



Designation: F1408 – 97 (Reapproved 2013)

Standard Practice for Subcutaneous Screening Test for Implant Materials¹

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1. Scope

1.1 This practice covers a short-term testing method to screen the subcutaneous tissue reaction to metallic or other implant candidate materials in small laboratory animals. The material may be dense or porous. The tissue reactions will be evaluated in comparison to those evoked by control materials that are accepted as clinical implant materials.

1.2 This practice, along with other appropriate biological tests (including other ASTM test methods), may be used to assess the biocompatibility of candidate materials for use in the fabrication of devices for clinical application. It may be also applied to evaluate the effect of special surface textures and preparations of known materials.

1.3 This experimental protocol is not designed to provide a comprehensive assessment of the systemic toxicity, carcinogenicity, teratogenicity, or mutagenicity of the material.

1.4 The values stated in SI units are to be regarded as standard. No other units of measurement are included in this standard.

1.5 *This standard does not purport to address all of the safety concerns, if any, associated with its use. It is the responsibility of the user of this standard to establish appropriate safety and health practices and determine the applicability of regulatory limitations prior to use.*

2. Referenced Documents

2.1 *ASTM Standards:*²

[F67 Specification for Unalloyed Titanium, for Surgical Implant Applications \(UNS R50250, UNS R50400, UNS R50550, UNS R50700\)](#)

[F75 Specification for Cobalt-28 Chromium-6 Molybdenum Alloy Castings and Casting Alloy for Surgical Implants \(UNS R30075\)](#)

¹ This practice is under the jurisdiction of ASTM Committee F04 on Medical and Surgical Materials and Devices and is the direct responsibility of Subcommittee F04.16 on Biocompatibility Test Methods.

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² For referenced ASTM standards, visit the ASTM website, www.astm.org, or contact ASTM Customer Service at service@astm.org. For *Annual Book of ASTM Standards* volume information, refer to the standard's Document Summary page on the ASTM website.

[F86 Practice for Surface Preparation and Marking of Metallic Surgical Implants](#)

[F136 Specification for Wrought Titanium-6Aluminum-4Vanadium ELI \(Extra Low Interstitial\) Alloy for Surgical Implant Applications \(UNS R56401\)](#)

[F138 Specification for Wrought 18Chromium-14Nickel-2.5Molybdenum Stainless Steel Bar and Wire for Surgical Implants \(UNS S31673\)](#)

[F648 Specification for Ultra-High-Molecular-Weight Polyethylene Powder and Fabricated Form for Surgical Implants](#)

[F763 Practice for Short-Term Screening of Implant Materials](#)

[F981 Practice for Assessment of Compatibility of Biomaterials for Surgical Implants with Respect to Effect of Materials on Muscle and Bone](#)

3. Summary of Practice

3.1 Under strict aseptic conditions, specimens of the candidate and control materials are implanted subcutaneously in the neck of mice (or other suitable animals). After one, three, and nine weeks the animals are anesthetized and the test samples are excised with an intact tissue envelope. On histologic sections the tissue reactions to the candidate materials are compared with the tissue response to clinically accepted control materials.

4. Significance and Use

4.1 This practice is a guideline for a short-term screening test for the evaluation of the tissue response to materials that may be selected for implantation in the human body. This test may be performed prior to longterm testing (for example, Practice F981) to eliminate unsuitable candidate materials early and to save further animal testing.

4.2 This practice may be used to detect toxic effects of materials in general (see [Appendix X1](#)). However, it is particularly suitable for the testing of materials that are intended to have contact with subcutaneous tissues or soft tissues in general. For materials intended to be inserted specifically into muscle tissues, Practice F763 should be considered as a short term test method.

4.3 The suggested implant specimens are cylindrical. A special grooved type of cylinder may be used (see [Fig. X2.1](#) of

Appendix X2) to allow tissue interlocking that could keep the implant in place and minimize tissue irritation through motion at the interface that otherwise could contribute to increased variance of the results. In case ungrooved cylinders are used (see Fig. X1.2 of **Appendix X2**), probable motion at the implant/tissue interface must be taken into account. Control cylinders should be shaped like the test cylinders.

4.4 The type of surface preparation of the specimens can affect the tissue reaction, therefore the preparation procedure should be noted in the report. The test may be used to compare the effect of different surface structures or conditions of the same material or to assess the effect of various treatments or modifications of a material.

5. Test Animals and Sites

5.1 Mice of an established strain, (preferably females), are used as test hosts. The test may be adapted to other suitable test animals (for example, rats).

5.2 The implant specimens of control and candidate materials are inserted subcutaneously in the neck of the host.

5.3 One implant is inserted per mouse. Therefore, the number of animals is identical with the number of test specimens. If rats or other larger suitable animals are used, more than one test specimen may be implanted, but the implants should never be allowed to come in contact with each other. If animals other than mice are large enough, cylinders of the candidate and control material may be implanted separately at the right and the left side of the neck in one animal.

6. Implant Specimens

6.1 *Specimen Design*—Cylinders of 7 mm length and 4 mm diameter are prepared for implantation in mice. Special specimens with two grooves are designed corresponding to the figures in **Appendix X2**. If larger animal hosts are used, the implant dimensions may be increased proportionally. If it is impossible to prepare specimens of this kind, the specimen configuration used must be described fully in the report. Implant specimens from the candidate and control material should always have the same dimensions.

6.2 *Selection of Control Materials*—Recommended metals for use as control materials include those given in Specifications **F67**, **F75**, **F136**, and **F138**. However, for specific applications any metal of known compatibility and standardized as implant material may be employed as control material for comparison. To study adverse tissue reactions, a non-compatible material like copper may be used as a positive control material. A suitable polymeric control material like the polyethylene USP negative control plastic, RS, or UHMWPE (see Specification **F648**) may be used.

6.3 *Specimen Surface*—The surface of specimens from prospective implant materials should be treated in the same manner as the implant intended for clinical application in the human patient. Depending on the particular issue, the control specimens should have either a surface condition as it is normally used for clinical applications or a surface condition

most similar to that of the tested candidate material. For preparation of metallic materials Practice **F86** should be considered.

6.4 *Numbers of Test and Control Implants*—Per each time period, at least six implant specimens of each candidate and control material should be evaluated in mice (one per mouse). If more than one specimen is implanted in larger test hosts, at least four animals should be used per material and time period.

6.5 *Conditioning*—The cleaning, sterilization, and packaging should be the same as used for implantation in the human patient. After surface preparation and sterilization the implant specimens should be protected from surface alterations and contamination and should be handled with non-metallic forceps when appropriate. When plastified forceps are used, be sure that no plastic material is transferred to the implant surface.

7. Procedure

7.1 *Implantation:*

7.1.1 Implant the specimens under sterile conditions in anesthetized animals. The incision site is remote from the implantation site to prevent infection around the implant. In mice, make a 1 cm long incision above the sacrum and prepare a subcutaneous tunnel toward the neck.

7.1.2 Push the implant through the tunnel and position at the neck. In some distance of the implant close the tunnel with three stitches with a thread of a non-metallic suture material to prevent moving of the implant. Then close the incision. (Do not place the implant directly underneath the incision to avoid infection.)

7.1.3 Keep the individually marked animals in standard cages that comply with current animal protection requirements. Keep mice up to three or four weeks in individual cages.

7.2 *Post-Operative Care*—Care of the animals should be in accordance with accepted standards as outlined in the *Guide for the Care and Use of Laboratory Animals*.³

7.2.1 Carefully observe each animal during the period of assay, and report any abnormal clinical findings.

7.2.2 If infection or injury of the test implant site invalidates the results, replace the animal so that the number of retrieved implants will be at least that of the schedule.

7.2.3 If an animal dies prior to the expected date of sacrifice, autopsy it and determine the cause of death. Replace the animal if the cause of death is unrelated to the test procedure or the test material. Include the test animal in the assay of data if the cause of death is related to the procedure or test material.

7.3 *Sacrifice and Implant Retrieval:*

7.3.1 Sacrifice the animals after one, three, and nine weeks. If longer time intervals are of interest, mice may be kept up to 24 weeks. Examine and report the status of the health of the animals prior to euthanasia.

7.3.2 At sacrifice, record any gross abnormalities of color or consistency observed on the tissues surrounding the implant.

³ *The Guide for Care and Use of Laboratory Animals*, Institute of Laboratory Animal Research Publication. Available from National Academy Press, 500 Fifth St., NW, Lockbox 285, Washington, DC 20055.