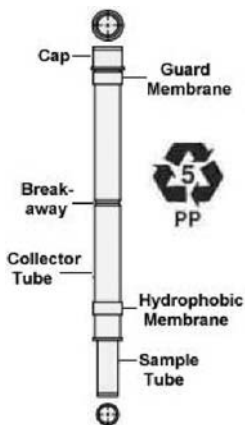


FIG. 2 MicroDist Sample Tube