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Standard Guide for Evaluating Non-culture Microbiological Tests Used for Enumerating Bacteria¹

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1. Scope

1.1 The purpose of this guide is to assist users and producers of non-culture tests in determining the applicability of the test for processing different types of samples and evaluating the accuracy of the results. Culture test procedures such as the Heterotrophic (Standard) Plate Count, the Most Probable Number (MPN) method and the Spread Plate Count are widely cited and accepted for the enumeration of microorganisms. However, these methods have their limitations, such as performance time and degree of accuracy. Moreover any given culture test method typically recovers only a fraction of the total viable microbes present in a sample. It is these limitations that have recently led to the marketing of a variety of non-culture procedures, test kits and instruments.

1.2 Culture test methods estimate microbial population densities based on the ability of microorganisms in a sample to proliferate in or on a specified growth medium, under specified growth conditions. Non-culture test methods attempt to provide the same or complimentary information through the measurement of a different parameter. This guide is designed to assist investigators in assessing the accuracy and precision of non-culture methods intended for the determination of microbial population densities or activities.

1.3 It is recognized that the Heterotrophic Plate Count (HPC) does not recover all microorganisms present in a product or a system (**1**, **2**).² When this problem occurs during the characterization of a microbiological population, alternative standard enumeration procedures may be necessary, as in the case of sulfate-reducing bacteria. At other times, chemical methods that measure the rates of appearance of metabolic derivatives, the utilization of contaminated product components or genetic profile of the microbial population might be indicated. In evaluating non-culture tests, it is possible that the use of these alternative standard procedures might be the only

means available for establishing correlation. In such cases, this guide can serve as a reference for those considerations.

1.4 Because there are so many types of tests that could be considered non-culture based, it is impossible to recommend a specific test protocol with statistical analyses for evaluating the tests. Instead, this guide should assist in determining what types of tests should be considered to verify the utility and identify the limitations of the nonconventional test.

1.5 The values stated in SI units are to be regarded as standard. No other units of measurement are included in this standard.

2. Referenced Documents

2.1 ASTM Standards:³

- D1129 Terminology Relating to Water
- D4012 Test Method for Adenosine Triphosphate (ATP) Content of Microorganisms in Water
- D5245 Practice for Cleaning Laboratory Glassware, Plasticware, and Equipment Used in Microbiological Analyses
- D5465 Practice for Determining Microbial Colony Counts from Waters Analyzed by Plating Methods
- E177 Practice for Use of the Terms Precision and Bias in ASTM Test Methods
- E691 Practice for Conducting an Interlaboratory Study to Determine the Precision of a Test Method
- E1601 Practice for Conducting an Interlaboratory Study to Evaluate the Performance of an Analytical Method
- E2756 Terminology Relating to Antimicrobial and Antiviral Agents

3. Terminology

3.1 Definitions:

3.1.1 For definitions of terms used in this guide refer to Terminologies D1129, E2756, and E177.

3.2 Abbreviations:

3.2.1 HPC—Heterotrophic Plate Count

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² The boldface numbers in parentheses refer to the list of references at the end of this guide.

³ For referenced ASTM standards, visit the ASTM website, www.astm.org, or contact ASTM Customer Service at service@astm.org. For *Annual Book of ASTM Standards* volume information, refer to the standard's Document Summary page on the ASTM website.

4. Summary of Guide

4.1 ASTM standard methods and practices are referenced for use by producers and users in order to determine the potential utility of a non-standard, non-culture test.

4.2 Recognizing that potential users of non-culture test methods might not have the resources with which or capabilities for evaluating the utility of non-standard, non-culture test methods, recommendations are provided to assist those users in identifying the capabilities that qualify microbiological laboratories to perform collaborative studies to evaluate those methods.

5. Significance and Use

5.1 This guide should be used by producers and potential producers of non-culture tests to determine the accuracy, selectivity, specificity, and reproducibility of the tests, as defined in Practice E691. Results of such studies should identify the limitations and indicate the utility or applicability of the non-culture test, or both, for use on different types of samples.

5.2 Non-culture test users and potential users should employ this guide to evaluate results of the non-culture test as compared to their present methods. Practices D5245 and D5465 should be reviewed in regards to the microbiological methods employed. If culture methods have not been used for monitoring the systems, then guidelines are included for obtaining microbiological expertise.

5.3 Utilization of a non-culture test can reduce the time required to determine the microbiological status of the system and detect microbe that are not detected by culture testing. Consequently, non-culture tests can contribute to the improvement in the overall operating efficiency of microbial contamination condition monitoring and diagnostic efforts, and microbicide performance evaluations.

5.4 Detecting microbial contamination levels that exceed predetermined upper control limits indicates the need for an addition of an antimicrobial agent or other corrective maintenance action. By accurately determining this in a shorter time period than is possible than by culture methods, treatment with antimicrobial agents may circumvent more serious problems

than if the treatment were postponed until culture results were available. If the antimicrobial treatment program relied on an inaccurate non-culture test, then unnecessary loss of product and problems associated with inappropriate selection or improper dosing with antimicrobial agents would exist.

5.5 Since many methods based on entirely different chemical and microbiological principles are considered, it is not possible to establish a unique design and recommend a specific method of statistical analyses for the comparisons to be made. It is only possible to present guides that should be followed while performing the experiments. It is also recommended that a statistician be involved in the study.

6. Procedures

6.1 Practice E1601 provides guidance on the evaluation of analytical method performance. The guidance provided below amplifies the processes described in Practice E1601 as they apply to microbiological test methods.

6.2 Although the heterotrophic plate count (HPC) has been used historically to determine the utility of newly developed non-culture methods, and can be an appropriate reference method in many cases (3), there are cases for which HPC is not an appropriate refer method

6.2.1 The choice of referee method to use for validating a new or proposed non-culture method should be determined based on the parameter the new method purports to be measuring.

6.2.2 Several methods used for the HPC are listed in Table 1.

6.2.3 When none of the Table 1 variations of the HPC (Heterotrophic Plate Count) are suitable refereed methods, Adenosine Triphosphate Concentration (Test Method D4012) or the Most Probable Number (MPN) technique (7) may be more appropriate.

6.2.4 Alternative standard enumeration methods or methods for measuring the rate of the appearance of derivatives or the rate of disappearance of components of the product in which the microbial contamination is being measured—where such

TABLE 1 Comparison of Selected Heterotrophic Plate Count Procedures for Samples from Various Sources

	Water (4)	Dairy (5)	Environment (6)	Food (7)	Cosmetic (7)	Paper (8)	Pharmaceutical (9)
Media	TGE, SM, R2A or m-HPC	SM	SM or TGE	SM	ML	TGE	SCD
Dilution, H ₂ O	KH ₂ PO ₄ + MgCl ₂	KH ₂ PO ₄	KH ₂ PO ₄	KH ₂ PO ₄	MLB	H ₂ O	KH ₂ PO ₄
Incubation, °C	35 ± 0.5 20 or 28 (R2A)	32 ± 1	35 ± 0.5	35	30 ± 2	36 ± 0.5	30–35
Incubation, h	48 ± 3 72 ± 4 (bottled water) 72–168 (R2A medium)	48 ± 3	48	48 ± 2	48	48	48–72
Amount of Agar, mL	10–12 (Pour Plate) 15 (Spread Plates) 5 (Membrane Filter)	10–12	10+	12–15	Spread Plates	15–20	15–20

TGE = Tryptone Glucose Extract Agar
SM = Standard Methods Agar (Tryptone Glucose Yeast Agar)
ML = Modified Lethen Agar
MLB = Modified Lethen Broth
SCD = Soybean Casein Digest Agar
R2A = Low-Nutrient Media (which may not be available in dehydrated form)
m-HPC = Formerly called m-SPC Agar (used for membrane filtration)