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Standard Test Method for Evaluating the Bacterial Filtration Efficiency (BFE) of Medical Face Mask Materials, Using a Biological Aerosol of Staphylococcus aureus¹

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INTRODUCTION

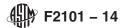
Workers, primarily those in the health care profession, involved in treating and caring for individuals injured or sick, as well as the patient, can be exposed to biological aerosols capable of transmitting disease. These diseases, which may be caused by a variety of microorganisms, can pose significant risks to life and health. Since engineering controls can not eliminate all possible exposures, attention is placed on reducing the potential of airborne exposure through the use of medical face masks.

1. Scope

- 1.1 This test method is used to measure the bacterial filtration efficiency (BFE) of medical face mask materials, employing a ratio of the upstream bacterial challenge to downstream residual concentration to determine filtration efficiency of medical face mask materials.
- 1.2 This test method is a quantitative method that allows filtration efficiency for medical face mask materials to be determined. The maximum filtration efficiency that can be determined by this method is 99.9 %.
- 1.3 This test method does not apply to all forms or conditions of biological aerosol exposure. Users of the test method should review modes for worker exposure and assess the appropriateness of the method for their specific applications.
- 1.4 This test method evaluates medical face mask materials as an item of protective clothing but does not evaluate materials for regulatory approval as respirators. If respiratory protection for the wearer is needed, a NIOSH-certified respirator should be used. Relatively high bacterial filtration efficiency measurements for a particular medical face mask material does not ensure that the wearer will be protected from biological aerosols since this test method primarily evaluates the performance of the composite materials used in the construction of the medical face mask and not its design, fit or facial sealing properties.
- 1.5 *Units*—The values stated in SI units or inch-pound units are to be regarded separately as standard. The values stated in each system may not be exact equivalents; therefore, each system shall be used independently of the other. Combining values from the two systems may result in nonconformance of the standard.
- 1.6 This test method does not address breathability of the medical face mask materials or any other properties affecting the ease of breathing through the medical face mask material.
- 1.7 This test method may also be used to measure the bacterial filtration efficiency (BFE) of other porous medical products such as surgical gowns, surgical drapes, and sterile barrier systems.
- 1.8 This standard does not purport to address all of the safety concerns, if any, associated with its use. It is the responsibility of the user of this standard to establish appropriate safety and health practices and determine the applicability of regulatory limitations prior to use.

¹ This test method is under the jurisdiction of ASTM Committee F23 on Personal Protective Clothing and Equipment and is the direct responsibility of Subcommittee F23.40 on Biological.

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2. Referenced Documents

2.1 ASTM Standards:²

E171 Practice for Conditioning and Testing Flexible Barrier Packaging

F1494 Terminology Relating to Protective Clothing

2.2 ANSI/ASQC Standard:³

ANSI/ASQC Z1.4 Sampling Procedures and Tables for Inspection by Attributes

2.3 ISO Standard:⁴

ISO 2859-1 Sampling Plans for Inspection by Attributes

2.4 Military Standard:⁵

MIL-STD 36954C (1973) Military Specification: Mask, Surgical, Disposable

3. Terminology

- 3.1 Definitions:
- 3.1.1 *aerosol*, *n*—a suspension of solid or liquid particles in a gas.
- 3.1.2 agar, n—a semi-solid culture medium used to support the growth of bacteria and other micro-organisms.
- 3.1.3 airborne exposure pathways, n—inhalation routes of exposure to the medical face mask wearer.
- 3.1.4 bacterial filtration efficiency (BFE), n—the effectiveness of a medical face mask material in preventing the passage of aerosolized bacteria; expressed in the percentage of a known quantity that does not pass the medical face mask material at a given aerosol flow rate.
 - 3.1.5 biological aerosol, n—a suspension of particles containing biological agents which have been dispersed in a gas.
- 3.1.6 *blood-borne pathogen, n*—an infectious bacterium or virus, or other disease inducing microbe carried in blood or other potentially infectious body fluids.
 - 3.1.7 body fluid, n—any liquid produced, secreted, or excreted by the human body.
- 3.1.8 *protective clothing, n*—an item of clothing that is specifically designed and constructed for the intended purpose of isolating all or part of the body from a potential hazard; or, isolating the external environment from contamination by the wearer of the clothing.
- 3.1.9 *medical face mask*, *n*—an item of protective clothing designed to protect portions of the wearer's face, including the mucous membrane areas of the wearer's nose and mouth, from contact with blood and other body fluids during medical procedures.

3.1.9.1 Discussion—

Medical face masks also function to partly limit the spread of biological contamination from the mask wearer (health care provider) to the patient.

3.2 For definitions of other protective clothing-related terms used in this test method, refer to Terminology F1494.

4. Summary of Test Method

- 4.1 The medical face mask material is clamped between a six-stage cascade impactor and an aerosol chamber. The bacterial aerosol is introduced into the aerosol chamber using a nebulizer and a culture suspension of *Staphylococcus aureus*. The aerosol is drawn through the medical face mask material using a vacuum attached to the cascade impactor. The six-stage cascade impactor uses six agar plates to collect aerosol droplets which penetrate the medical face mask material. Control samples are collected with no test specimen clamped in the test apparatus to determine the upstream aerosol counts.
- 4.2 The agar plates from the cascade impactor are incubated for 48 h and counted to determine the number of viable particles collected. The ratio of the upstream counts to the downstream counts collected for the test specimen are calculated and reported as a percent bacterial filtration efficiency.

5. Significance and Use

5.1 This test method offers a procedure for evaluation of medical face mask materials for bacterial filtration efficiency. This test method does not define acceptable levels of bacterial filtration efficiency. Therefore, when using this test method it is necessary to describe the specific condition under which testing is conducted.

² For referenced ASTM standards, visit the ASTM website, www.astm.org, or contact ASTM Customer Service at service@astm.org. For *Annual Book of ASTM Standards* volume information, refer to the standard's Document Summary page on the ASTM website.

³ Available from American Society for Quality (ASQ), 600 N. Plankinton Ave., Milwaukee, WI 53203, http://www.asq.org.

⁴ Available from American National Standards Institute (ANSI), 25 W. 43rd St., 4th Floor, New York, NY 10036, http://www.ansi.org. C/astm-f2 101-14

⁵ Available from Standardization Documents Order Desk, Bldg. 4 Section D, 700 Robbins Ave., Philadelphia, PA 19111-5094, Attn: NPODS.

- 5.2 This test method has been specifically designed for measuring bacterial filtration efficiency of medical face masks, using Staphylococcus aureus as the challenge organism. The use of S. aureus is based on its clinical relevance as a leading cause of nosocomial infections.
- 5.3 This test method has been designed to introduce a bacterial aerosol challenge to the test specimens at a flow rate of 28.3 L/mm. (1 ft³/min). This flow rate is within the range of normal respiration and within the limitations of the cascade impactor.
- 5.4 This test method allows the aerosol challenge to be Unless otherwise specified, the testing shall be performed with the inside of the medical face mask in contact with the bacterial challenge. Testing may be performed with the aerosol challenge directed through either the face side or liner side of the test specimen, thereby, allowing evaluation of filtration efficiencies which relate to both patient-generated aerosols and wearer-generated aerosols.
- 5.5 Degradation by physical, chemical, and thermal stresses could negatively impact the performance of the medical face mask material. The integrity of the material can also be compromised during use by such effects as flexing and abrasion, or by wetting with contaminants such as alcohol and perspiration. Testing without these stresses could lead to a false sense of security. If these conditions are of concern, evaluate the performance of the medical face mask material for bacterial filtration efficiency following an appropriate pretreatment technique representative of the expected conditions of use. Consider preconditioning to assess the impact of storage conditions and shelf life for disposable products, and the effects of laundering and sterilization for reusable products.
- 5.6 If this procedure is used for quality control, perform proper statistical design and analysis of larger data sets. This type of analysis includes, but is not limited to, the number of individual specimens tested, the average percent bacterial filtration efficiency, and standard deviation. Data reported in this way help to establish confidence limits concerning product performance. Examples of acceptable sampling plans are found in references such as ANSI/ASQC Z1.4 and ISO 2859-1.

6. Apparatus and Materials

- 6.1.1 Autoclave, capable of maintaining 121-123°C.

 Standards
- 6.1.2 *Incubator*, capable of maintaining $37 \pm 2^{\circ}$ C.
- 6.1.3 Analytical Balance, capable of weighing 0.001 g.
- 6.1.4 Vortex Mixer, capable of mixing the contents of 16 mm × 150 mm test tubes.
- 6.1.5 Orbital Shaker, capable of achieving 100-250 rpm.
- 6.1.6 Refrigerator, capable of maintaining 2-8°C.
- 6.1.7 Six-Stage Viable Particle Cascade Impactor.
- 6.1.8 Vacuum Pump, capable of 57 L/m (2 ft³/mm).
- 6.1.9 Air Pump/Compressor, capable of 15 psig minimum.
- 6.1.10 Peristaltic Pump, capable of delivering 0.01 mL/min.
- 6.1.11 Nebulizer, capable of delivering a mean particle size of 3.0 μ m \pm 0.3 μ m and a challenge level of 2200 \pm 500 viable particles per test, as determined according to step 12.3.
 - 6.1.12 Glass Aerosol Chamber, 60 cm by 8 cm diameter tube.
 - 6.1.13 Colony Counter, manual or automatic, capable of counting up to 400 colonies/plate.
 - 6.1.14 Timers, capable of 0.1 s accuracy.
 - 6.1.15 Automatic Pipetor, capable of delivering 1.0 mL \pm 0.05 mL.
 - 6.1.16 Flow Meters, capable of 28.3 L/min.
 - 6.1.17 Aerosol Condenser.
 - 6.1.18 Pressure Gauge, capable of 35 kPa ± 1 kPa accuracy.
 - 6.1.19 Air Regulator.
 - 6.2 Materials:
 - 6.2.1 Flasks, 250-500 mL Erlenmeyer.
 - 6.2.2 Petri Dishes, sterile 15 by 100 mm.
 - 6.2.3 Pipettes, 1 mL, 5 mL, and 10 mL.
 - 6.2.4 Test Tube Rack, stainless
 - 6.2.5 Bottles, sterile, glass, 100-500 mL capacity.
 - 6.2.6 *Inoculating Loop.*
 - 6.2.7 Stoppers/Closures, of appropriate size to fit test tubes.
 - 6.2.8 Test Tubes, 16 mm × 150 mm.