



Designation: **F720–81 (Reapproved 2012) F720 – 13**

Standard Practice for Testing Guinea Pigs for Contact Allergens: Guinea Pig Maximization Test¹

This standard is issued under the fixed designation F720; the number immediately following the designation indicates the year of original adoption or, in the case of revision, the year of last revision. A number in parentheses indicates the year of last reapproval. A superscript epsilon (ϵ) indicates an editorial change since the last revision or reapproval.

1. Scope

- 1.1 This practice is intended to determine the potential for a substance, or material extract, to elicit contact dermal allergenicity.
- 1.2 The values stated in SI units are to be regarded as standard. No other units of measurement are included in this standard.
- 1.3 *This standard does not purport to address all of the safety concerns, if any, associated with its use. It is the responsibility of the user of this standard to establish appropriate safety and health practices and determine the applicability of regulatory limitations prior to use.*

2. Referenced Documents

2.1 *ASTM Standards:*²

[F619 Practice for Extraction of Medical Plastics](#)

2.2 *ISO Standards:*²

[ISO 10993–10 Biological evaluation of medical devices—Part 10: Tests for irritation and skin sensitization](#)

3. Summary of Practice

3.1 After a two-stage induction employing Freund's complete adjuvant and sodium lauryl sulfate, the substance or extract is placed on patches and then placed on the skin of guinea pigs. After 24 h, the patches are removed and the skin examined for allergic reaction, and the intensity of the reaction scored at the time of removal and 24 and 48 h subsequent to removal.

4. Significance and Use

4.1 In selecting a new material for human contact in medical applications, it is important to ensure that the material will not stimulate the immune system to produce an allergic reaction. The reaction would be due to substances which could leach out of a material. Therefore, this practice provides for using material extracts. The rationale for this practice is based on the fact that the guinea pig has been shown to be the best animal model for human allergic contact dermatitis. The use of Freund's complete adjuvant and sodium lauryl sulfate tends to enhance the potential of a material to cause an allergy. Therefore, this test, while not guaranteeing that a material is nonallergenic, is the most severe animal test in common use today.

5. Materials and Manufacturer

5.1 *Hartley Strain Guinea Pigs*, male, 300 to 500 g.

5.1.1 Ten animals are used for each test material.

5.2 *Freund's Complete Adjuvant*.

5.3 *Occlusive Surgical Tape*, 3.75 cm in width.

5.4 *Elastic Bandage*.

5.5 *Sodium Lauryl Sulfate* (10 weight %) in USP petroleum jelly.

5.6 *Positive Control Substance*.

¹ This practice is under the jurisdiction of ASTM Committee F04 on Medical and Surgical Materials and Devices and is the direct responsibility of Subcommittee F04.16 on Biocompatibility Test Methods.

Current edition approved Oct. 1, 2012 June 1, 2013. Published October 2012 July 2013. Originally approved in 1981. Last previous edition approved in 2007 2012 as F720 – 81 (2007)(2012).¹; DOI: 10.1520/F0720-81R12; 10.1520/F0720-13.

² For referenced ASTM standards, visit the ASTM website, www.astm.org, or contact ASTM Customer Service at service@astm.org. For *Annual Book of ASTM Standards* volume information, refer to the standard's Document Summary page on the ASTM website.³

³ Available from International Organization for Standardization (ISO), 1, ch. de la Voie-Creuse, CP 56, CH-1211 Geneva 20, Switzerland, <http://www.iso.org>.

5.6.1 5 % formaldehyde for water-soluble test substances.

6. Preparation of Test Samples

6.1 Samples for Intradermal Injection:

6.1.1 Water-Soluble Constituents or Water Extract Liquids:

6.1.1.1 Dissolve the water-soluble constituent up to its maximum solubility, not to exceed a concentration of 10 weight %, or obtain a water extraction liquid as described in Practice F619.

6.1.1.2 Combine equal volumes of the liquid described in 6.1.1.1 and Freund's complete adjuvant. Homogenize by continuous and vigorous vortex mixing for a minimum of 5 min. Emulsification is complete when a drop placed on the surface of a water-ice bath remains intact.

6.1.1.3 Also prepare the constituent or extract to the same concentration in water without Freund's complete adjuvant.

6.1.2 Oil Soluble Constituents:

6.1.2.1 Dissolve oil-soluble constituents in Freund's complete adjuvant to a concentration of 10 weight %.

6.1.2.2 Combine equal volumes of the 10 % Freund's adjuvant solution with an equal volume of water by slowly adding the water to the adjuvant while homogenizing with a rotating stirrer. Homogenize by continuous and vigorous mixing for a minimum of 5 min. Emulsification is complete when a drop placed on the surface of a water-ice bath remains intact.

6.1.2.3 Also prepare the constituent to an equal concentration without Freund's complete adjuvant.

6.1.3 Vegetable Oil Extract Liquids:

6.1.3.1 Mix equal volumes of oil extract liquid obtained in accordance with Practice F619 with an equal volume of Freund's complete adjuvant.

6.1.3.2 Also prepare extract to an equal concentration in water without Freund's complete adjuvant.

6.1.4 Alternative Sample Preparation: As an alternative method for sample extraction, the method described in ISO 10993–10, Annex E is recommended for consideration for the extraction of polymeric materials.

6.1.5 Prepare control substances consistent with 6.1.1-6.1.3.2.

NOTE 1—If the final concentration of the suspected allergen results in deleterious effects such as ulceration, necrosis, or systemic toxicity, use the maximum tolerable concentration.

6.2 Samples for Topical Application:

6.2.1 *Liquids*—For all liquids, use highest concentration not causing excessive irritation or deleterious to general health of test animals. (See Note 1.)

6.2.1.1 For oil-miscible liquids, dilute with petroleum jelly if necessary.

6.2.1.2 For water-miscible liquids, dilute with water if necessary.

7. Procedure for Test and Control Substances

7.1 Induction:

7.1.1 Intradermal Injection:

7.1.1.1 Clip the shoulder region of each guinea pig free of hair, exposing a 4 by 6-cm area.

7.1.1.2 Three injection sites on each side of the spine should be identified with at least 1.5 cm between sites. Intradermal injections are then made as indicated below, with Site 1 being closest to the animal's head.

(1) Site 1—0.1 mL of adjuvant without test sample

(2) Site 2—0.1 mL of test sample without adjuvant

(3) Site 3—0.1 mL of test sample emulsified in complete adjuvant

7.1.2 Topical Application:

7.1.2.1 One week after the intradermal injections, clip the test area of the guinea pigs.

7.1.2.2 If the test sample is non-irritating, treat each test area with 10 % sodium lauryl sulfate (SLS) in petroleum jelly, 24 h before applying the test patches. Massage the SLS into the skin with a glass rod.

7.1.2.3 Apply the test sample to a 2 by 4-cm patch of qualitative filter paper. Apply the test agent in petroleum jelly to the filter in a thick, even layer. Apply the test liquid to the test paper until saturated.

7.1.2.4 Apply filter paper to the injection site of the guinea pigs. Cover with 3.75-cm occlusive surgical tape and wrap an elastic bandage around the torso to secure the tape.

7.1.2.5 Leave in place for 48 h.

7.2 Challenge:

7.2.1 Two weeks following the induction described in 7.1, perform the following:

7.2.2 Shave a 5 by 5-cm area on the animal's flank.

7.2.3 Delivery of Test Sample:

7.2.3.1 Test solids as a 25 % concentration (providing this concentration is not irritating) in petroleum jelly by weight. Apply on a 2 by 2-cm piece of filter paper as outlined in 7.1.2.3.

7.2.3.2 Test liquids in undiluted form consistent with 6.1.5. Apply on a 2 by 2-cm piece of filter paper as outlined in 7.1.2.3.

7.2.4 Cover the filter paper and sample with occlusive surgical tape and an elastic bandage as described in 7.1.2.4.