

## **Animal and vegetable fats and oils — Determination of fatty-acid-bound chloropropanediols (MCPDs) and glycidol by GC/MS —**

### **Part 3: Method using acid transesterification and measurement for 2-MCPD, 3-MCPD and glycidol**

*Corps gras d'origines animale et végétale — Détermination des esters de chloropropanediols (MCPD) et d'acides gras et des esters de glycidol et d'acides gras par CPG/SM —*

*Partie 3: Méthode par transestérification acide et mesure ~~pour le~~ 2-MCPD, ~~le~~ 3-MCPD et ~~le~~ glycidol*

<https://standards.iteh.ai/catalog/standards/iso/3df5ca72-aa17-41b4-a993-f79fa448af60/iso-fdis-18363-3>

**FDIS stage**

© ISO 2024

All rights reserved. Unless otherwise specified, or required in the context of its implementation, no part of this publication may be reproduced or utilized otherwise in any form or by any means, electronic or mechanical, including photocopying, or posting on the internet or an intranet, without prior written permission. Permission can be requested from either ISO at the address below or ISO's member body in the country of the requester.

ISO copyright office  
CP 401 • Ch. de Blandonnet 8  
CH-1214 Vernier, Geneva  
Phone: + 41 22 749 01 11  
E-mail: [copyright@iso.org](mailto:copyright@iso.org)  
Website: [www.iso.org](http://www.iso.org)

Published in Switzerland

# iTeh Standards (<https://standards.iteh.ai>) Document Preview

[ISO/FDIS 18363-3](#)

<https://standards.iteh.ai/catalog/standards/iso/3df5ca72-aa17-41b4-a993-f79fa448af60/iso-fdis-18363-3>

**Contents**

Foreword .....iv

Introduction..... v

1 Scope ..... 1

2 Normative references..... 1

3 Terms and definitions ..... 1

4 Principle..... 2

5 Reagents..... 2

5.1 Standard and reference compounds..... 2

5.2 Standard solutions..... 3

5.2.1 General..... 3

5.2.2 Stock solutions (1 mg/ml)..... 3

5.2.3 Working solutions..... 3

5.3 Other reagents..... 4

5.4 Reagent solutions..... 4

6 Apparatus ..... 5

7 Sample ..... 5

7.1 Sampling ..... 5

7.2 Preparation of the test sample ..... 5

8 Procedure..... 6

8.1 Test sample preparation..... 6

8.2 Preparation of the calibration curve..... 7

8.3 Gas chromatography/mass spectrometry references..... 7

9 Expression of results ..... 8

9.1 Quantification of 3-MCPD esters ..... 8

9.2 Quantification of 2-MCPD esters ..... 8

9.3 Quantification of glycidyl esters ..... 9

10 Precision ..... 10

10.1 General ..... 10

10.2 Repeatability..... 10

10.3 Between-day reproducibility ..... 10

11 Test report ..... 10

Annex A (informative) Construction of the calibration curves ..... 11

Annex B (informative) Results of the interlaboratory test ..... 18

Bibliography ..... 20

## Foreword

ISO (the International Organization for Standardization) is a worldwide federation of national standards bodies (ISO member bodies). The work of preparing International Standards is normally carried out through ISO technical committees. Each member body interested in a subject for which a technical committee has been established has the right to be represented on that committee. International organizations, governmental and non-governmental, in liaison with ISO, also take part in the work. ISO collaborates closely with the International Electrotechnical Commission (IEC) on all matters of electrotechnical standardization.

The procedures used to develop this document and those intended for its further maintenance are described in the ISO/IEC Directives, Part 1. In particular, the different approval criteria needed for the different types of ISO documents should be noted. This document was drafted in accordance with the editorial rules of the ISO/IEC Directives, Part 2 (see [www.iso.org/directives](http://www.iso.org/directives)).

~~Attention is drawn~~ISO draws attention to the possibility that ~~some of the~~ ~~elements~~ ~~implementation~~ of this document may ~~be involve~~ the ~~subject~~ ~~use of (a) patent(s)~~. ISO takes no position concerning the evidence, validity or applicability of ~~any claimed~~ patent rights in respect thereof. As of the date of publication of this document, ISO had not received notice of (a) patent(s) which may be required to implement this document. However, implementers are cautioned that this may not represent the latest information, which may be obtained from the patent database available at [www.iso.org/patents](http://www.iso.org/patents). ISO shall not be held responsible for identifying any or all such patent rights. ~~Details of any patent rights identified during the development of the document will be in the Introduction and/or on the ISO list of patent declarations received (see [www.iso.org/patents](http://www.iso.org/patents)).~~

Any trade name used in this document is information given for the convenience of users and does not constitute an endorsement.

For an explanation ~~on~~of the voluntary nature of standards, the meaning of ISO specific terms and expressions related to conformity assessment, as well as information about ISO's adherence to the World Trade Organization (WTO) principles in the Technical Barriers to Trade (TBT), see [www.iso.org/iso/foreword.html](http://www.iso.org/iso/foreword.html) see the following URL: [www.iso.org/iso/foreword.html](http://www.iso.org/iso/foreword.html).

~~This document was prepared by Technical Committee ISO/TC 34, Food products, Subcommittee SC 11, Animal and vegetable fats and oils, in collaboration with the European Committee for Standardization (CEN) Technical Committee CEN/TC 307, Oilseeds, vegetable and animal fats and oils and their by-products – Methods of sampling and analysis, in accordance with the Agreement on technical cooperation between ISO and CEN (Vienna Agreement).~~

~~This second edition cancels and replaces the first edition (ISO 18363-3:2017), of which it constitutes a minor revision was performed to bring. The changes are as follows:~~

~~— the text of the Introduction in line has been revised to be consistent with the latest part in the ISO 18363 series. 4:2021.~~

A list of all parts in the ISO 18363 series can be found on the ISO website.

~~Any feedback or questions on this document should be directed to the user's national standards body. A complete listing of these bodies can be found at [www.iso.org/members.html](http://www.iso.org/members.html).~~

## Introduction

The ISO 18363 series ~~is a family of International Standards which~~ can be used for the determination of ester-bound MCPD and glycidol. This introduction describes the methods specified in the ~~four documents currently published or proposed~~ different parts so that the analyst can decide which methods are suitable for application. The detailed application of each method is contained within the scope of each individual method.

ISO 18363-1 is a differential method equivalent to ~~the~~-DGF standard C-VI 18 (10)<sup>[11]</sup> and identical to AOCS Official Method Cd 29c-13<sup>[10]</sup>. In brief, it is based on a fast alkaline catalysed release of 3-MCPD and glycidol from the ester derivatives. Glycidol is subsequently converted into induced 3-MCPD. It consists of two parts. The first part (A) allows the determination of the sum of ester-bound 3-MCPD and ester-bound glycidol, whereas the second part (B) determines ester-bound 3-MCPD only. Both assays are based on the release of the target analytes 3-MCPD and glycidol from the ester-bound form by an alkaline catalysed alcoholysis carried out at room temperature. In part A, an acidified sodium chloride solution is used to stop the reaction and subsequently convert the glycidol into induced 3-MCPD. Thus, 3-MCPD and glycidol become indistinguishable in part A. In part B, the reaction stop is achieved by the addition of an acidified chloride-free salt solution which also prevents the conversion of glycidol into induced MCPD. Consequently, part B allows the determination of the genuine 3-MCPD content. Finally, the glycidol content of the sample is proportional to the difference of both assays (A – B) and can be calculated when the transformation ratio from glycidol to 3-MCPD has been determined. ISO 18363-1 is applicable to the fast determination of ester-bound 3-MCPD and glycidol in refined and non-refined vegetable oils and fats. ISO 18363-1 can also apply to animal fats and used frying oils and fats, but a validation study must be undertaken before the analysis of these matrices. Any free analytes within the sample would be included in the results, but the document does not allow the distinction between free and bound analytes. However, as of publication of this document, research has not shown any evidence of a free analyte content as high as the esterified analyte content in refined vegetable oils and fats. In principle, ISO 18363-1 can also be modified in such a way that the determination of 2-MCPD is feasible, but again, a validation study must be undertaken before the analysis of this analyte.

ISO 18363-2 represents ~~the~~-AOCS Official Method Cd 29b-13<sup>[9]</sup>. In brief, it is based on a slow alkaline release of MCPD and glycidol from the ester derivatives. Glycidol is subsequently converted into 3-MBPD. ISO 18363-2 consists of two sample preparations that differ in the use of internal standards. Both preparations are used for the determination of ester-bound 2-MCPD and 3-MCPD. In part A, a preliminary result for ester-bound glycidol is determined. Because the 3-MCPD present in the sample is converted to some minor extent into induced glycidol by the sample preparation, part B serves to quantify this amount of induced glycidol that is subsequently subtracted from the preliminary glycidol result of part A. By the use of isotopically labelled free MCPD isomers in assay A and isotopically labelled ester-bound 2-MCPD and 3-MCPD in part B, the efficiency of ester cleavage can be monitored. Both assays A and B are based on the release of the target analytes 2-MCPD, 3-MCPD, and glycidol from the ester-bound form by a slow alkaline catalysed alcoholysis in the cold. In both sample preparations, the reaction is stopped by the addition of an acidified concentrated sodium bromide solution so as to convert the unstable and volatile glycidol into 3-MBPD which shows comparable properties to 3-MCPD with regard to its stability and chromatographic performance. Moreover, the major excess of bromide ions prevents the undesired formation of 3-MCPD from glycidol in the case of samples which contain naturally occurring amounts of chloride. ISO 18363-2 is applicable to the determination of ester-bound 3-MCPD, 2-MCPD, and glycidol in refined and unrefined vegetable oils and fats. It also applies to animal fats and used frying oils and fats, but a validation study must be undertaken before the analysis of these matrices. Any free analytes within the sample are included in the results, but the document does not allow the distinction between free and bound analytes. However, as of publication of this document, research has not shown any evidence of a free analyte content as high as the esterified analyte content in refined vegetable oils and fats.

This document (*i.e.* ISO 18363-3) represents AOCS Official Method Cd 29a-13<sup>[8]</sup>. In brief, it is based on the conversion of glycidyl esters into 3-MBPD esters and a slow acid catalysed release of MCPD and MBPD from the ester derivatives. This document is based on a single sample preparation in which glycidyl esters are converted into MBPD monoesters, and subsequently, the free analytes 2-MCPD, 3-MCPD, and 3-MBPD are released by a slow acid catalysed alcoholysis. The 3-MBPD represents the genuine content of bound glycidol.

## ISO/FDIS 18363-3:2024(en)

This document is applicable ~~for~~to the determination of ester-bound 2-MCPD, 3-MCPD, and glycidol in refined and non-refined vegetable oils and fats. It also applies to animal fats and used frying oils and fats, but a validation study must be undertaken before the analysis of these matrices. The method is suited for the analysis of bound (esterified) analytes, but if required, this document can also be performed without the initial conversion of glycidyl esters. In such a setup, both free and bound 2-MCPD and 3-MCPD forms are included in the results and the amount of free analytes can be calculated as the difference between two determinations performed in both setups. However, as of publication of this document, research has not shown any evidence of a free analyte content as high as the esterified analyte content in refined vegetable oils and fats.

ISO 18363-4 specifies a rapid procedure based on fast alkaline cleavage of the MCPD and glycidyl esters. The released glycidol is subsequently converted into 3-MBPD. The pH of the fast alkaline cleavage generally causes the released MCPD to partially convert to glycidol during the cleavage of the esters, leading to overestimation of the glycidyl ester content of the sample. By adding two distinct isotopically labelled ester-bound 3-MCPD and glycidol internal standards, it is possible to quantify the amount of labelled glycidol resulting from the degradation of the released internal standard. This information can be used to correct for overestimation of the glycidyl ester induced glycidol by 3-MCPD induced glycidol. The same two internal standards are used for quantification of the bound MCPD and glycidol, requiring a single sample preparation to quantify bound 2-MCPD-, 3-MCPD- and glycidol esters. In analogue with ISO 18363-1, ISO 18363-2 and ~~ISO 18363-3~~this document, the released MCPDs and 3-MBPD are derivatized with phenylboronic acid before GC-MS/MS analysis. In contrast to the other parts of the ISO 18363 series, ISO 18363-4 requires GC-MS/MS instrumentation to unambiguously detect each of the (isotopically labelled) MBPDs required for correct quantification of the glycidyl ester induced glycidol. ISO 18363-4 is applicable to the determination of ester-bound 3-MCPD, 2-MCPD and glycidol in refined and unrefined vegetable oils and fats. It also applies to animal fats and used frying oils and fats, but a validation study must be undertaken before analysis of these matrices. Any free analytes within the sample are included in the results, but ISO 18363-4 will not allow the distinction between free and bound analytes. However, as of publication of this document, research has not shown any evidence of a free analyte content as high as the esterified analyte content in refined vegetable oils and fats.

## Document Preview

[ISO/FDIS 18363-3](https://standards.iteh.ai/catalog/standards/iso/3df5ca72-aa17-41b4-a993-f79fa448af60/iso-fdis-18363-3)

<https://standards.iteh.ai/catalog/standards/iso/3df5ca72-aa17-41b4-a993-f79fa448af60/iso-fdis-18363-3>

# Animal and vegetable fats and oils — Determination of fatty-acid-bound chloropropanediols (MCPDs) and glycidol by GC/MS —

## Part 3:

## Method using acid transesterification and measurement for 2-MCPD, 3-MCPD and glycidol

### 1 Scope

This document specifies a procedure for the simultaneous determination of 2-MCPD esters (bound 2-MCPD), 3-MCPD esters (bound 3-MCPD) and glycidyl esters (bound glycidol) in a single assay, based on acid catalysed ester cleavage and derivatization of cleaved (free) analytes with phenylboronic acid (PBA) prior to GC/MS analysis.

This document is applicable to solid and liquid fats and oils. For all three analytes the limit of quantification (LOQ) is 0,1 mg/kg and the limit of detection (LOD) is 0,03 mg/kg.

### 2 Normative references

The following documents are referred to in the text in such a way that some or all of their content constitutes requirements of this document. For dated references, only the edition cited applies. For undated references, the latest edition of the referenced document (including any amendments) applies.

ISO 3696, *Water for analytical laboratory use — Specification and test methods*

### 3 Terms and definitions

For the purposes of this document, the following terms and definitions apply.

ISO and IEC maintain **terminological terminology** databases for use in standardization at the following addresses:

— IEC Electropedia: available at

— ISO Online browsing platform: available at <https://www.iso.org/obp>

— IEC Electropedia: available at <https://www.electropedia.org/>

#### 3.1

##### bound 2-MCPD

amount of 2-MCPD cleaved from its esterified (bound) forms by acid catalysed transesterification according to the reference method

Note 1 to entry: The content of 2-MCPD is calculated and reported as a mass fraction, in milligrams per kilogram (mg/kg).

#### 3.2

##### bound 3-MCPD

amount of 3-MCPD cleaved from its esterified (bound) forms by acid catalysed transesterification according to the reference method

Note 1 to entry: The content of 3-MCPD is calculated and reported as a mass fraction, in milligrams per kilogram (mg/kg).



### 3.3

#### bound glycidol

amount of glycidol cleaved from its esterified (bound) forms by acid catalysed transesterification according to the reference method

Note 1 to entry: The content of glycidol is calculated and reported as a mass fraction, in milligrams per kilogram (mg/kg).

## 4 Principle

The oil/fat sample is dissolved in tetrahydrofuran, and the internal standards (pentadeuterated 3-MCPD diester and pentadeuterated glycidyl ester) are added. During the first step of sample preparation, glycidyl esters are converted into 3-MBPD monoesters by the addition of an acidified solution of sodium bromide. Upon completion of the reaction, the organic phase, containing 2- and 3-MCPD esters and 3-MBPD esters, is separated and evaporated to dryness. In the second step the residue is dissolved in tetrahydrofuran and the acid transesterification is initiated by the addition of an acid alcoholic solution. After 16 h incubation at 40 °C, the sample mixture is neutralized and the fatty acid methyl esters generated during the transesterification are removed. Finally, the purified sample [containing cleaved (free) analytes] is derivatized with phenylboronic acid prior to GC/MS analysis.

The quantification of 2- and 3-MCPD esters (expressed as bound 2- and 3-MCPD) is based on the 2-MCPD/3-MCPD-d5 and 3-MCPD/3-MCPD-d5 signal ratio, respectively. The quantification of glycidyl esters (expressed as bound glycidol) is based on the 3-MBPD/3-MBPD-d5 signal ratio.

This method allows the simultaneous quantification of all three analytes in a single assay.

## 5 Reagents

**WARNING** — This document requires handling of hazardous substances. Technical, organizational, and personal safety measures shall be followed.

Unless otherwise stated, analytically pure reagents shall be used. Water shall ~~comply with~~ conform to grade 3 of ISO 3696.

### 5.1 Standard and reference compounds

**5.1.1 1,2-Dipalmitoyl-3-chloropropanediol (PP-3-MCPD)**, purity  $\geq 95$  % (e.g. from a supplier or synthesized from 3-MCPD and palmitoyl chloride as described by Reference [1]).

NOTE 1,2-Dipalmitoyl-3-chloropropanediol can be substituted by 1,2-dioleoyl-3-chloropropanediol or other fatty acid diesters of 3-MCPD with similar chain length (C16-C18 are preferred as they are the most abundant in the majority of oils/fats).

**5.1.2 1,3-Dipalmitoyl-2-chloropropanediol (PP-2-MCPD)**, purity  $\geq 95$  % (e.g. synthesized from 2-MCPD and palmitoyl chloride as described by Reference [1]).

NOTE In analogy with the recommendations given for PP-3-MCPD, 1,3-dipalmitoyl-2-chloropropanediol can be substituted by other fatty acid diesters of 2-MCPD with similar chain length (C16-C18 are preferred as they are the most abundant in the majority of oils/fats).

**5.1.3 Pentadeuterated 1,2-dipalmitoyl-3-chloropropanediol (PP-3-MCPD-d5)**, purity  $\geq 95$  %.

NOTE The same consideration applied to 1,2-dipalmitoyl-3-chloropropanediol is valid also for its pentadeuterated analogue, see the ~~Note~~note in 5.1.1.

**5.1.4 Glycidyl palmitate (Gly-P)**, purity  $\geq 98$  %.



NOTE Glycidyl palmitate can be substituted by glycidyl oleate or other fatty acid esters of glycidol with similar chain length (C16-C18 are preferred as they are the most abundant in the majority of oils/fats).

### 5.1.5 Pentadeuterated glycidyl palmitate (Gly-P-d5), purity $\geq$ 98 %.

NOTE The same consideration applied to glycidyl palmitate is valid also for its pentadeuterated analogue, see the ~~Note~~note in [5.1.4](#).

## 5.2 Standard solutions

### 5.2.1 General

All standard solutions can be prepared with either toluene ([5.3.5](#)) or tetrahydrofuran ([5.3.1](#)). Toluene is preferred for standard solutions containing glycidyl esters.

### 5.2.2 Stock solutions (1 mg/ml)

- a) Weigh 10 mg of PP-3-MCPD ([5.1.1](#)) in a 10 ml volumetric flask. Fill up to the mark, making sure that the standard is completely dissolved in the solvent.
- b) Weigh 10 mg of PP-2-MCPD ([5.1.2](#)) in a 10 ml volumetric flask. Fill up to the mark, making sure that the standard is completely dissolved in the solvent.
- c) Weigh 10 mg of PP-3-MCPD-d5 ([5.1.3](#)) in a 10 ml volumetric flask. Fill up to the mark, making sure that the standard is completely dissolved in the solvent.
- d) Weigh 10 mg of Gly-P ([5.1.4](#)) in a 10 ml volumetric flask. Fill up to the mark, making sure that the standard is completely dissolved in the solvent.
- e) Weigh 10 mg of Gly-P-d5 ([5.1.5](#)) in a 10 ml volumetric flask. Fill up to the mark, making sure that the standard is completely dissolved in the solvent.

NOTE Stock solutions are stable for at least three months when stored at  $-18\text{ }^{\circ}\text{C}$ .

### 5.2.3 Working solutions

- a) Calibration I (PP-3-MCPD, 55  $\mu\text{g/ml}$ ). Pipette 550  $\mu\text{l}$  of the stock solution [[5.2.2 a\)](#)] into a 10 ml volumetric flask and fill up to the mark with the solvent.
- b) Calibration II (PP-3-MCPD, 5,5  $\mu\text{g/ml}$ ). Pipette 1 ml of the Calibration I solution [[5.2.3 a\)](#)] into a 10 ml volumetric flask and fill up to the mark with the solvent.
- c) Calibration III (PP-2-MCPD, 55  $\mu\text{g/ml}$ ). Pipette 550  $\mu\text{l}$  of the stock solution [[5.2.2 b\)](#)] into a 10 ml volumetric flask and fill up to the mark with the solvent.
- d) Calibration IV (PP-2-MCPD, 5,5  $\mu\text{g/ml}$ ). Pipette 1 ml of the Calibration III solution [[5.2.3 c\)](#)] into a 10 ml volumetric flask and fill up to the mark with the solvent.
- e) Calibration V (Gly-P, 100  $\mu\text{g/ml}$ ). Pipette 1 ml of the stock solution [[5.2.2 d\)](#)] into a 10 ml volumetric flask and fill up to the mark with the solvent.
- f) Calibration VI (Gly-P, 10  $\mu\text{g/ml}$ ). Pipette 1 ml of the Calibration V solution [[5.2.3 e\)](#)] into a 10 ml volumetric flask and fill up to the mark with the solvent.
- g) Internal standard I (PP-3-MCPD-d5, 40  $\mu\text{g/ml}$ ). Pipette 400  $\mu\text{l}$  of the stock solution [[5.2.2 c\)](#)] into a 10 ml volumetric flask and fill up to the mark with the solvent.

- h) Internal standard II (Gly-P-d5, 50 µg/ml). Pipette 500 µl of the stock solution [5.2.2 e)] into a 10 ml volumetric flask and fill up to the mark with the solvent.

NOTE 1 As an alternative to the preparation of separate standard solutions for each analyte, the three [5.2.3 a), c), and e)] can be combined in one single standard solution at high concentration of all three analytes (“mixed Calibration I”). To prepare the mixed solution, pipette 550 µl of PP-3-MCPD stock solution [5.2.2 a)], 550 µl of PP-2-MCPD stock solution [5.2.2 b)], and 1 ml of Gly-P stock solution [5.2.2 d)] into a 10 ml volumetric flask and fill up to the mark with the solvent. Also, the solutions [5.2.3 b), d), and f)] can be combined in one single standard solution at low concentration of all three analytes (“mixed Calibration II”). To prepare the mixed solution, pipette 1 ml of mixed Calibration I into a 10 ml volumetric flask and fill up to the mark with the solvent.

NOTE 2 The internal standard solutions [5.2.3 g and h)] can also be combined in a single solution (“mixed internal standard”). To prepare the mixed solution, pipette 400 µl of PP-3-MCPD-d5 [5.2.2 c)] and 500 µl of Gly-P-d5 [5.2.2 e)] into a 10 ml volumetric flask and fill up to the mark with solvent.

### 5.3 Other reagents

5.3.1 **Tetrahydrofuran**, anhydrous.

5.3.2 **Methanol**, analytical grade.

5.3.3 **n-Heptane**, analytical grade.

5.3.4 **Acetone**, analytical grade.

5.3.5 **Toluene**, analytical grade.

5.3.6 **Water**, ultra-pure (e.g. obtained by using a purification system).

5.3.7 **Sulfuric acid**, purity ≥ 95 %.

5.3.8 **Sodium hydrogen carbonate**, purity ≥ 99 %.

5.3.9 **Sodium sulfate**, purity ≥ 99 %.

5.3.10 **Phenylboronic acid**, purity ≥ 97 %.

5.3.11 **Sodium bromide**, purity ≥ 99,5 %.

### 5.4 Reagent solutions

5.4.1 **Acid aqueous solution of sodium bromide** [sodium bromide 3 mg/ml, sulfuric acid 5 % (volume fraction)]. Prepare a concentrated aqueous solution of sodium bromide by dissolving 1 g of sodium bromide (5.3.11) in 10 ml of ultrapure water (5.3.6). Transfer 180 µl of the concentrated solution into an empty conical flask. Add 0,3 ml of sulfuric acid (5.3.7) and 5,5 ml of ultra-pure water (5.3.6). Shake vigorously.

It is advisable to freshly prepare the solution on daily basis.

5.4.2 **Sodium hydrogen carbonate solution** (0,6 %<sub>w</sub> mass concentration). Weigh 0,6 g of sodium hydrogen carbonate (5.3.8) in a 100 ml volumetric flask and fill up to the mark with ultra-pure water (5.3.6). Use an ultrasonic bath (6.3) to ensure the complete dissolution of the reagent.

NOTE As an alternative, the solution can be prepared by dilution of the sodium hydrogen carbonate saturated solution (5.4.4).